

Lacewings and their Allies



Part 2

Introduction - Part 2

- Part 2 of the two part webinar series on British Isles Lacewing and Allies identification
- Identification using a microscope
- Some features will help you identify from photographs
- Take away – be aware of important identification features for different groups and species and where to find them on the insect



Coverage

NEUROPTERA (Lacewings)

CONIOPTERYGIDAE (Waxflies)

- *Conwentzia pineticola*
- *Conwentzia psociformis*
- *Coniopteryx borealis*
- *Coniopteryx tineiformis*
- *Coniopteryx pygmaea*
- *Coniopteryx esbenpeterseni*
- *Coniopteryx lentiae*
- *Semidalis aleyrodiformis*
- *Semidalis pseudouncinata*
- *Parasemidalis fuscipennis*
- *Aleuropteryx juniperi*
- *Helicoconis hirtinervis*

SISYRIDAE (Spongeflies)

- *Sisyra dalii*
- *Sisyra nigra*
- *Sisyra terminalis*

CHRYSOPIIDAE (Green Lacewings)

- *Chrysopa abbreviata*
- *Chrysopa commata*
- *Chrysopa dorsalis*
- *Chrysopa pallens*
- *Chrysopa perla*
- *Chrysopa phyllochoma*
- *Chrysoperla carnea*
- *Chrysoperla lucasina*
- *Chrysoperla pallida*
- *Chrysopidia ciliata*
- *Cunctochrysa albolineata*
- *Cunctochrysa cosmia*
- *Apertochrysa flavifrons*
- *Apertochrysa prasina*
- *Apertochrysa ventralis*
- *Nineta flava*
- *Nineta vittata*
- *Nineta inpunctata*
- *Nineta pallida*
- *Nothochrysa capitata*

- *Nothochrysa fulviceps*
- *Peyerimhoffina gracilis*

HEMEROBIIDAE (Brown Lacewings)

- *Psectra diptera*
- *Micromus variegatus*
- *Micromus angulatus*
- *Micromus paganus*
- *Hemerobius humulinus*
- *Hemerobius perelegans*
- *Hemerobius simulans*
- *Hemerobius stigma*
- *Hemerobius atrifrons*
- *Hemerobius pini*
- *Hemerobius contumax*
- *Hemerobius striatus*
- *Hemerobius nitidulus*
- *Hemerobius micans*
- *Hemerobius lutescens*
- *Hemerobius marginatus*
- *Wesmaelius malladai*
- *Wesmaelius mortoni*
- *Wesmaelius ravus*
- *Wesmaelius balticus*
- *Wesmaelius nervosus*
- *Wesmaelius subnebulosus*
- *Wesmaelius concinnus*
- *Wesmaelius quadrifasciatus*
- *Sympherobius elegans*
- *Sympherobius pygmaeus*
- *Sympherobius pellucidus*
- *Sympherobius fuscescens*
- *Sympherobius klapaleki*
- *Megalomus hirtus*

MEGALOPTERA

SIALIDAE (Alderflies)

- *Sialis fuliginosa*
- *Sialis lutaria*
- *Sialis nigripes*

MECOPTERA

PANORPIDAE (Scorpionflies) females

- *Panorpa cognata*
- *Panorpa communis*
- *Panorpa germanica*

Covered in Part 1:

RAPHIDIOPTERA (Snakeflies)

RAPHIDIIDAE

- *Subilla confinis*
- *Xanthostigma xanthostigma*
- *Atlantoraphidia maculicollis*
- *Phaeostigma notata*

NEUROPTERA (Lacewings)

OSMYLIDAE (Giant Lacewings)

- *Osmylus fulvicephalus*

MYRMELEONTIDAE (Antlions)

- *Euroleon nostras*
- *Myrmeleon formicarius*

HEMEROBIIDAE (Brown Lacewings)

- *Drepanepteryx phalaenoides*

MECOPTERA

PANORPIDAE (Scorpionflies) Males

- *Panorpa cognata*
- *Panorpa communis*
- *Panorpa germanica*

BOREIDAE (Snow Fleas)

- *Boreus hyemalis*

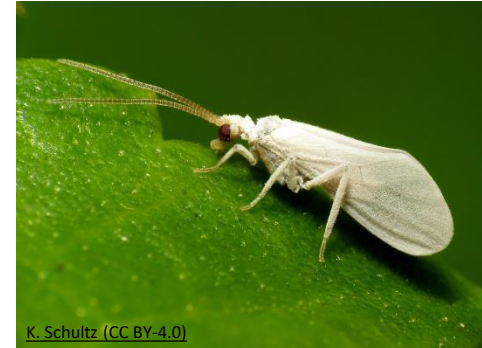
Preparation

- Can be pinned
- Preserved in alcohol
80% ethanol or isopropyl alcohol
- Label with important info.
- Most can be identified without much preparation
- Some require their abdomen clearing
- Some require dissection



Clearing abdomen

- 10% Potassium hydroxide
- Detach abdomen – with coniopterygids clear whole insect
- Put in test tube (5mm deep)
- Put in bowl/pan with boiling water (from kettle) leave 30 mins
- Or leave in cold solution overnight
- Remove from solution and wash with water on shallow tray under microscope
- Be careful when using chemicals and always read instructions and warnings



CBG Photography Group, Centre for Biodiversity Genomics (CC BY)

Megaloptera (Alderflies)

- Can be preserved pinned or in alcohol
- Wing venation and general morphology similar between all species
- Can only be identified from genitalia, both male and female



O. Fogh Nielsen (CC BY 4.0)

Sialis lutaria



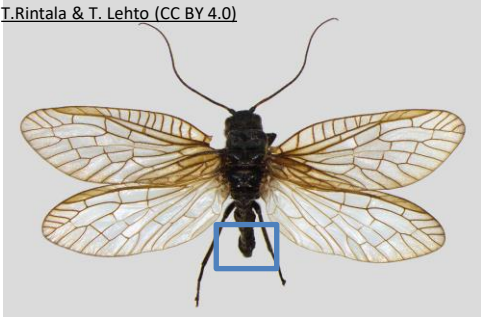
O. Fogh Nielsen (CC BY 4.0)

Sialis fuliginosa



O. Fogh Nielsen (CC BY 4.0)

Sialis nigripes



Side



Underneath

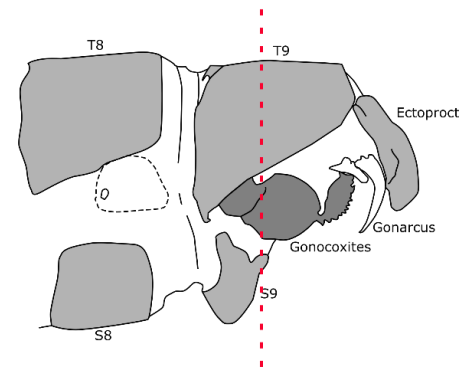
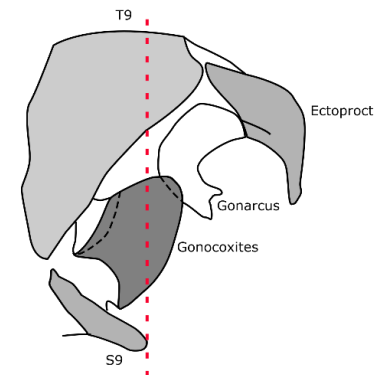
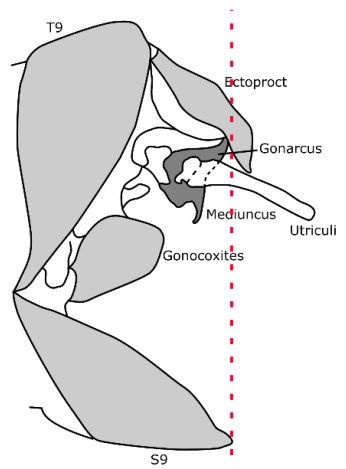


Sialis lutaria

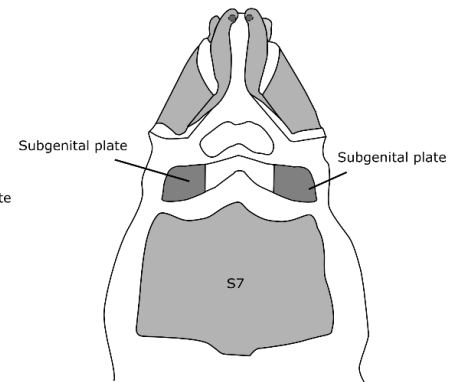
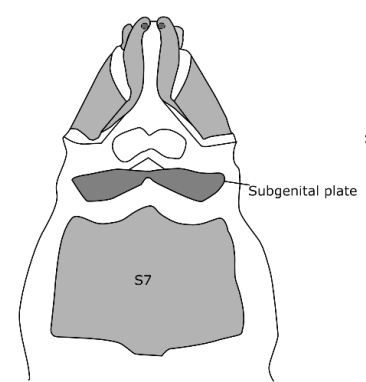
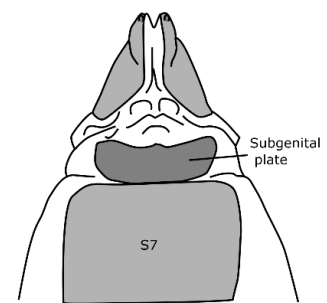
Sialis fuliginosa

Sialis nigripes

MALE



FEMALE



Sialis lutaria

Sialis fuliginosa

Sialis nigripes

Neuroptera (Lacewings)

- Can be preserved pinned or in alcohol
- Important diagnostic features from body morphology, wing venation, genitalia

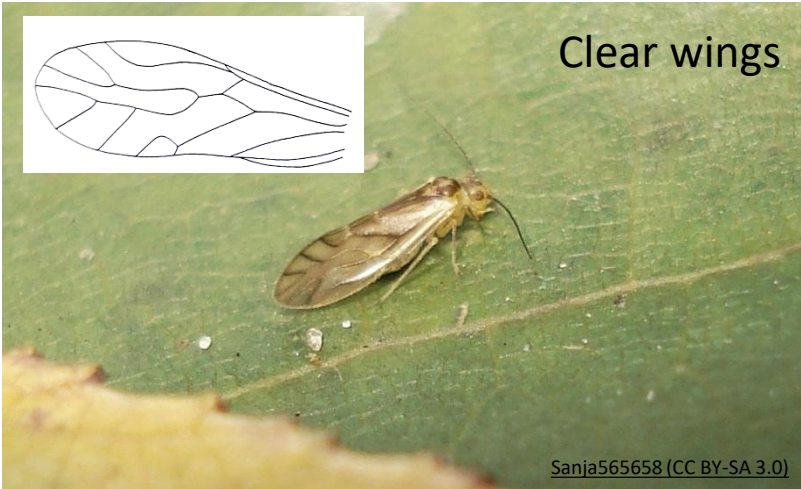


Coniopterygidae (Wax Flies)

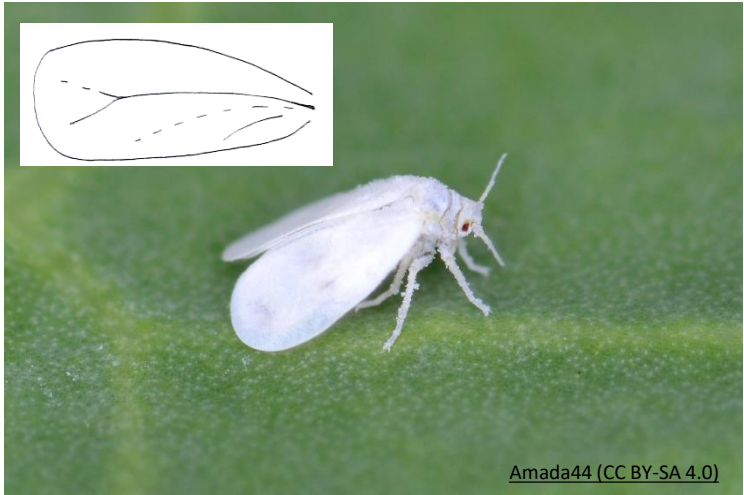
- Smallest neuropterans
- Best to preserve in alcohol
- Male specimens need to be collected for Identification
- Male genitalia diagnostic
- Many specimens need to have abdomen cleared



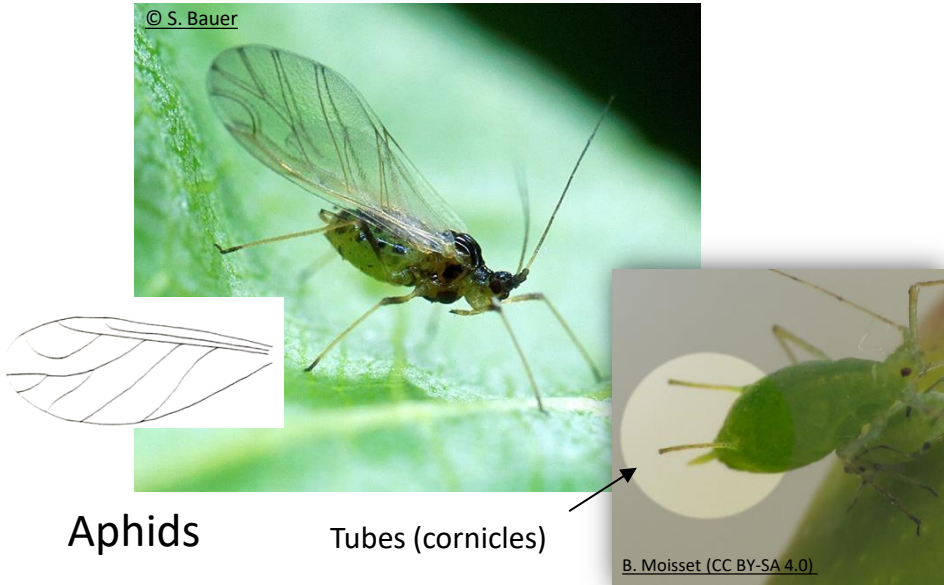
Confusion



Psocoptera

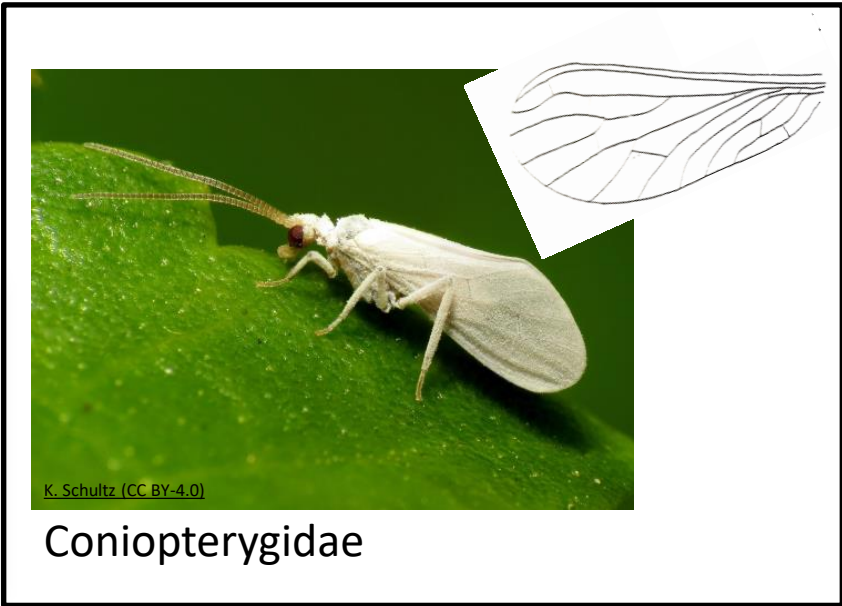


Whitefly (Hemiptera: Aleyrodidae)



Aphids

Tubes (cornicles)

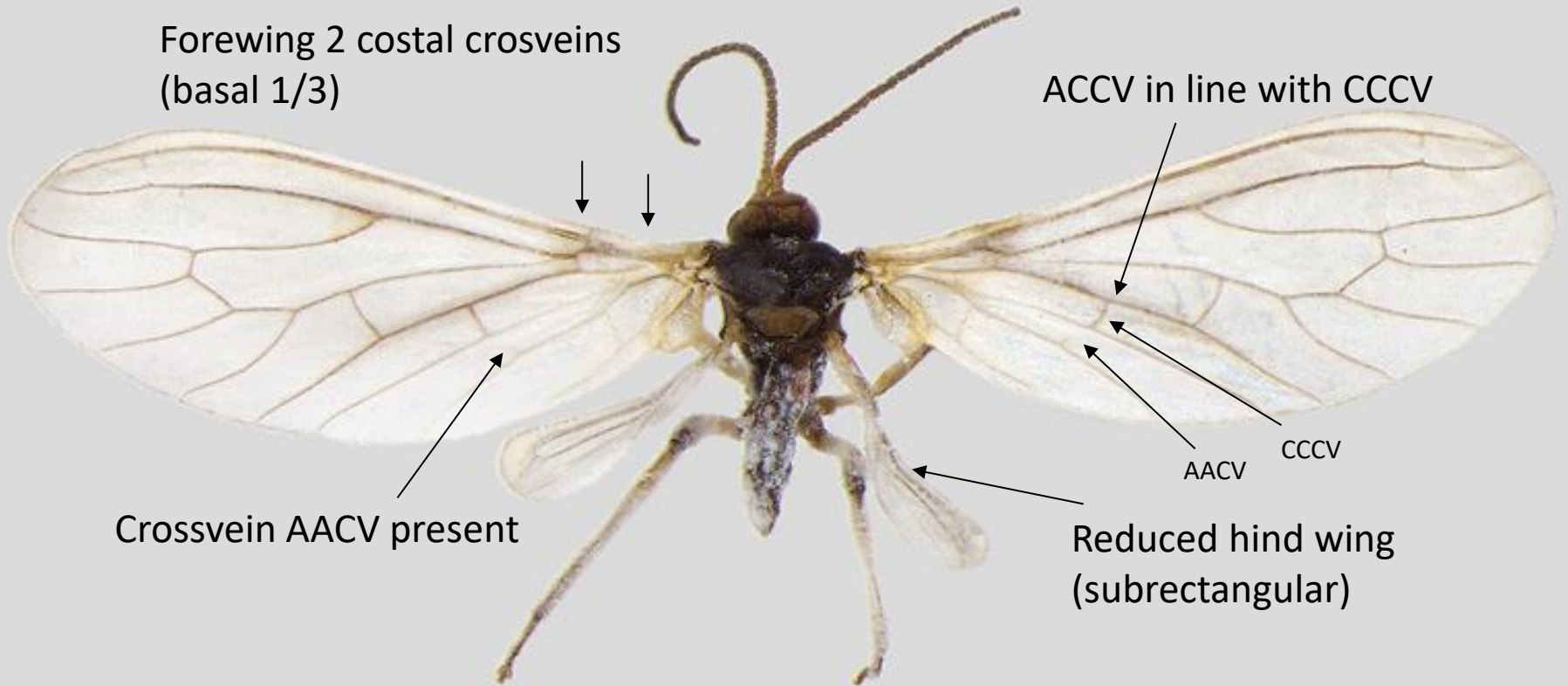


Coniopterygidae

Conwentzia

Forewing 2 costal crossveins
(basal 1/3)

ACCV in line with CCCV



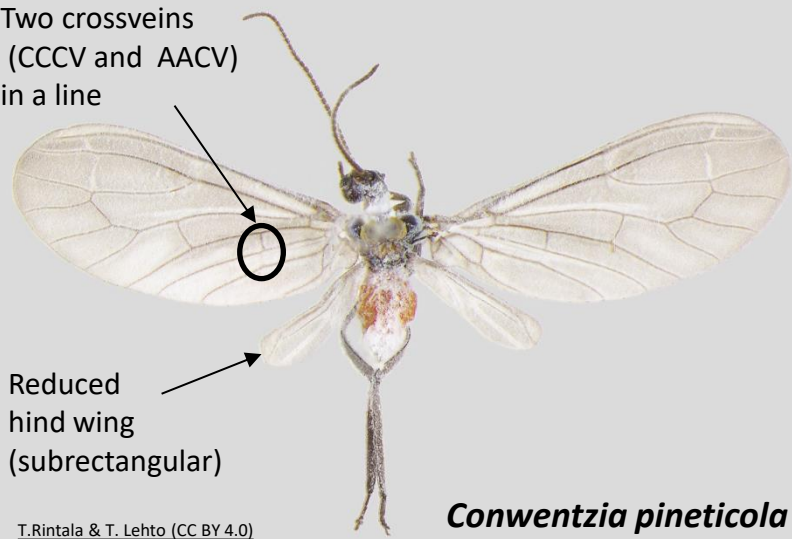
Crossvein AACV present

AACV CCCV

Reduced hind wing
(subrectangular)

Conwentzia species

Two crossveins
(CCCV and AACV)
in a line

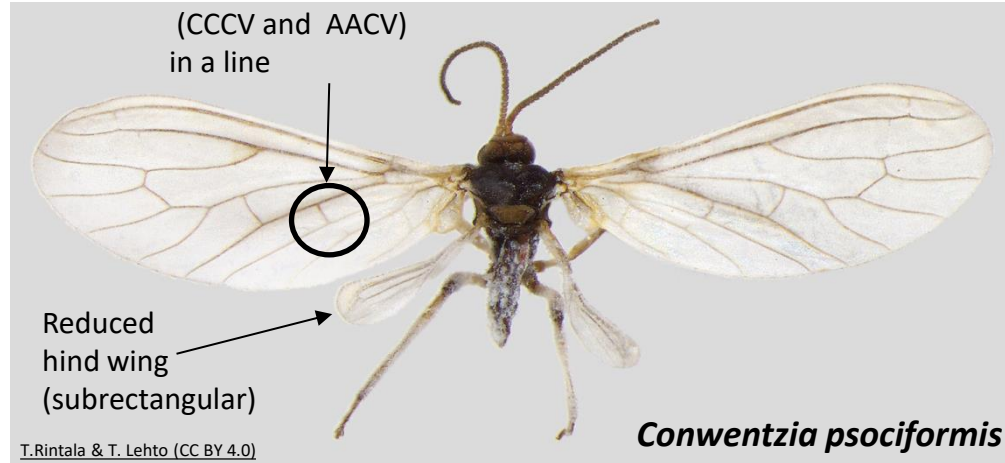


Reduced
hind wing
(subrectangular)

T.Rintala & T. Lehto (CC BY 4.0)

Conwentzia pineticola

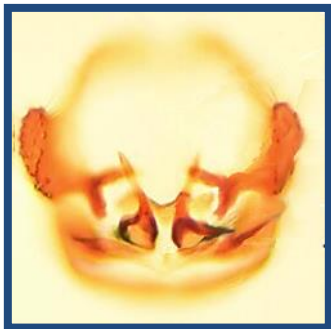
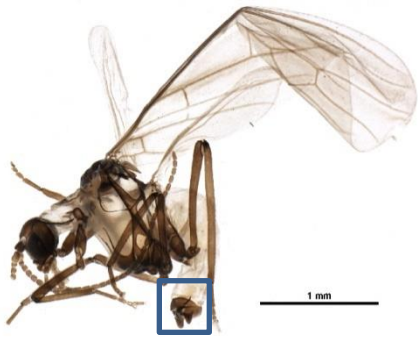
Two crossveins
(CCCV and AACV)
in a line



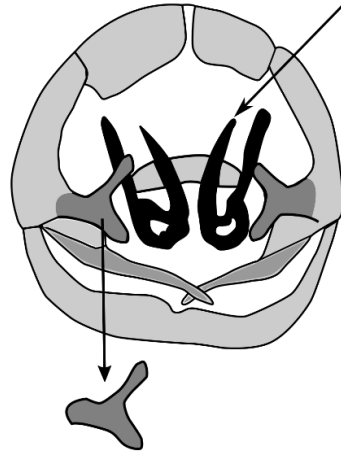
Reduced
hind wing
(subrectangular)

T.Rintala & T. Lehto (CC BY 4.0)

Conwentzia psociformis



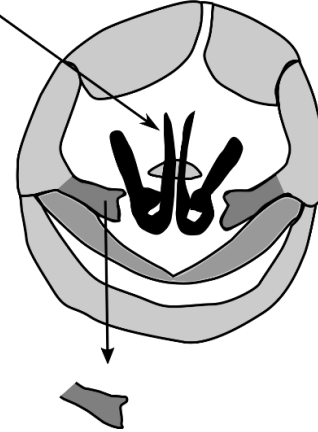
diverging wideley



process of ectoproct forked
Conwentzia pineticola

Parameres

parallel diverging only slightly



process of ectoproct not as forked
Conwentzia psociformis

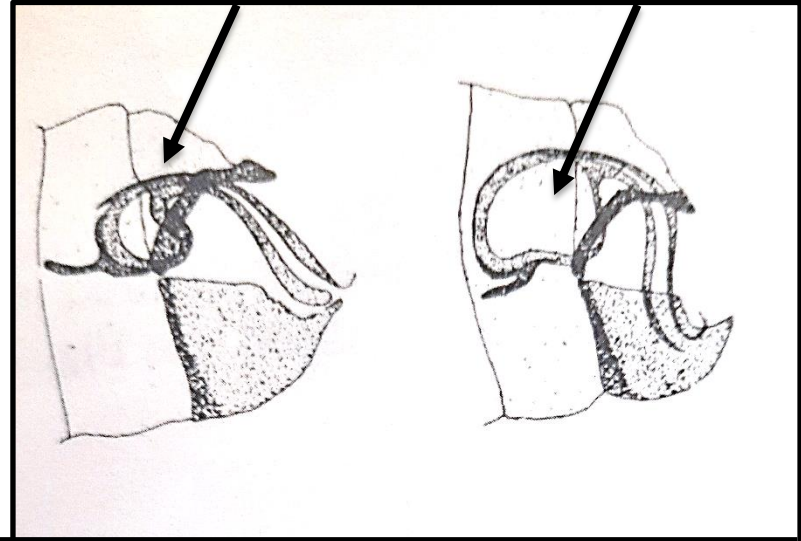
Coniopteryx



Subgenera: *Metaconiapteryx* and *Coniapteryx*

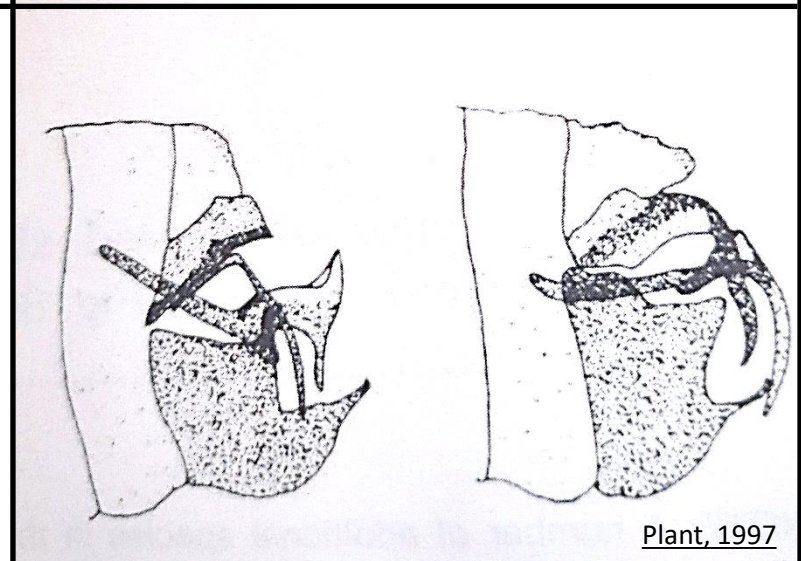
Metaconiapteryx

Internal genital structures in a ring

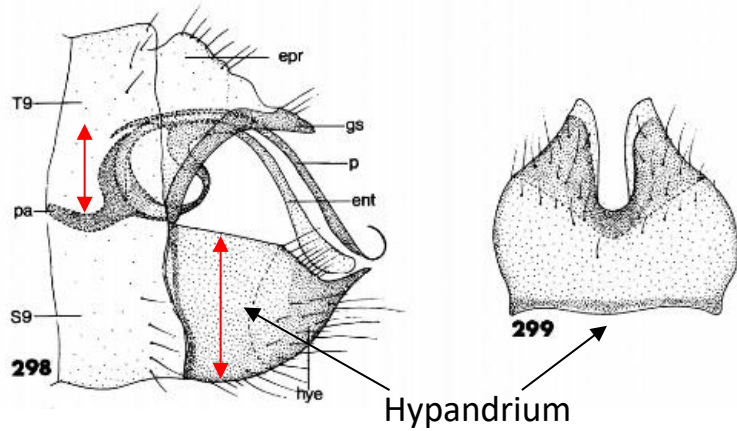


Coniapteryx

Internal genital structures **NOT** in a ring



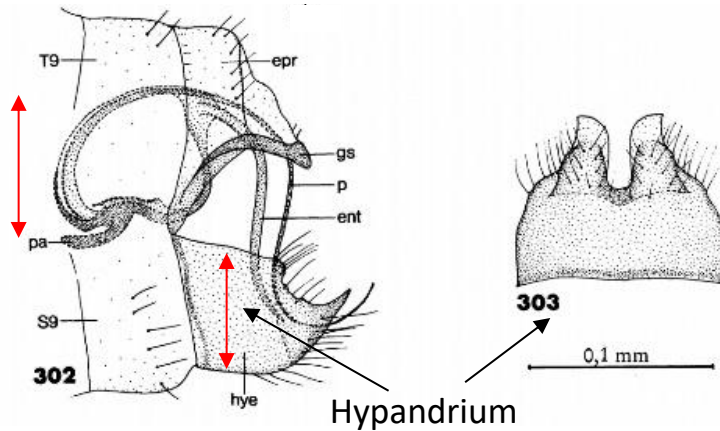
Coniopteryx (Metaconiopteryx)



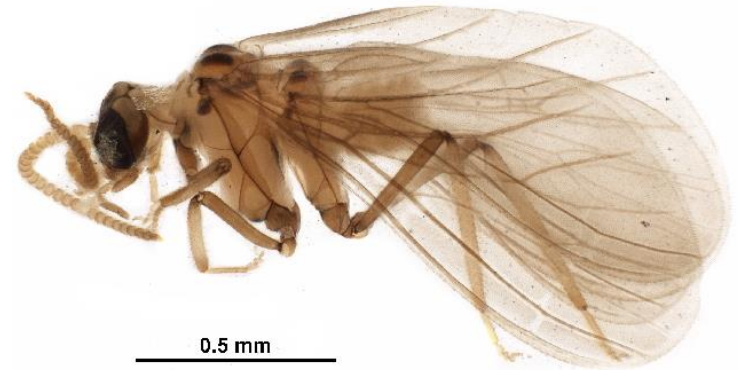
Coniopteryx (Metaconiopteryx) esbenpeterseni



SNSB, Zoologische Staatssammlung Muenchen (CC BY-NC-SA)

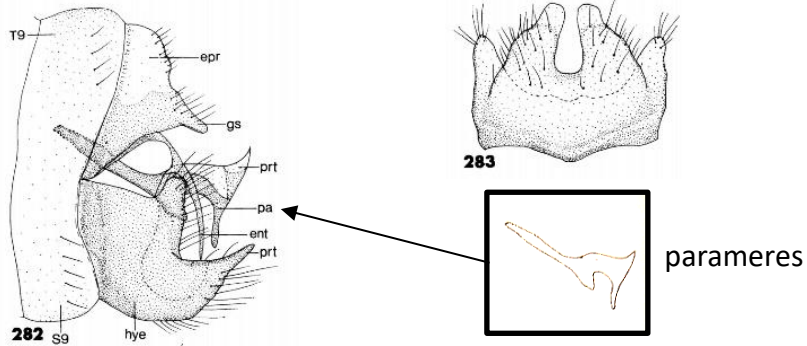


Coniopteryx (Metaconiopteryx) lentiae

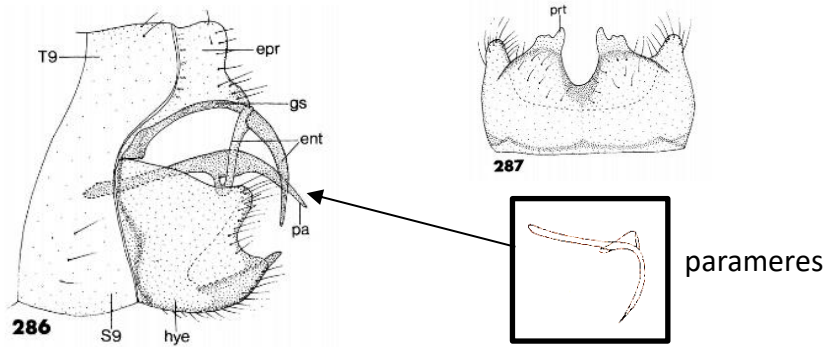
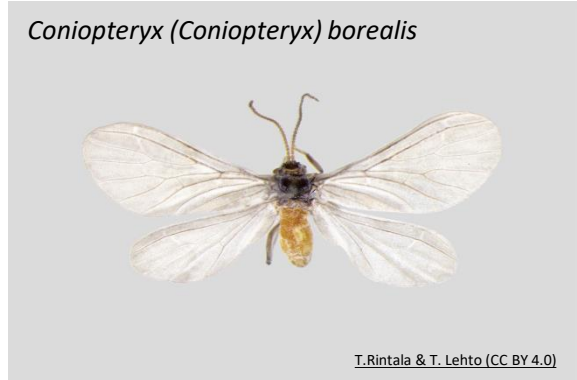


Aletardi (CC-BY)

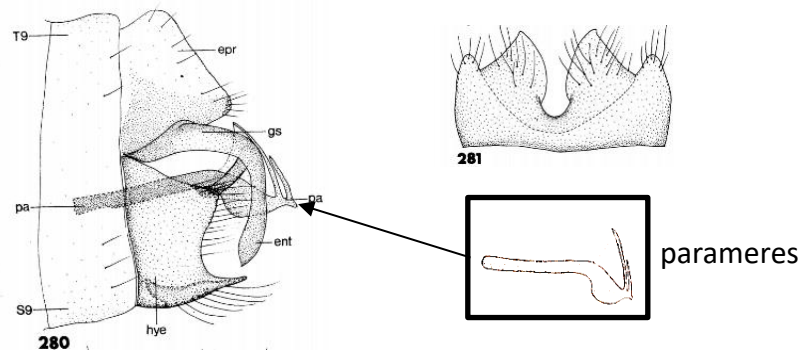
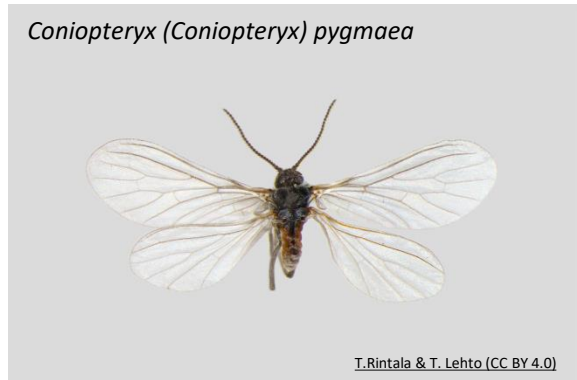
Coniopteryx (Coniopteryx)



Coniopteryx (Coniopteryx) borealis



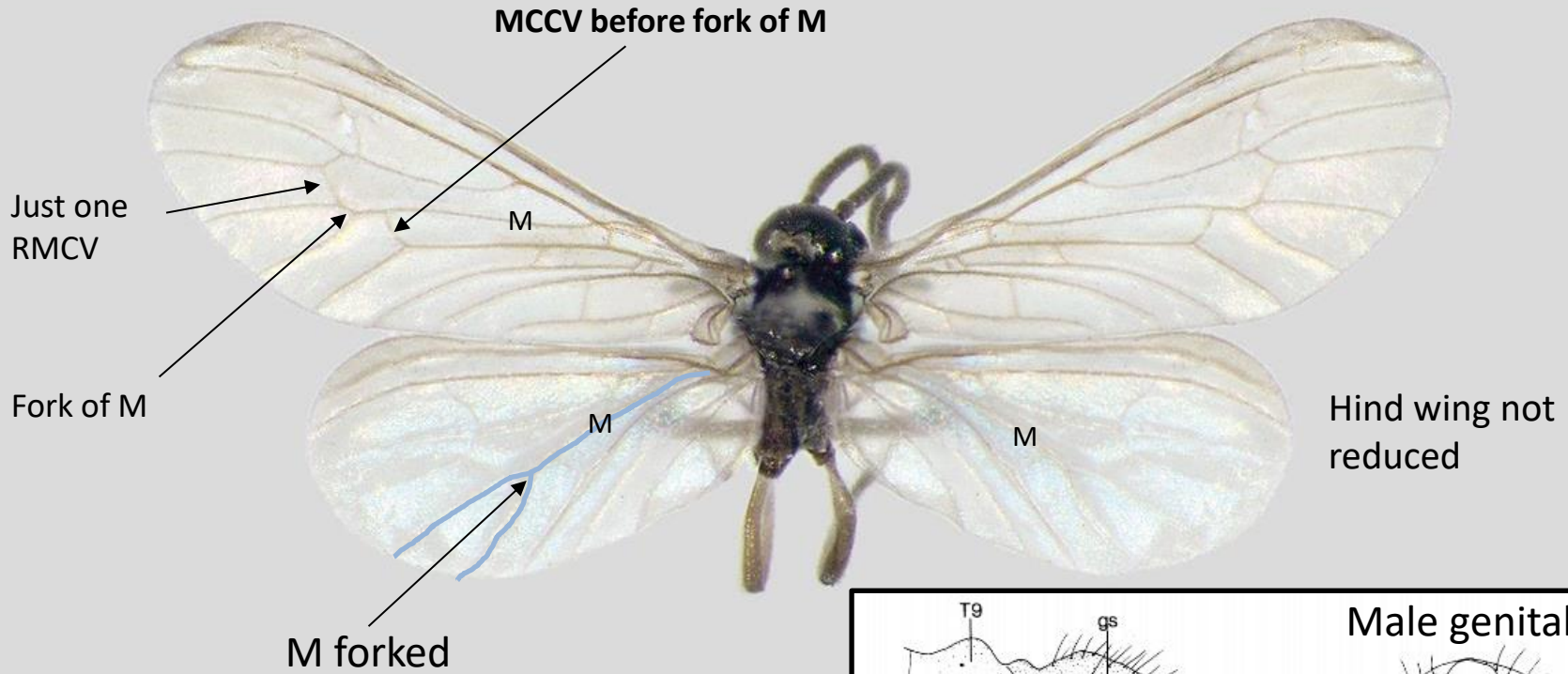
Coniopteryx (Coniopteryx) pygmaea



Coniopteryx (Coniopteryx) tineiformis

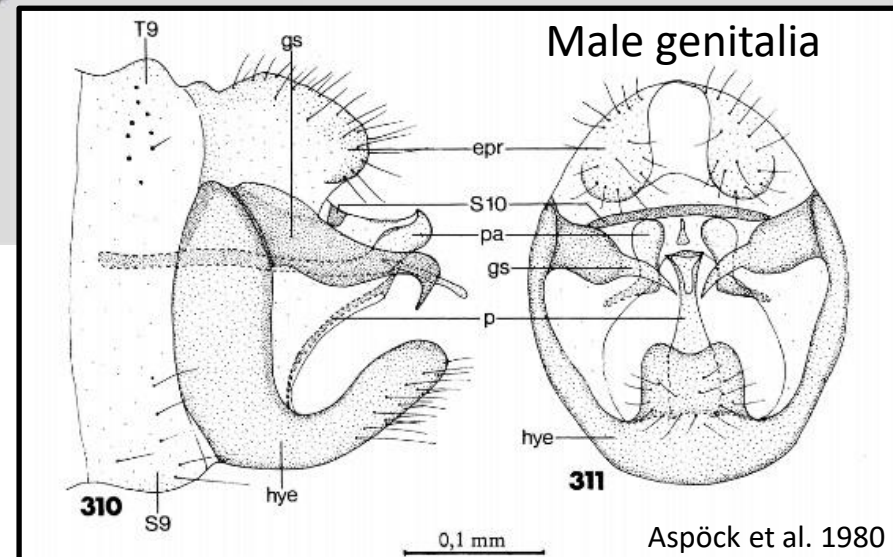


Parasemidalis fuscipennis

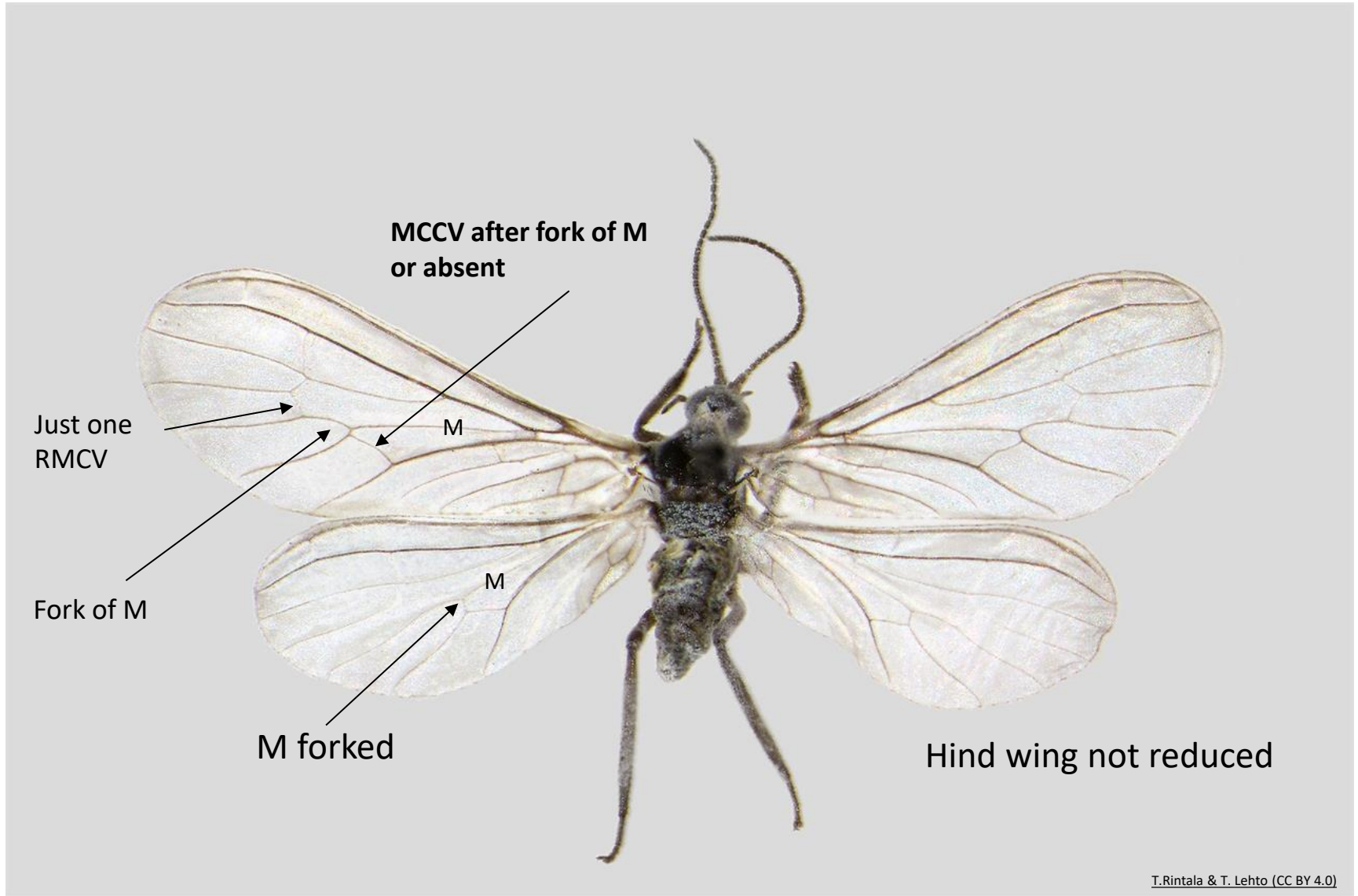


T.Rintala & T. Lehto (CC BY 4.0)

Only coniopterygid in British Isles
that can be identified by wing venation



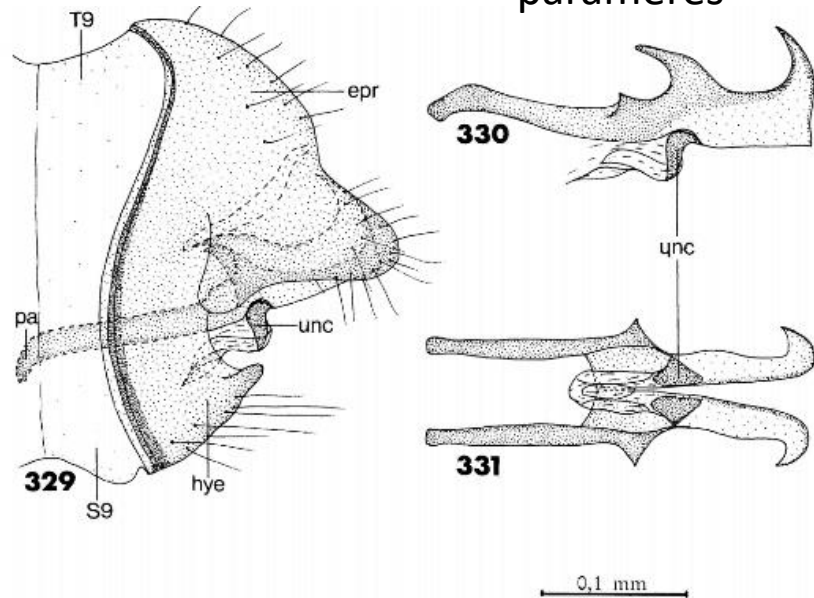
Semidalis



Male genitalia

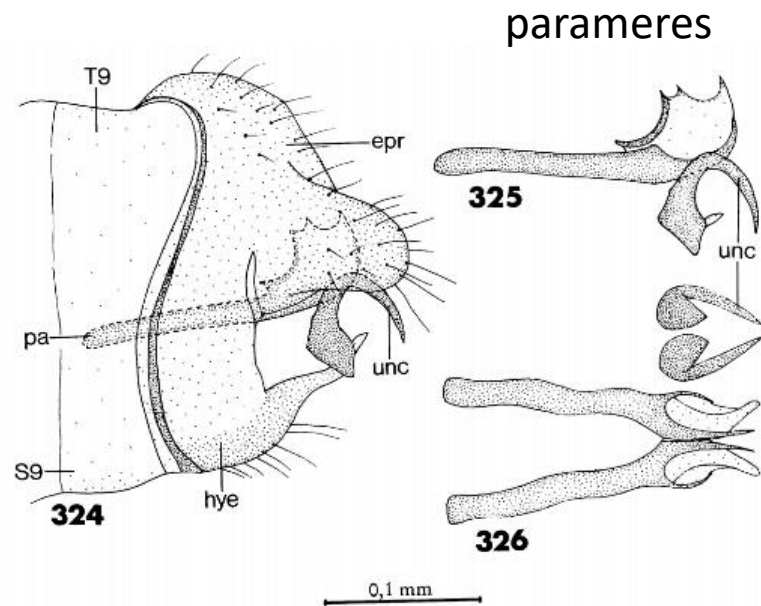


Semidalis aleyrodiformis

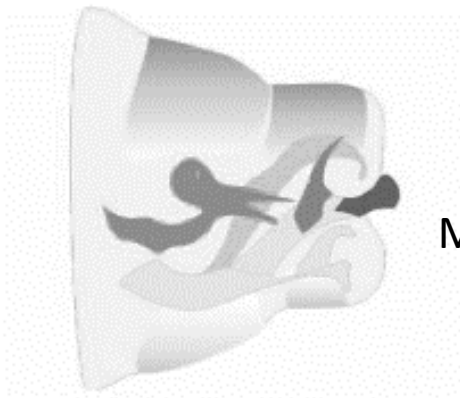
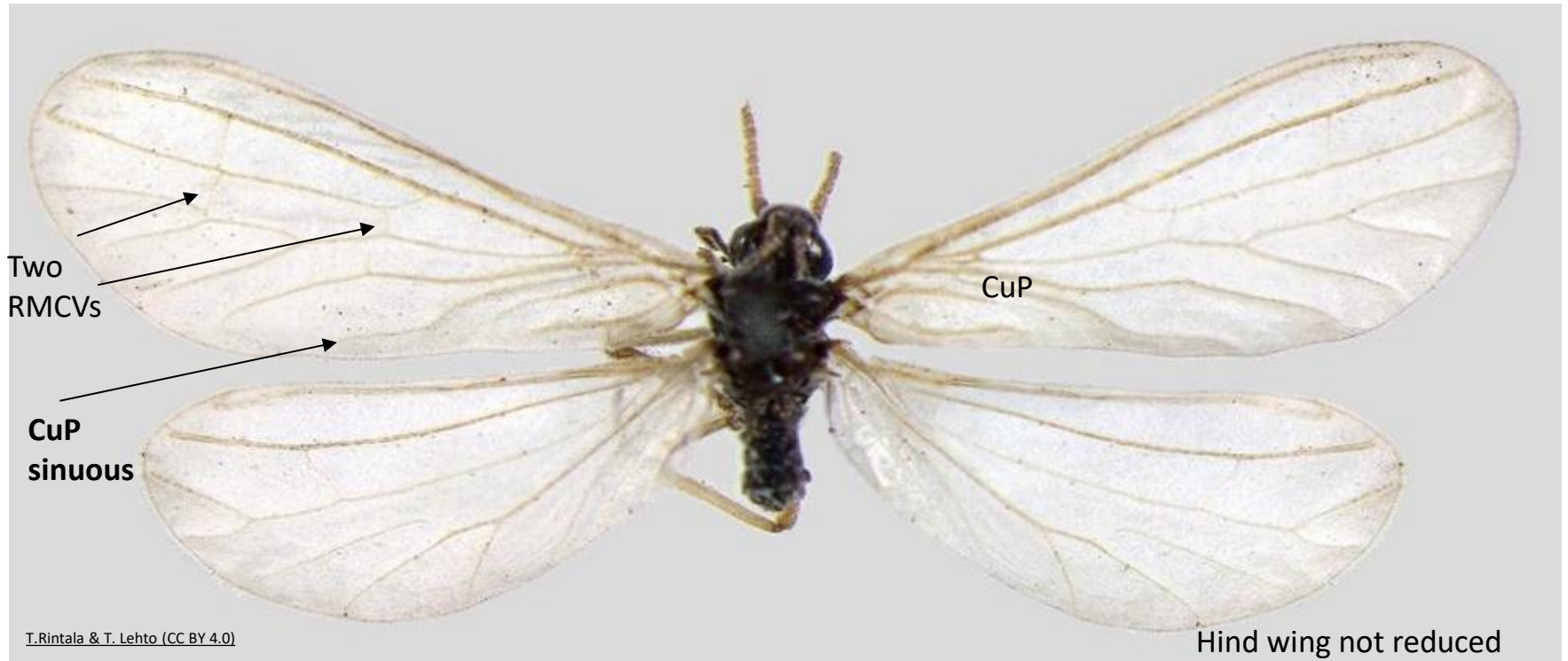


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Semidalis pseudouncinata



Aleuropteryx juniperi



Male genitalia

Need to check male genitalia as *Aleuropteryx loewii* is widespread in Europe and could be found in Britain

Helicoconis hirtinervis

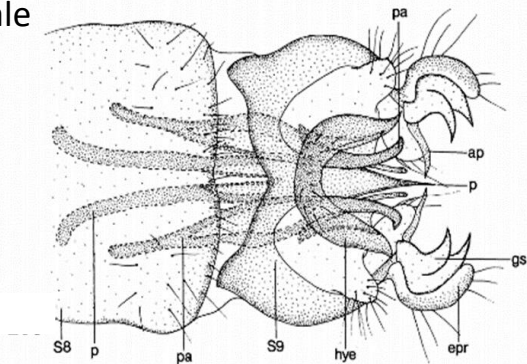
T.Rintala & T. Lehto (CC BY 4.0)

Male

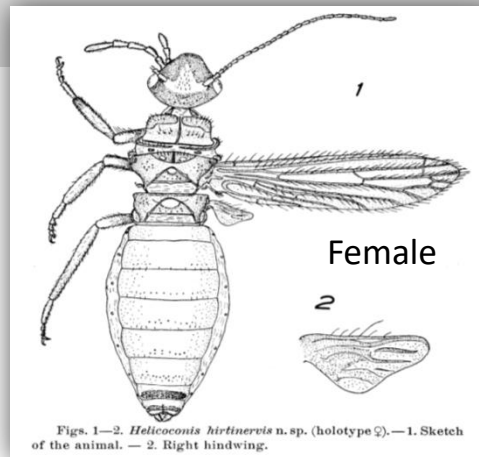


Hind wing not reduced in male

Male



Aspöck et al. 1980



Tjeder, 1960

Female fore- and hind wing reduced

Hemerobiidae (Brown Lacewings)

- Can be pinned or preserved in alcohol
- May need to dissect or clear the abdomen
- Genitalia often needs to be observed to confirm identity

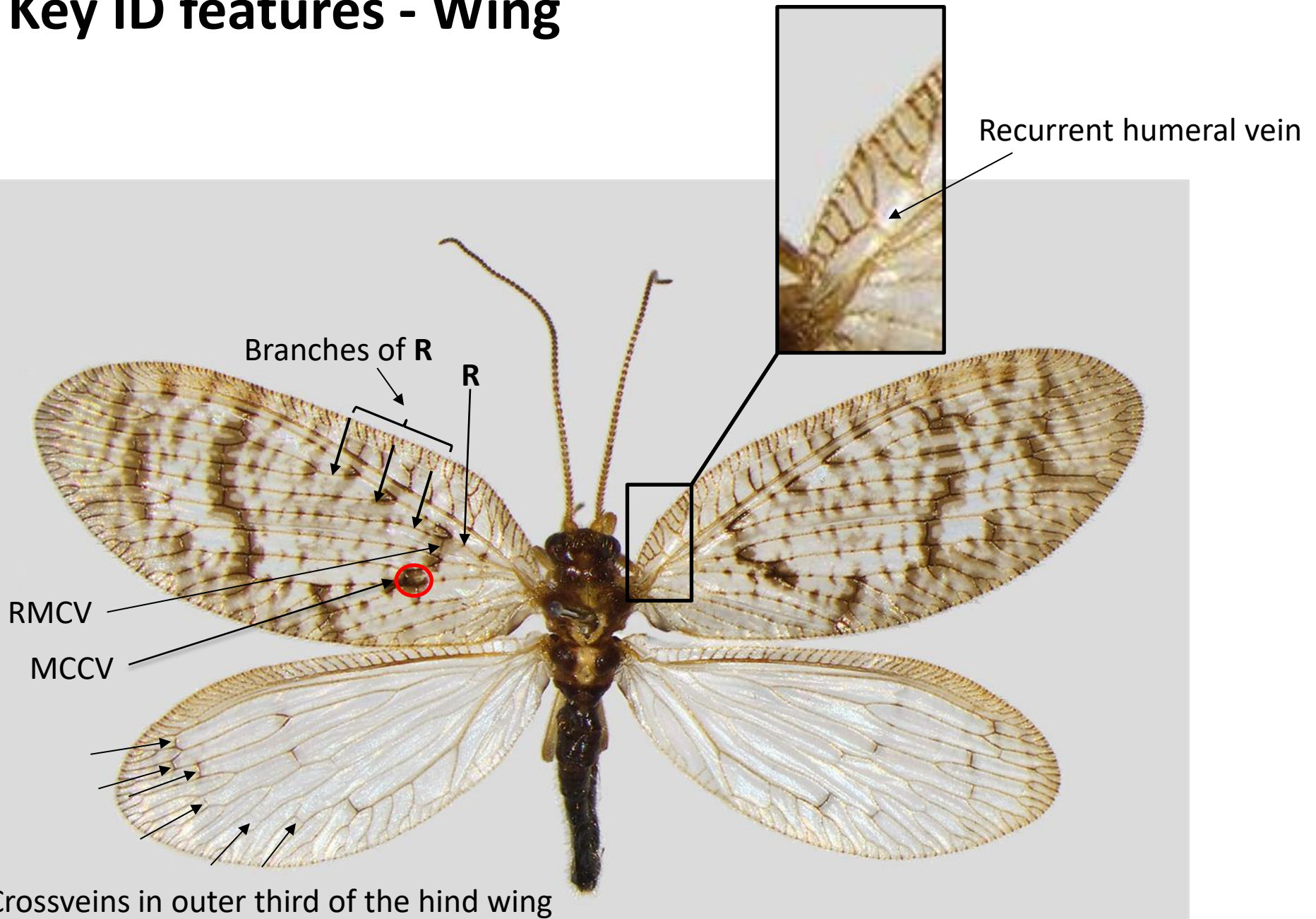


O. Fogh Nielsen (CC BY 4.0)



O. Fogh Nielsen (CC BY 4.0)

Key ID features - Wing



Recurrent humeral vein absent

T.Rintala & T. Lehto (CC BY 4.0)



T.Rintala & T. Lehto (CC BY 4.0)



Splits of two genera: *Psectra* and *Micromus*

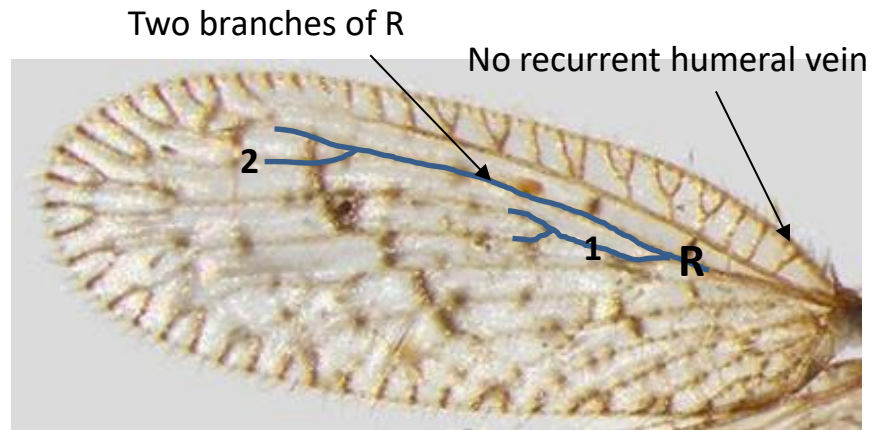
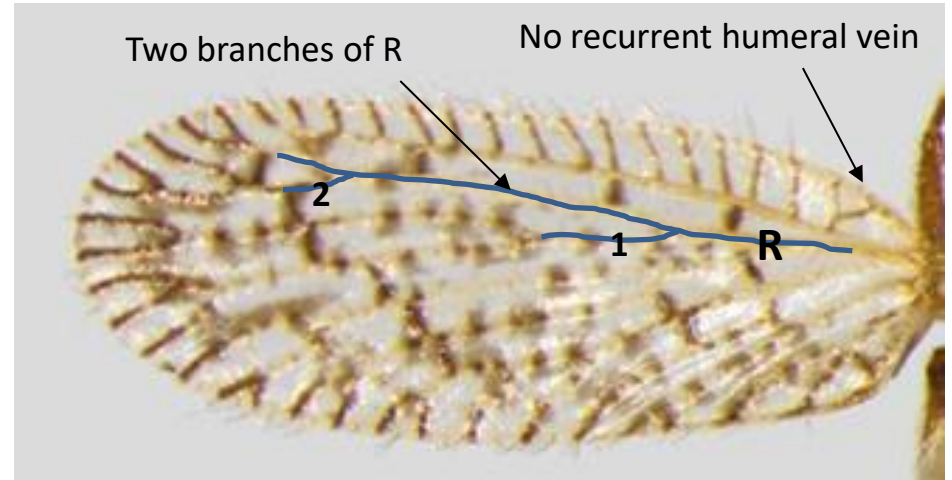
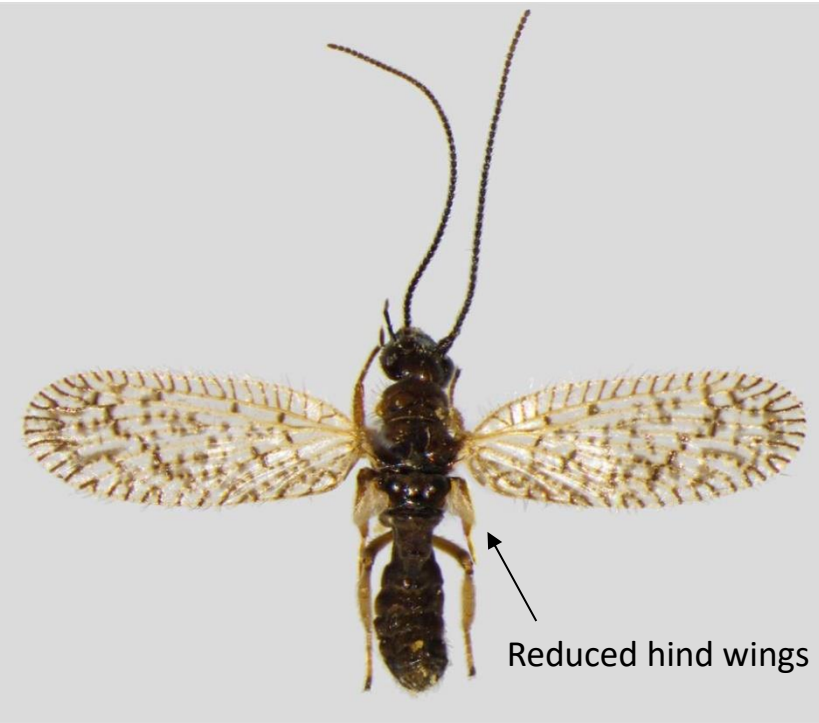


Recurrent humeral vein

First basal vein in costal area
Curves strongly backwards
and has numerous branches
coming off it

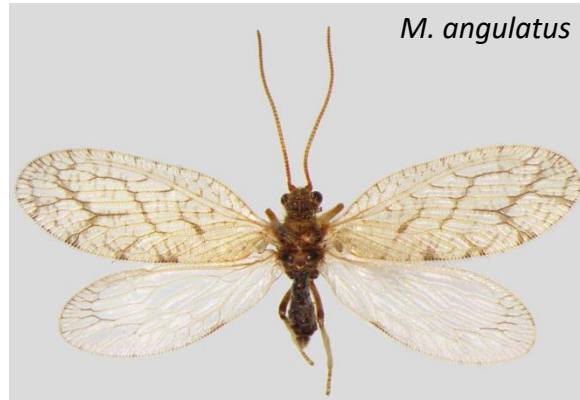
T.Rintala & T. Lehto (CC BY 4.0)

Psectra diptera



Hind wings not reduced

Micromus



(Two or) three branches R

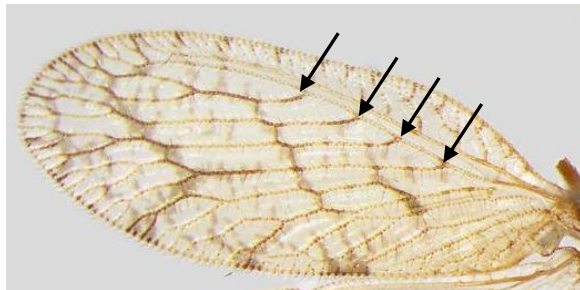
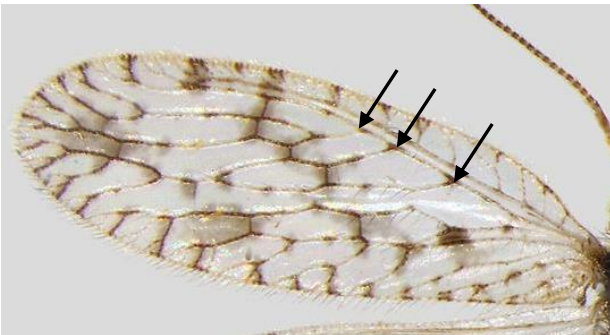


Four branches R

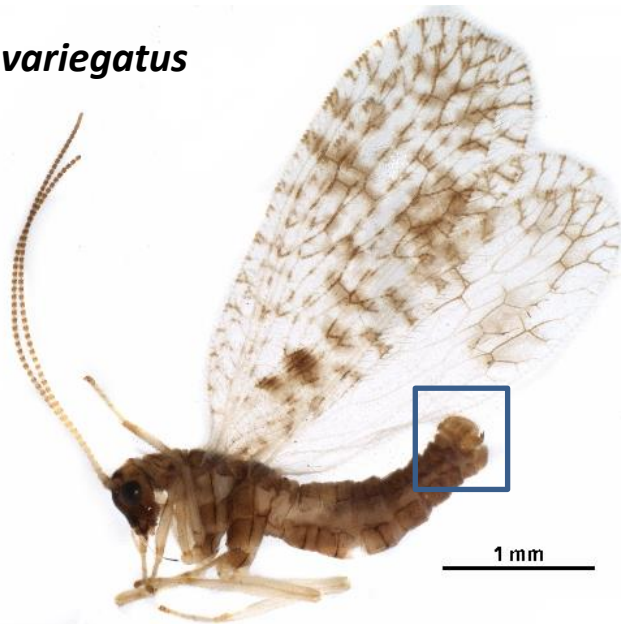


(Four or) five branches R

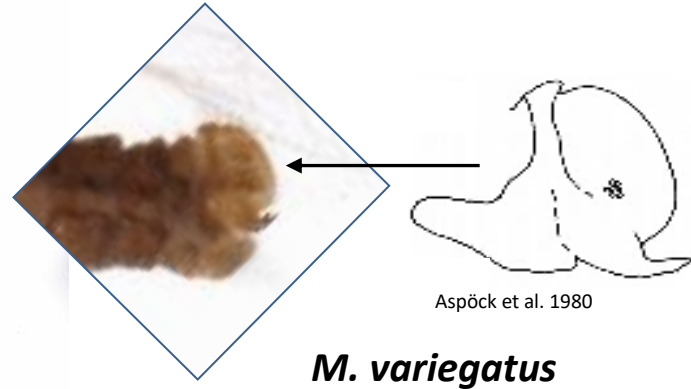
No recurrent humeral vein



Micromus variegatus

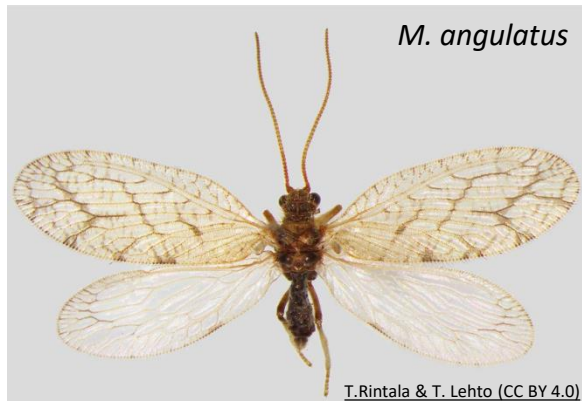


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Aspöck et al. 1980

M. variegatus



M. angulatus

T.Rintala & T. Lehto (CC BY 4.0)



Aspöck et al. 1980



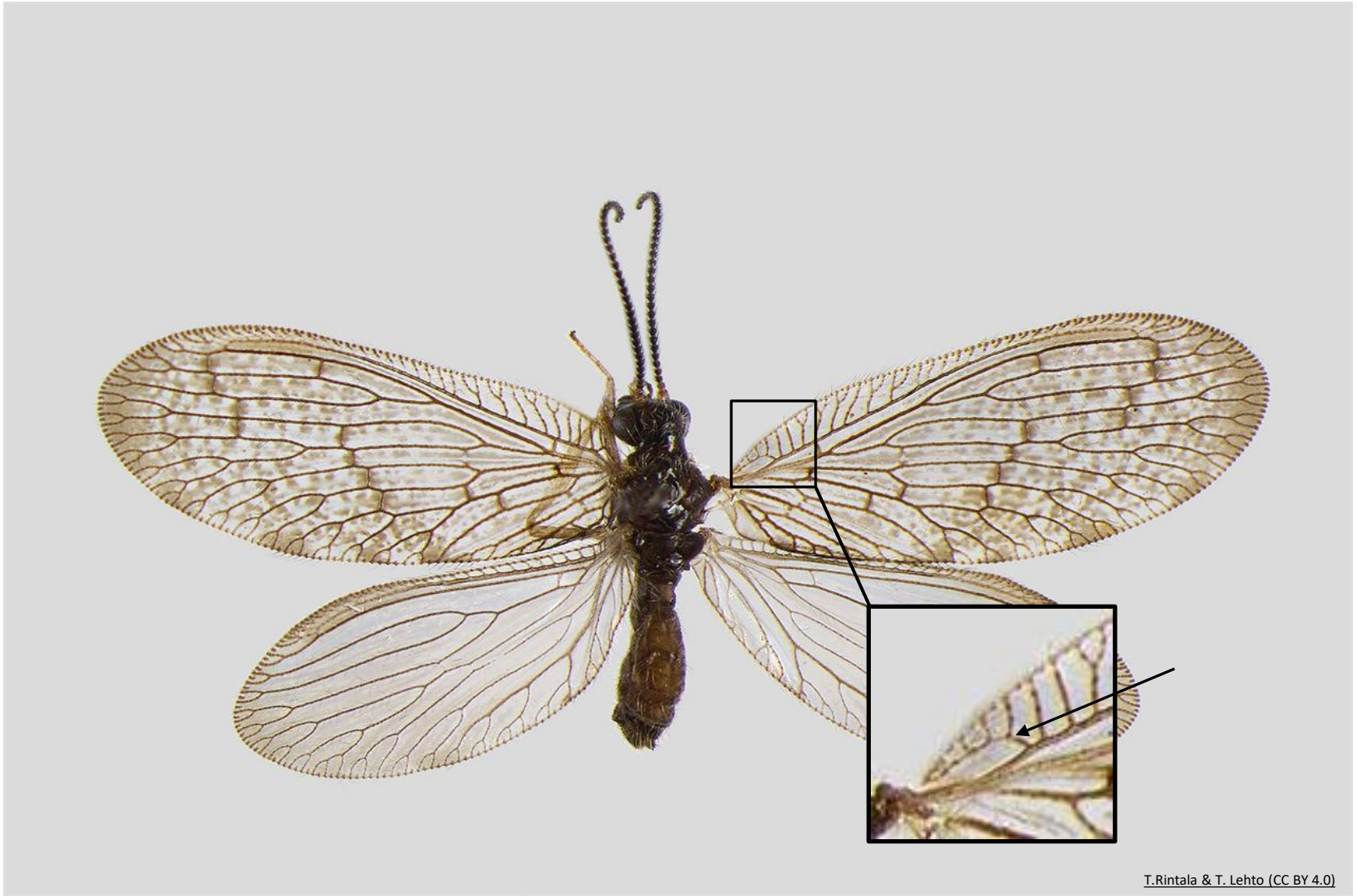
M. paganus

T.Rintala & T. Lehto (CC BY 4.0)



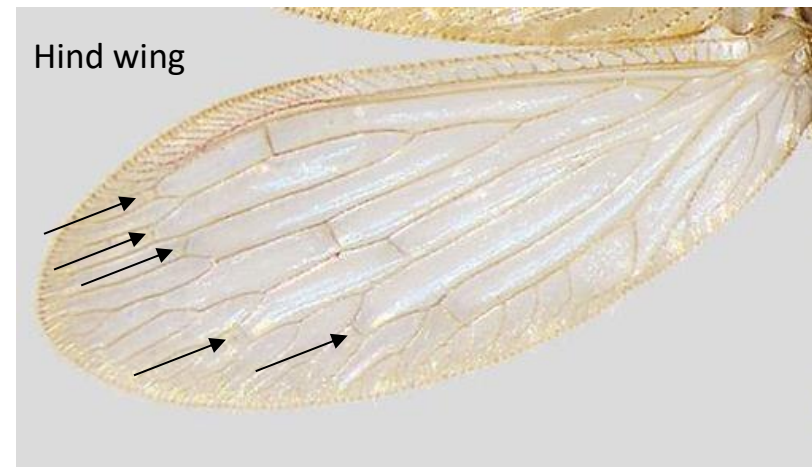
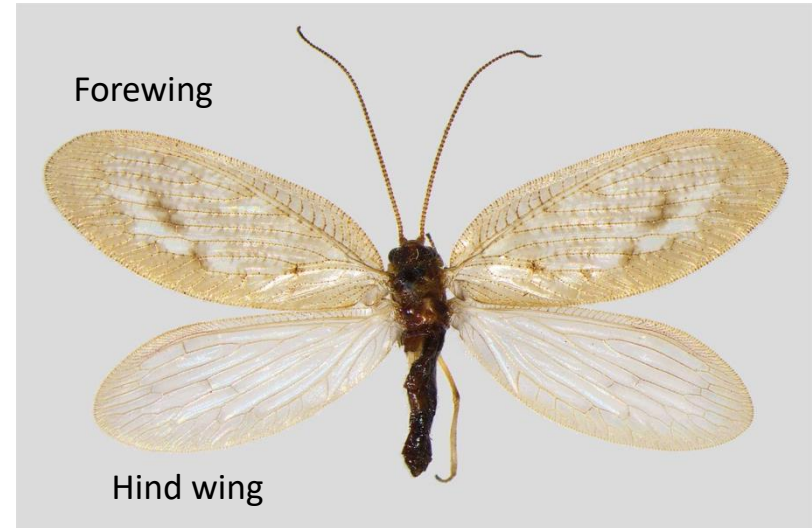
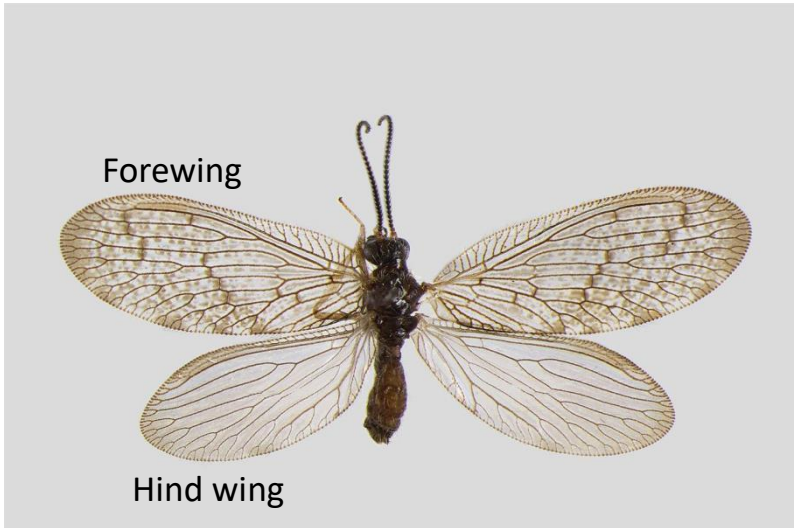
Aspöck et al. 1980

Recurrent humeral vein present



Symphorobius, Megalomus, Hemerobius, Wesmaelius

Hind wing crossveins



No more than 4 crossveins in outer third hind wing

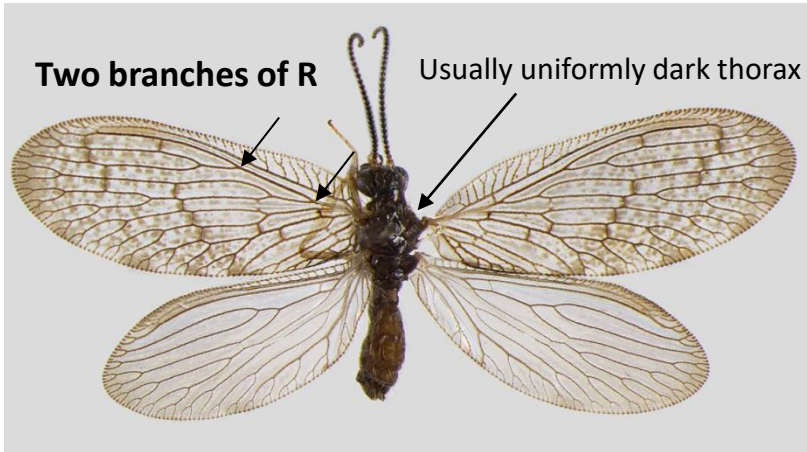
At least 5 crossveins in outer third hind wing

Sympherobius

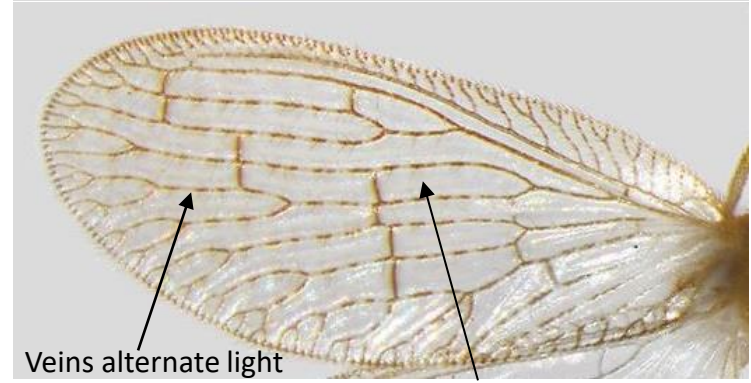
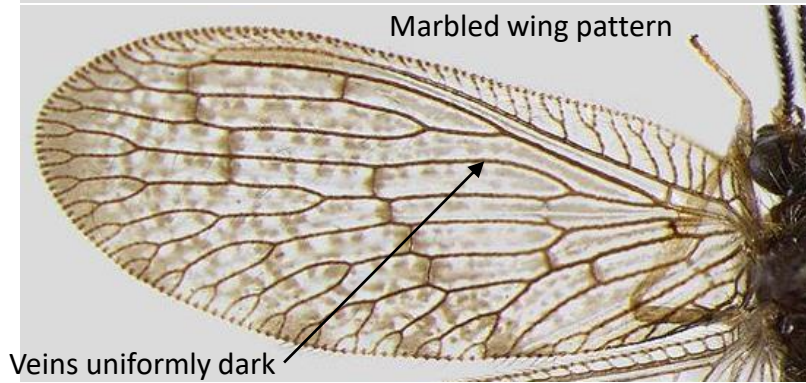
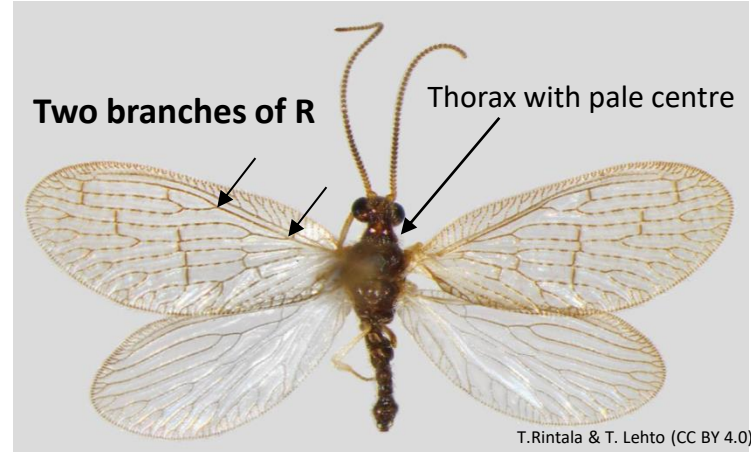
Megalonus, Hemerobius, Wesmaelius

Symphorobius

Symphorobius elegans



Symphorobius pygmaeus



Male genitalia

Aspöck et al. 1980

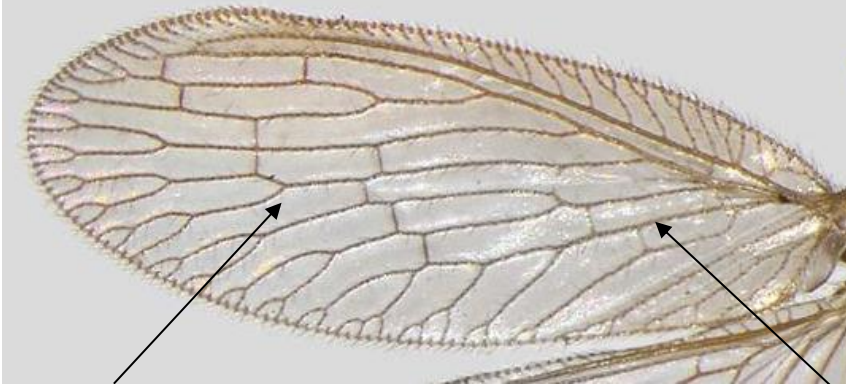
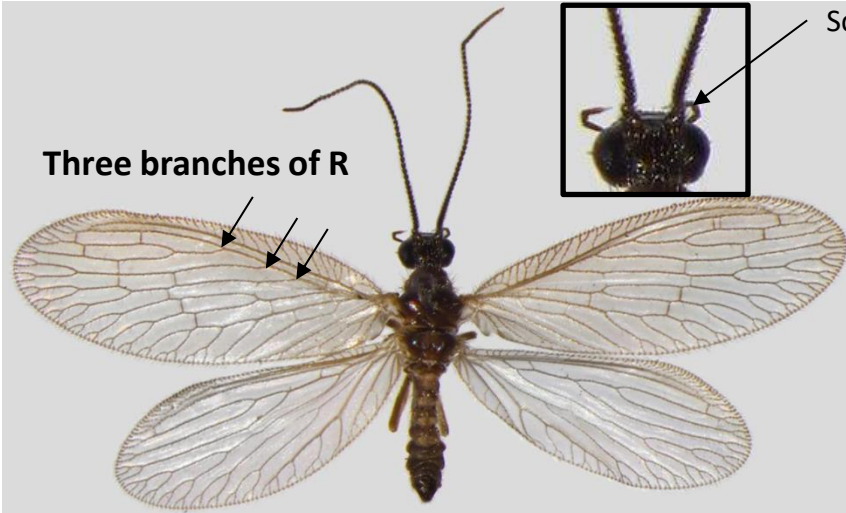


Male genitalia

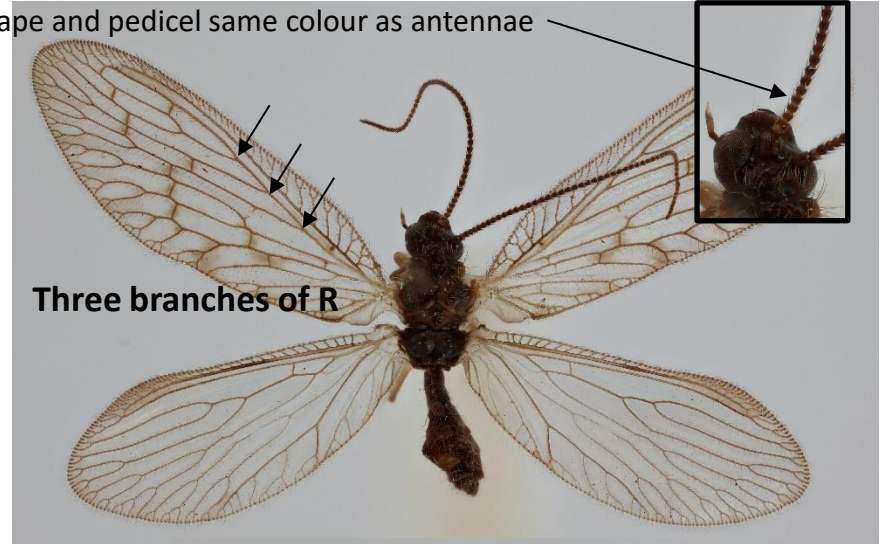
Aspöck et al. 1980

All photos: T.Rintala & T. Lehto (CC BY 4.0)

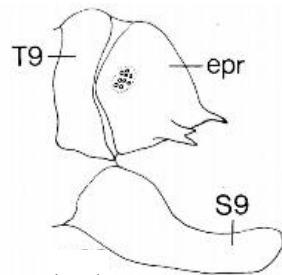
Symphorobius fuscescens



Symphorobius pellucidus



Male genitalia



Aspöck et al. 1980

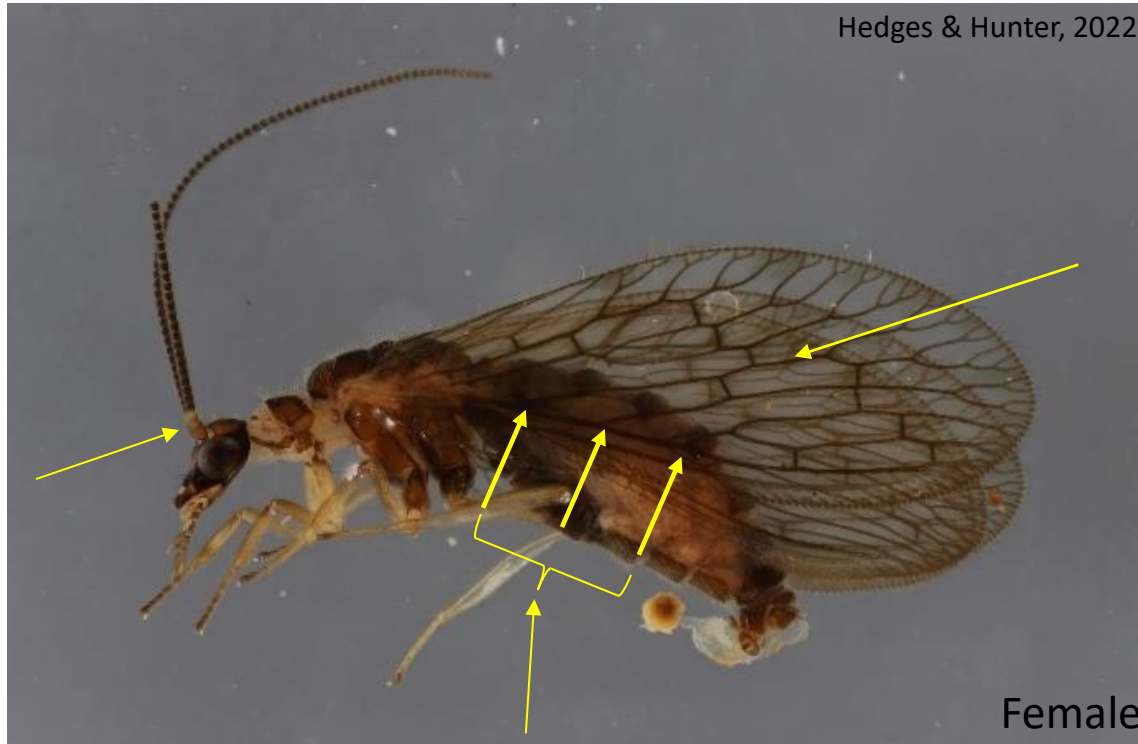
Male genitalia



Aspöck et al. 1980

Symphorobius klapaleki

Hedges & Hunter, 2022



Veins uniformly dark not alternating light and dark

Scape and pedicel pale yellow different to rest of antennae

3 branches of R

Female



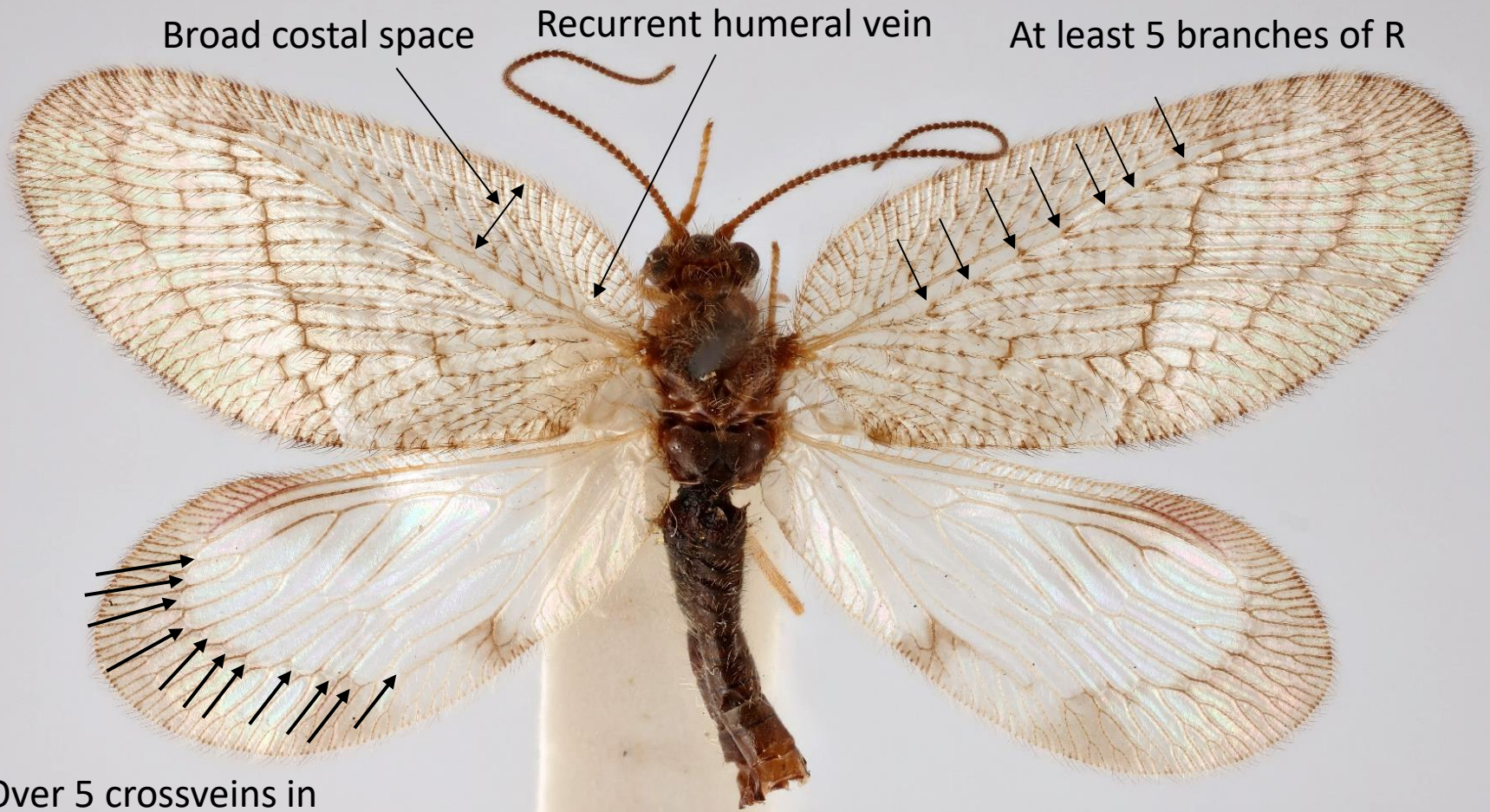
Male genitalia

Aspöck et al. 1980



At least 5 crossveins in outer third hind wing
Megalomus, Hemerobius, Wesmaelius

Megalomus hirtus



Broad costal space

Recurrent humeral vein

At least 5 branches of R

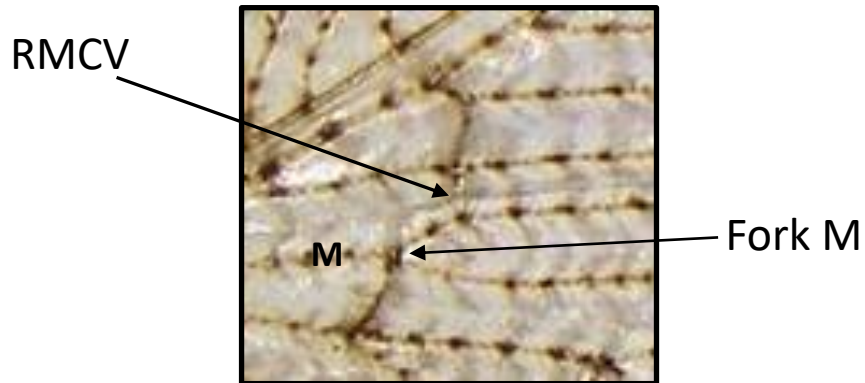
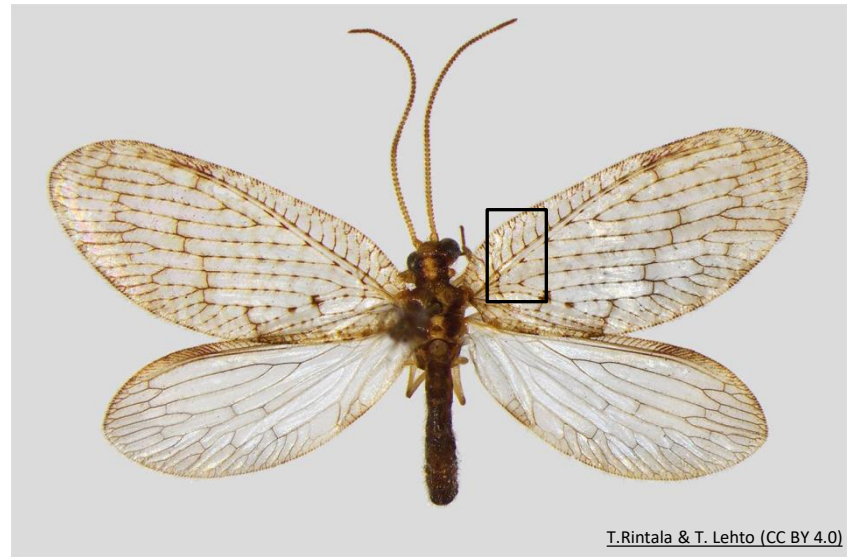
Over 5 crossveins in
Outer third of hind wing

RMCV and fork of M

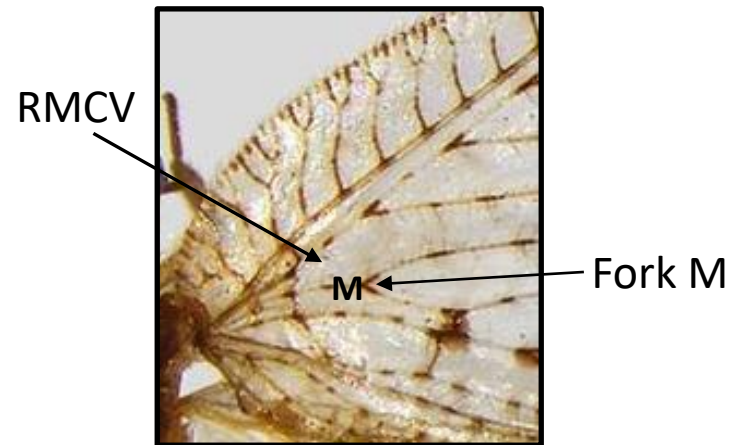
RMCV dark after fork of M



RMCV pale before (or at) fork of M



Wesmaelius



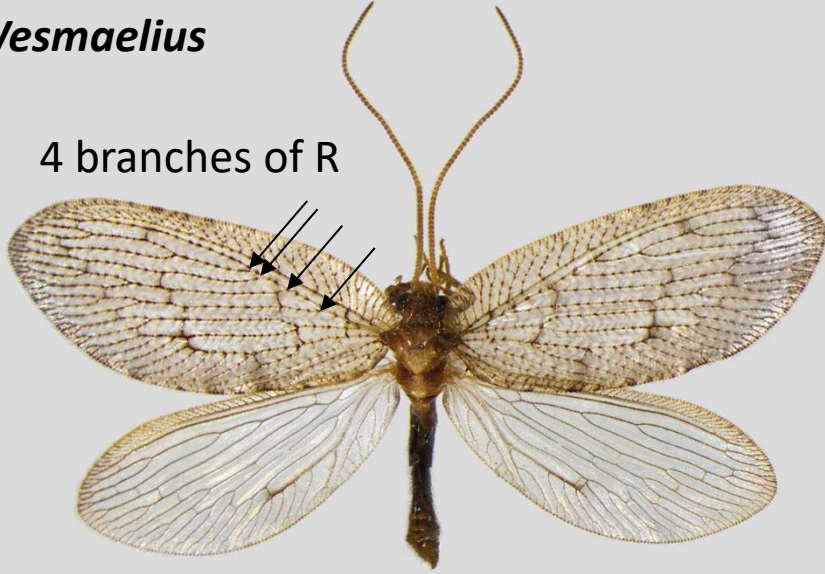
Hemerobius

Wesmaelius

Two subgenera: *Wesmaelius* and *Kimminsia*

Wesmaelius

4 branches of R



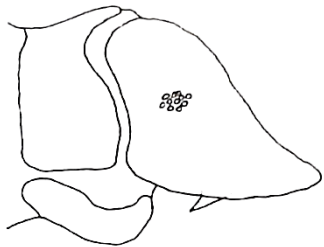
T.Rintala & T. Lehto (CC BY 4.0)

Kimminsia

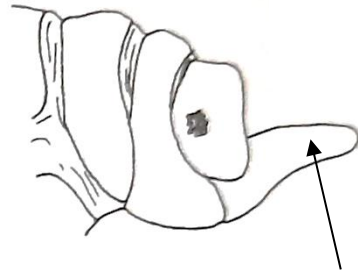
3 branches of R



T.Rintala & T. Lehto (CC BY 4.0)



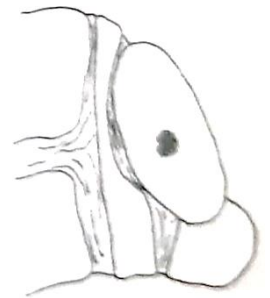
Male genitalia triangular



Female genitalia with long process



Male genitalia not triangular
various shapes

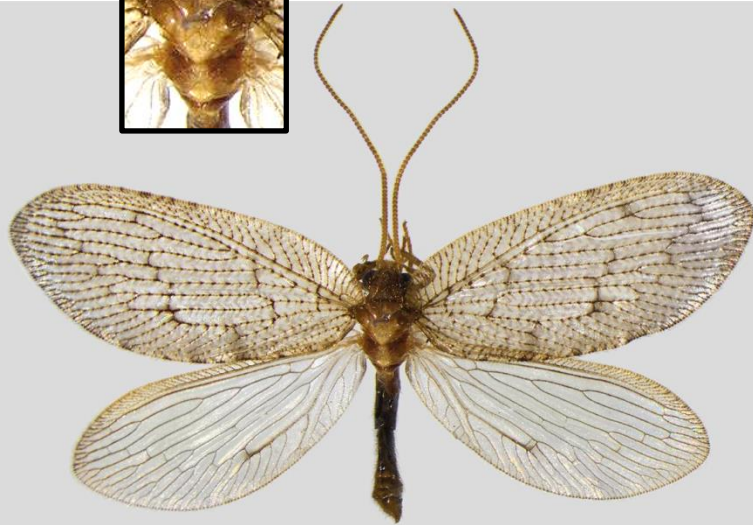


Female genitalia with
final segment rounded
not elongate

Wesmaelius (Wesmaelius) concinnus



orange-brown
no stripe



T.Rintala & T. Lehto (CC BY 4.0)

Wesmaelius (Wesmaelius) quadrifasciatus



pale stripe

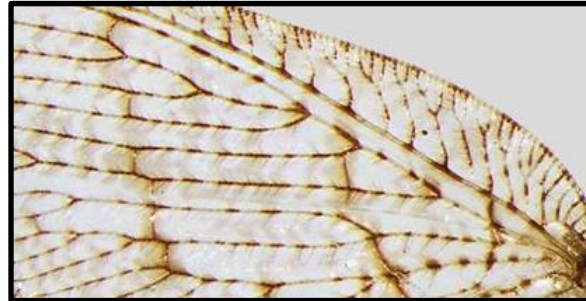


T.Rintala & T. Lehto (CC BY 4.0)



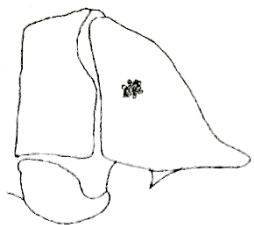
Longitudinal veins with spots (1.5-2x width of vein)

Forewing with V on membrane centred on spots, angle of V Points to wing base



Longitudinal veins with streaks (2x+ width of vein)

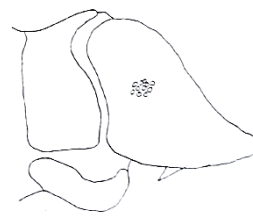
Forewing membrane with V pattern running into each other - broader



Male genitalia



Female anal plates



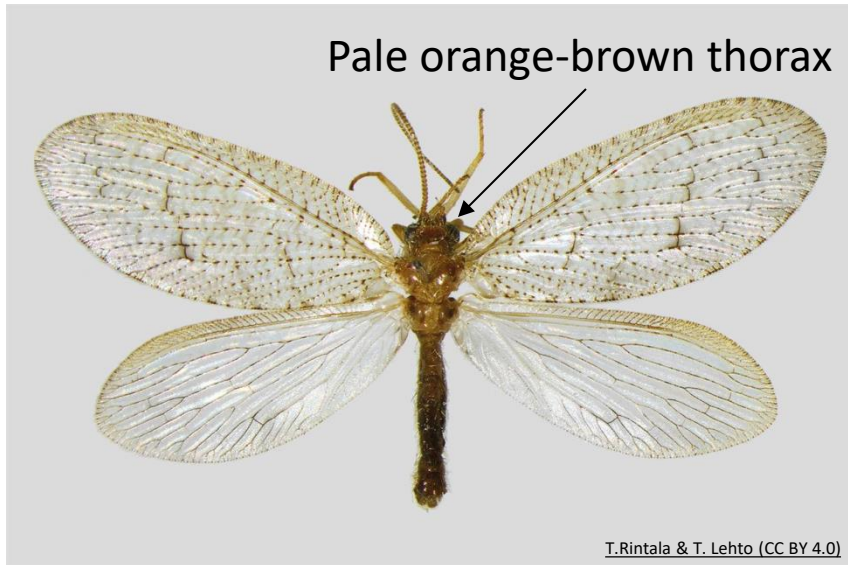
Male genitalia



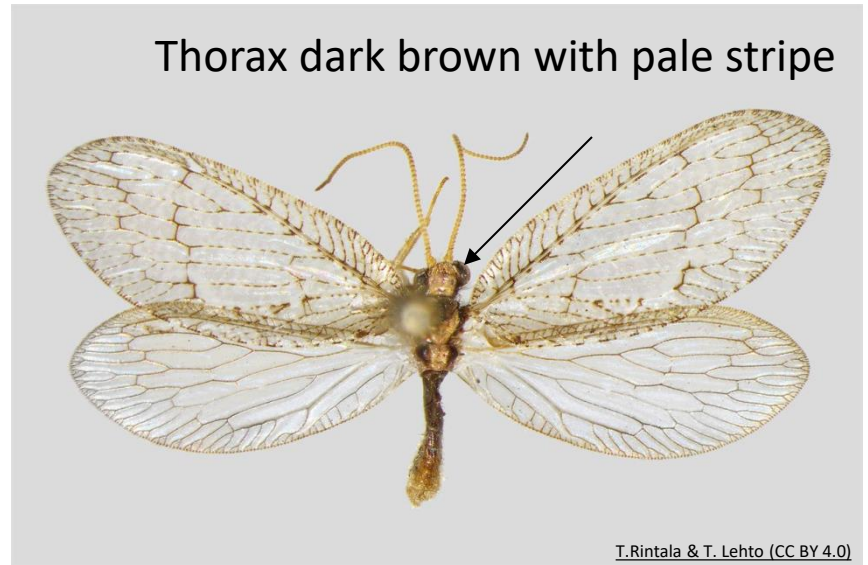
Female anal plates

Kimminsia

- Six species
- Only identified by genitalia



Wesmaelius (Kimminsia) mortoni
Wesmaelius (Kimminsia) balticus



Wesmaelius (Kimminsia) subnebulosus
Wesmaelius (Kimminsia) nervosus
Wesmaelius (Kimminsia) malladai
Wesmaelius (Kimminsia) ravus

Wesmaelius (Kimminsia) mortoni

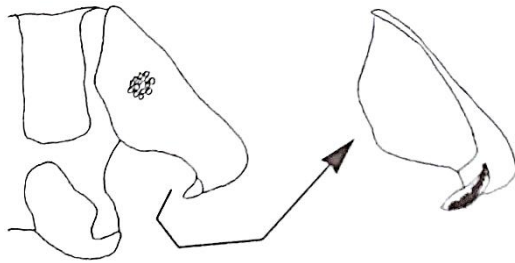
Not recorded since 1898 - extinct in British Isles?
Was recorded in Scotland
If found voucher specimen should be retained

No stripe on thorax

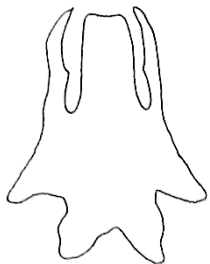


T.Rintala & T. Lehto (CC BY 4.0)

Forewing longitudinal veins with spots



Male genitalia



Female anal plate

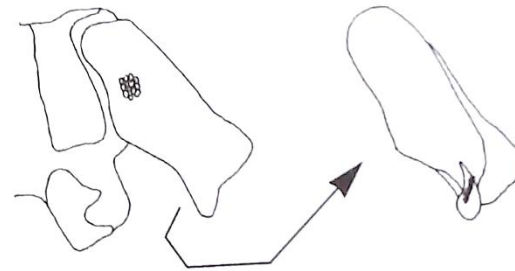
Wesmaelius (Kimminsia) balticus

No stripe on thorax

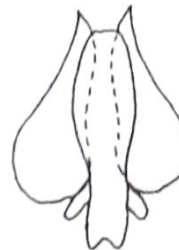


T.Rintala & T. Lehto (CC BY 4.0)

Forewing longitudinal veins without spots



Male genitalia



Female anal plate

Pale stripe on thorax

W. (Kimminsia) subnebulosus



T.Rintala & T. Lehto (CC BY 4.0)

W. (Kimminsia) nervosus



T.Rintala & T. Lehto (CC BY 4.0)

W. (Kimminsia) malladai



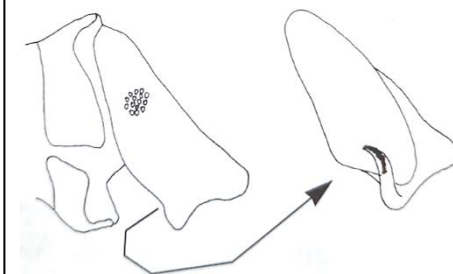
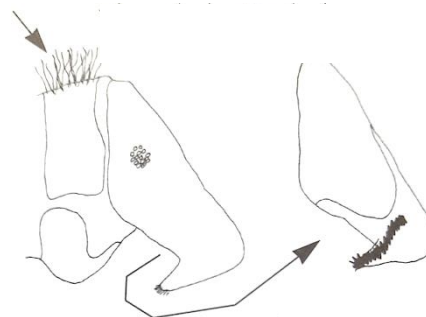
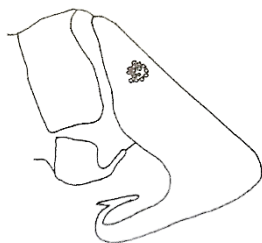
T.Rintala & T. Lehto (CC BY 4.0)

W. (Kimminsia) ravus

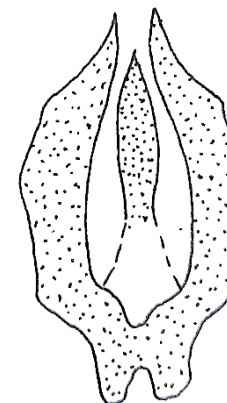
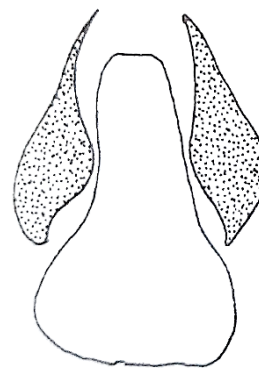
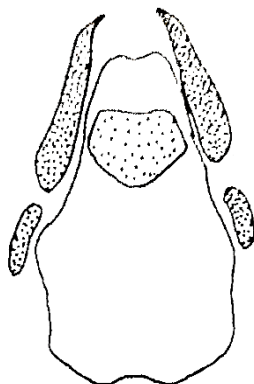
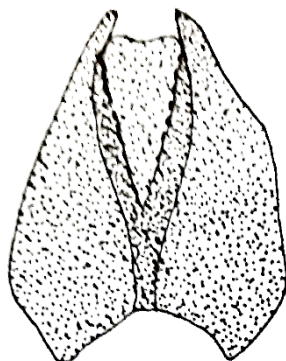


T.Rintala & T. Lehto (CC BY 4.0)

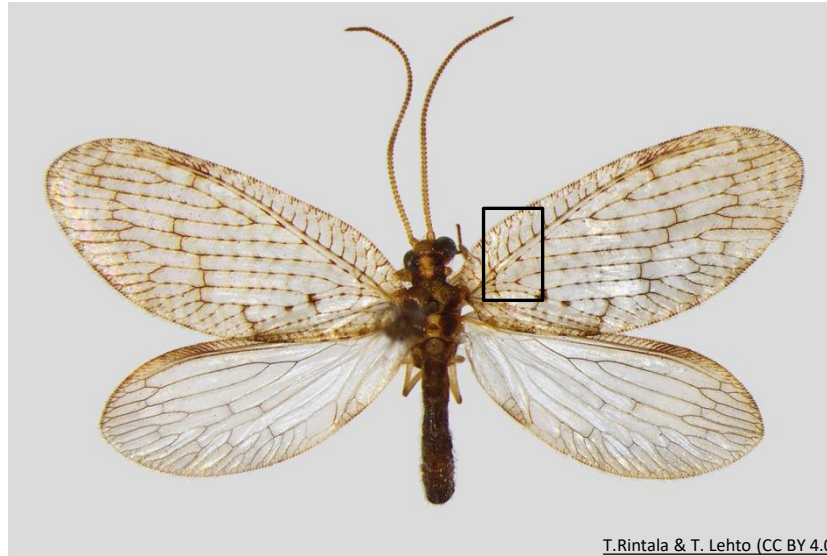
Male genitalia:



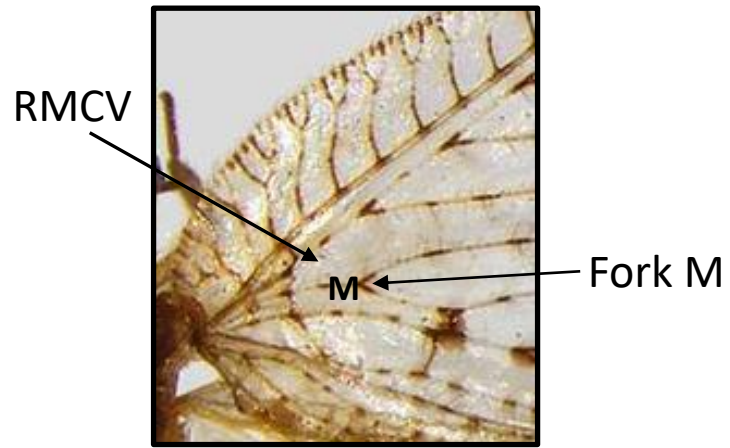
Female anal plates:



RMCV pale and before fork of M



T.Rintala & T. Lehto (CC BY 4.0)



Hemerobius

Media-Cubitus crossvein (MCCV) and Forewing patterns



MCCV colourless (if dark membrane around clear)
Forewing no pattern

Hemerobius nitidulus, H. micans



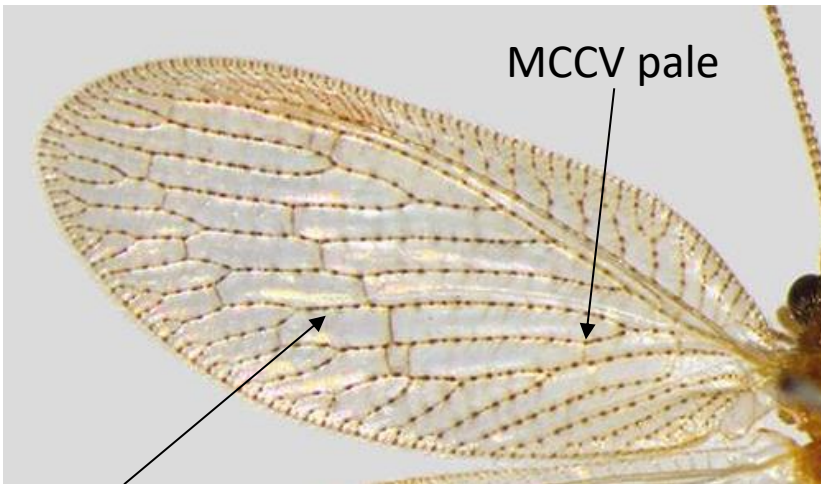
MCCV dark
Forewing with pattern

Hemerobius stigma, H. atrifrons, H. marginatus, H. lutescens, H. striatus, H. contumax, H. pini, H. simulans, H. perelegans, H. humulinus

Hemerobius nitidulus

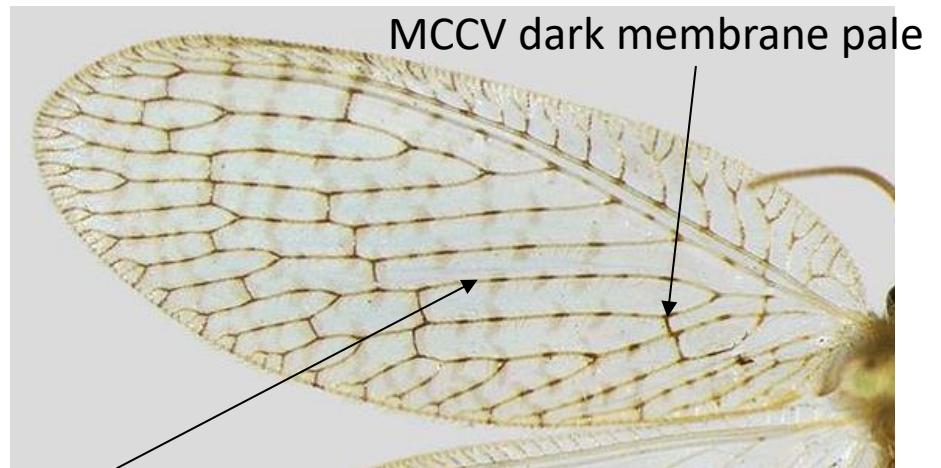


Hemerobius micans



MCCV pale

Main veins with spots
Wing hairs from dots dark



MCCV dark membrane pale

Main veins with dashes (2x long as width)
Wing hairs from dashes pale



MCCV dark
Forewing with pattern

***Hemerobius stigma, H. atrifrons, H. marginatus,
H. lutescens H. striatus, H. contumax, H. pini,
H. simulans, H. perelegans, H. humulinus***

Thorax stripe

Orange coloured pterostigma



Without stripe



Hemerobius stigma



With stripe



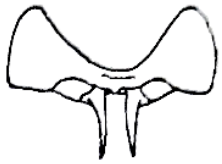
Hemerobius atrifrons, *H. marginatus*,
H. lutescens, *H. striatus*, *H. contumax*, *H. pini*,
H. simulans, *H. perelegans*, *H. humulinus*

Face colour

Front of head shining glossy black



Not shiny, brown face



Plant, 1997

Internal male
genitalia



Hemerobius atrifrons

H. marginatus, *H. lutescens*, *H. striatus*,
H. contumax, *H. pini*, *H. simulans*,
H. perelegans, *H. humulinus*

Basal SC-R crossvein



Pale or only slightly darkened
at the end that joins to R

Hemerobius marginatus, H. lutescens



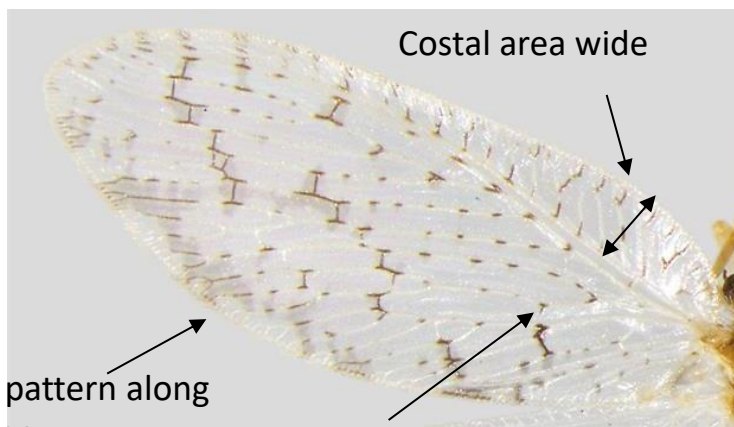
Dark, usually black

Hemerobius striatus,
H. contumax, H. pini, H. simulans,
H. perelegans, H. humulinus

Hemerobius marginatus



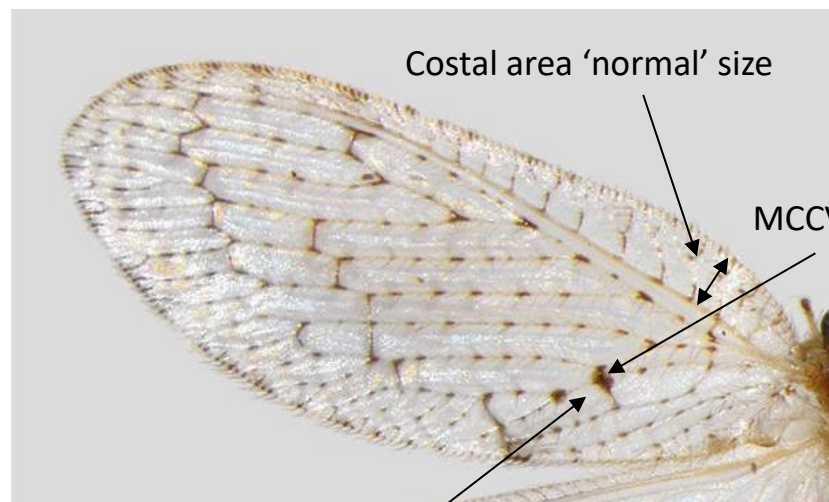
Hemerobius lutescens



Costal area wide

Distinct pattern along wing edge

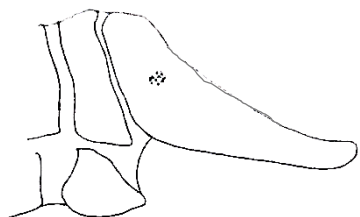
MCCV dark



Costal area 'normal' size

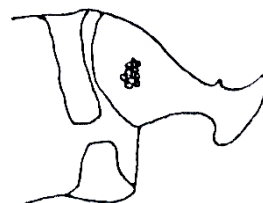
MCCV dark

Pattern at base of wing



Male genitalia

Plant, 1997



Male genitalia

Plant, 1997

Basal SC-R crossvein



Dark, usually black

Hemerobius striatus,
H. contumax, H. pini, H. simulans,
H. perelegans, H. humulinus

Forewing hind margin shading



Uniform shading

Hemerobius striatus, *H. contumax*, *H. pini*



Not uniform some transparent areas

H. simulans, *H. perelegans*, *H. humulinus*

Forewing inner and outer gradate series of crossveins

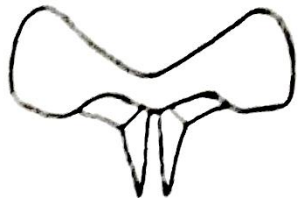
O. Fogh Nielsen (CC BY 4.0)



inner outer

Inner dark and outer pale

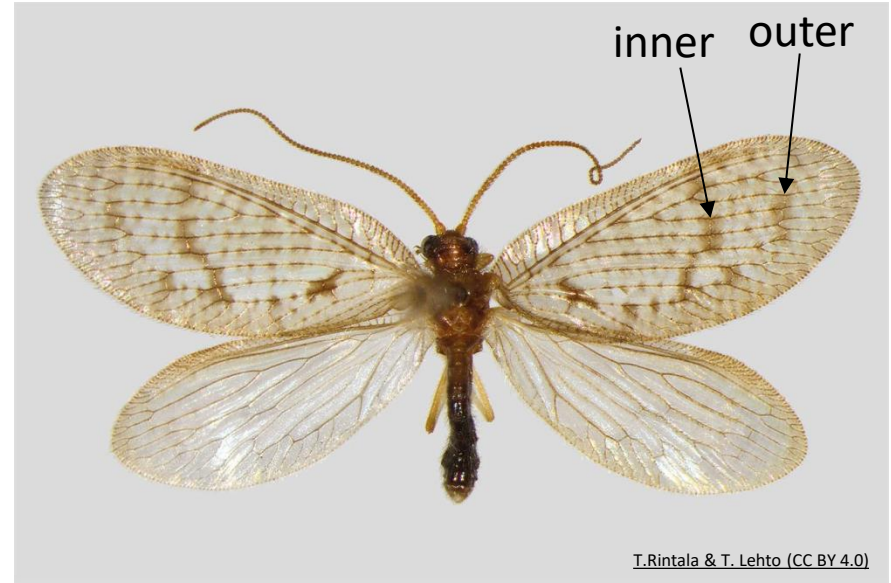
Hemerobius striatus (= *Hemerobius fenestratus*)



Internal male genitalia

Plant, 1997

Recently found
Records require verification



inner outer

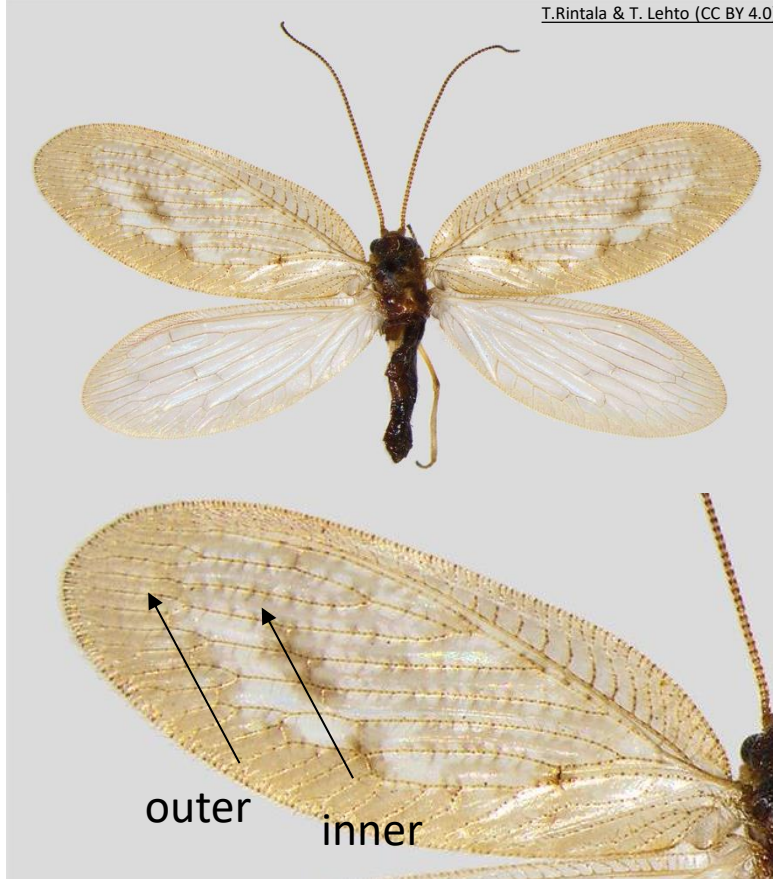
T.Rintala & T. Lehto (CC BY 4.0)

Both inner and outer dark

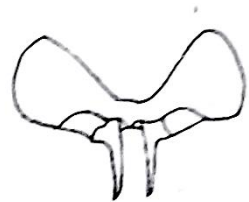
Hemerobius contumax, *H. pini*

Hemerobius contumax

T.Rintala & T. Lehto (CC BY 4.0)



Gradate series of crossveins close and parallel



Plant, 1997

Internal male genitalia

Not recorded since 1952 – possibly extinct in Britain
Was recorded in Surrey, Hampshire, Buckinghamshire, Sussex
Records require verification

Hemerobius pini

T.Rintala & T. Lehto (CC BY 4.0)



Gradate series of crossveins diverging not parallel



Plant, 1997

Internal male genitalia

Forewing hind margin shading not uniform

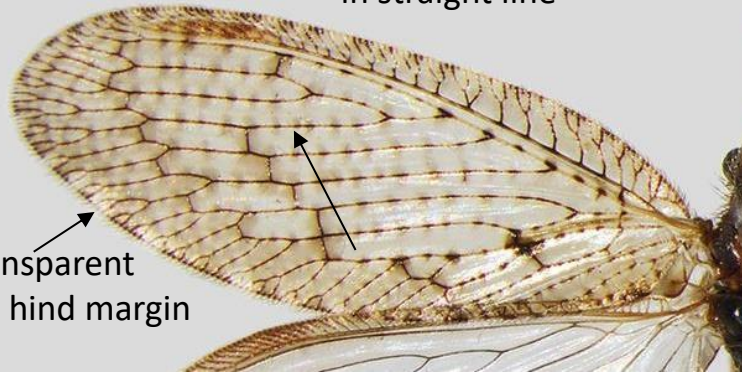


Not uniform some transparent areas
H. simulans, *H. perelegans*, *H. humulinus*

T.Rintala & T. Lehto (CC BY 4.0)

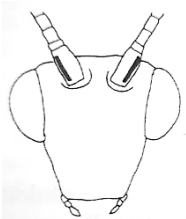


Inner gradate series
in straight line



Small transparent
Areas on hind margin

Wing span greater than 20 mm



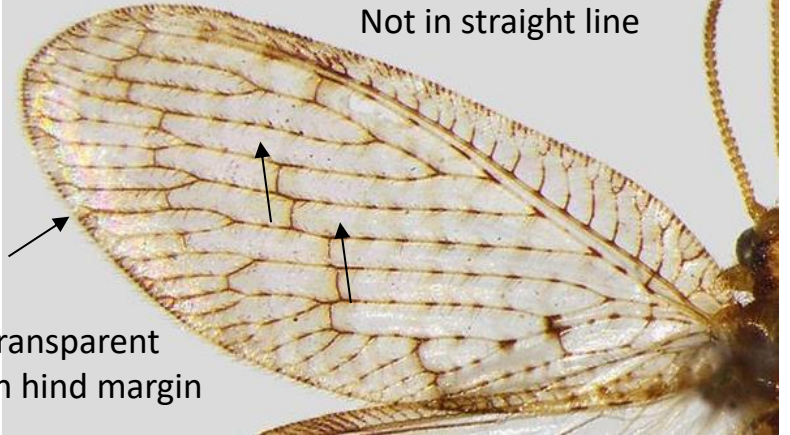
Scape usually with dark stripe

Hemerobius simulans

T.Rintala & T. Lehto (CC BY 4.0)



Inner gradate series
Not in straight line



Larger transparent
Areas on hind margin

Wing span less than 18 mm

Scape without dark stripe

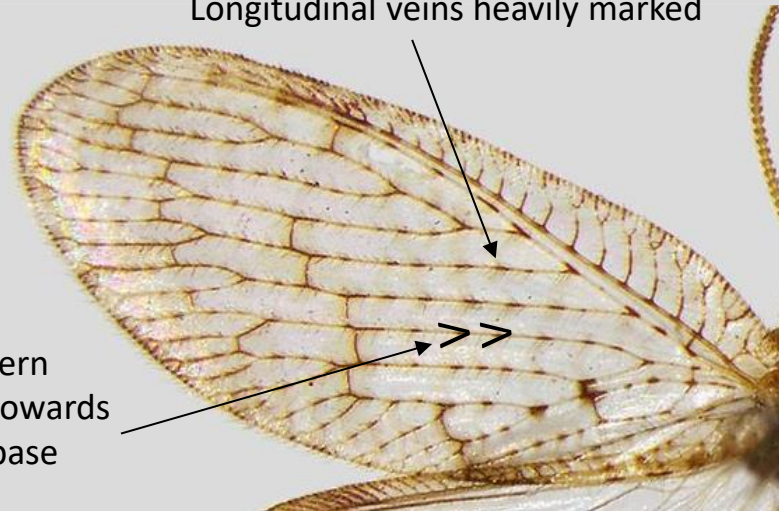
H. perelegans, H. humulinus

Hemerobius perelegans

T.Rintala & T. Lehto (CC BY 4.0)



Longitudinal veins heavily marked



V pattern apex towards wing base

Red brown forewings

Hemerobius humulinus

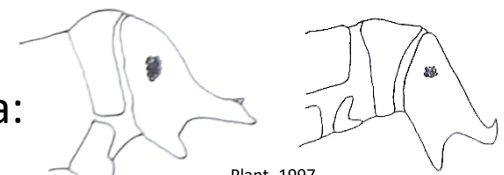
T.Rintala & T. Lehto (CC BY 4.0)



Pale forewings

Any V shape pattern pale and not standing out

Male genitalia:



Plant, 1997

Chrysopidae (Green Lacewings)

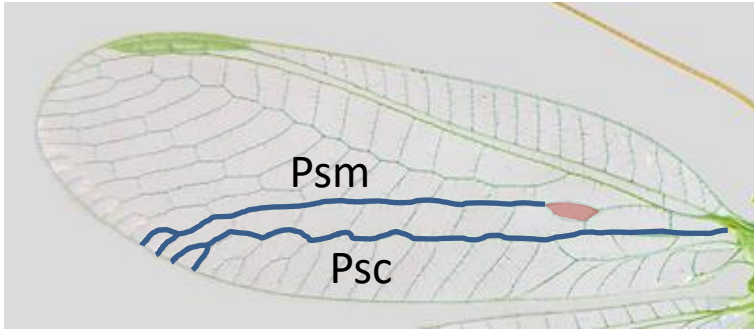
- Can be preserved as both pinned and in alcohol, usually pinned
- Can generally be identified without abdomen clearing or dissection



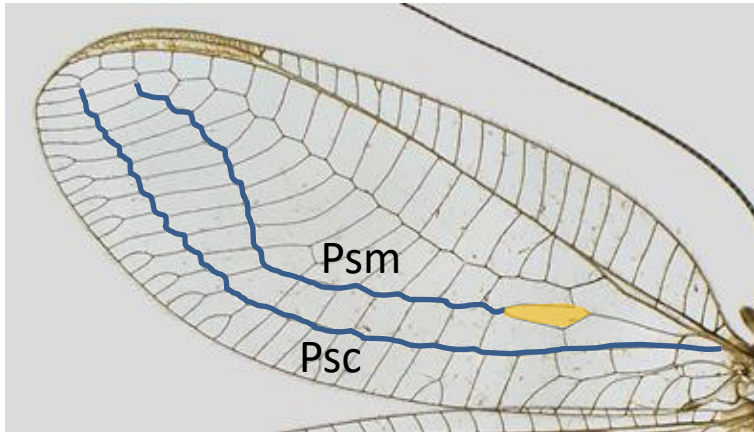
Pseudomesia and Pseudocubitus

Psm = Pseudomesia

Psc = Pseudocubitus



Other genera



Nothochrysa species



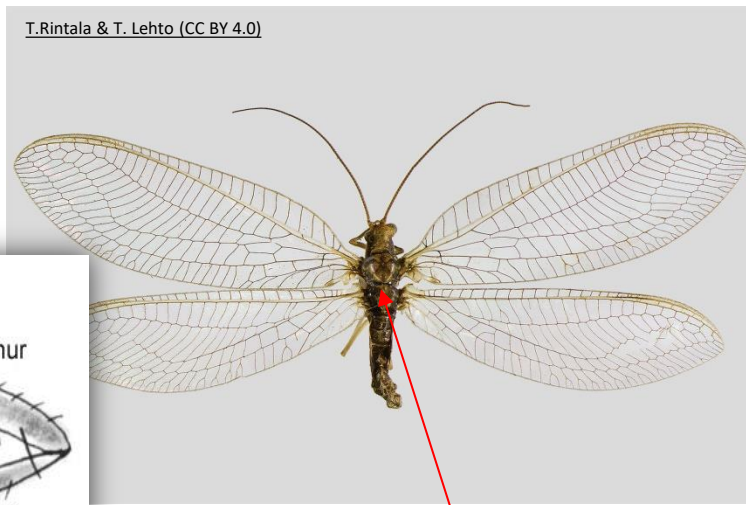
Nothochrysa capitata

T.Rintala & T. Lehto (CC BY 4.0)



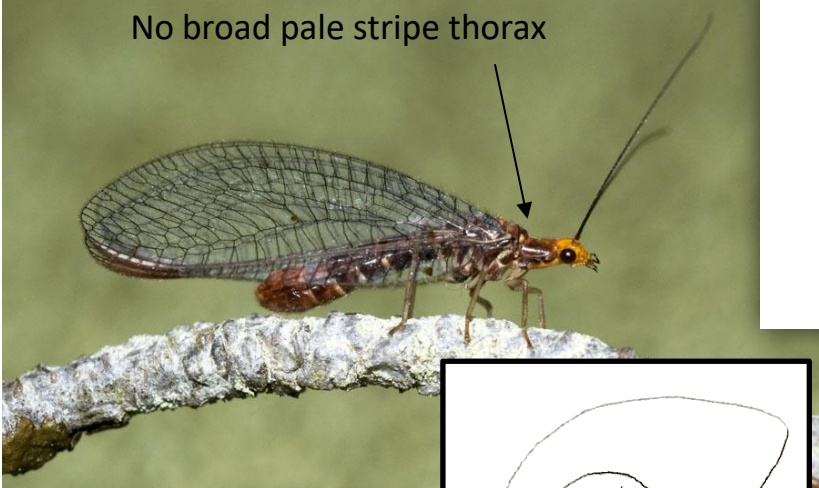
Nothochrysa fulviceps

T.Rintala & T. Lehto (CC BY 4.0)

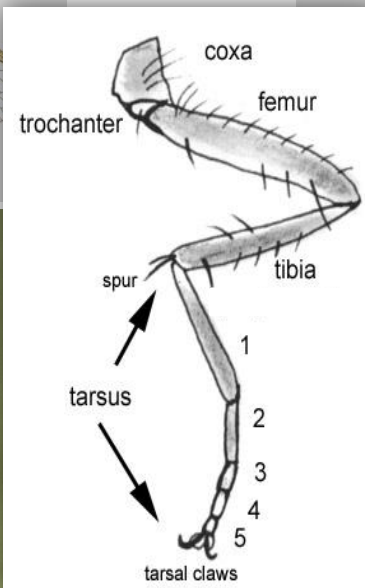
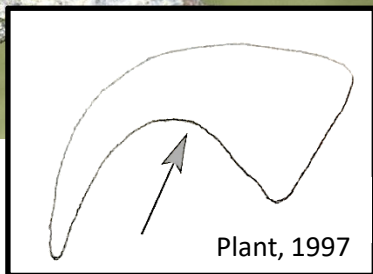


O. Fogh Nielsen (CC BY 4.0)

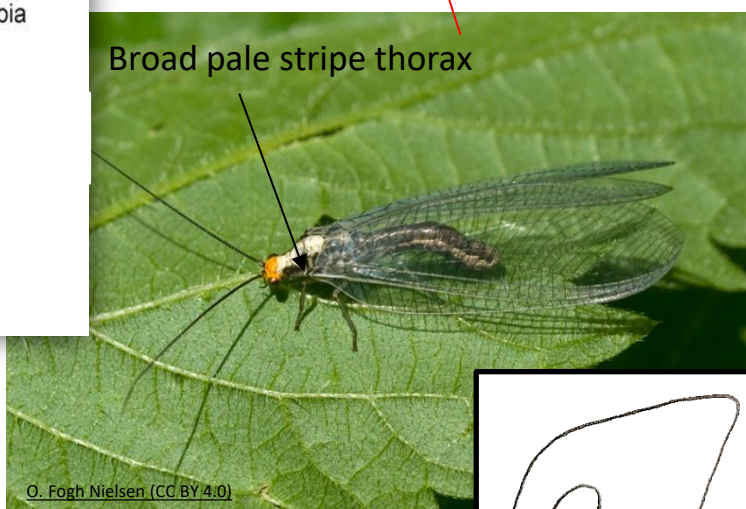
No broad pale stripe thorax



Tarsal claw simple

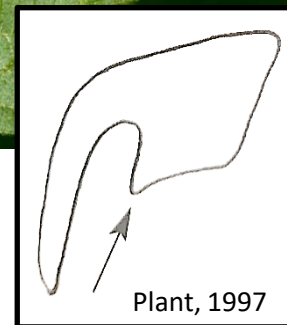


Broad pale stripe thorax



O. Fogh Nielsen (CC BY 4.0)

Tarsal claw with tooth

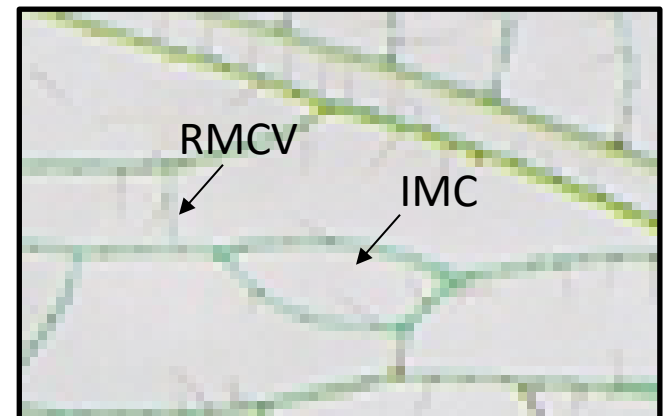


Relationship of Intramedial cell and Radial-Media Crossvein



T.Rintala & T. Lehto (CC BY 4.0)

- Intermedial cell (IMC)
- Radial-Media crossvein (RMCV)



Peyerimhoffina gracilis

Inner gradate series
of crossveins with at least
2x as many as outer series



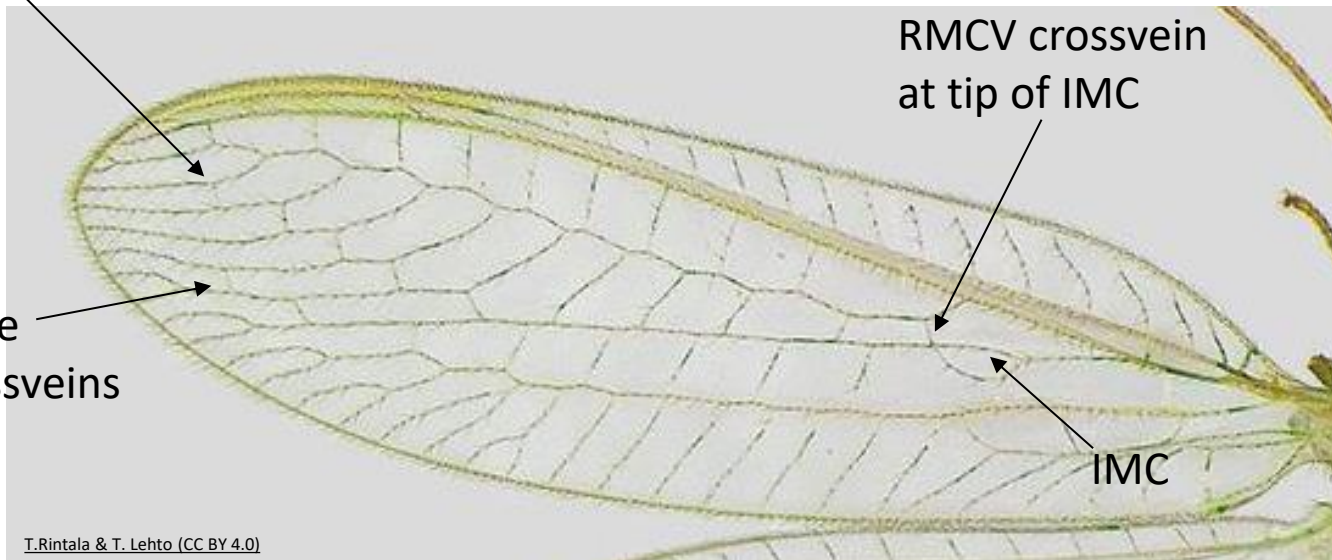
T.Rintala & T. Lehto (CC BY 4.0)

Inner gradate series
of crossveins

Outer gradate
series of crossveins

RMCV crossvein
at tip of IMC

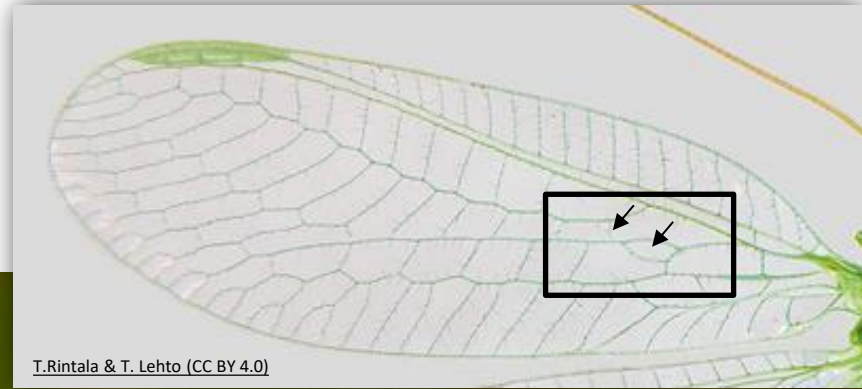
IMC



T.Rintala & T. Lehto (CC BY 4.0)

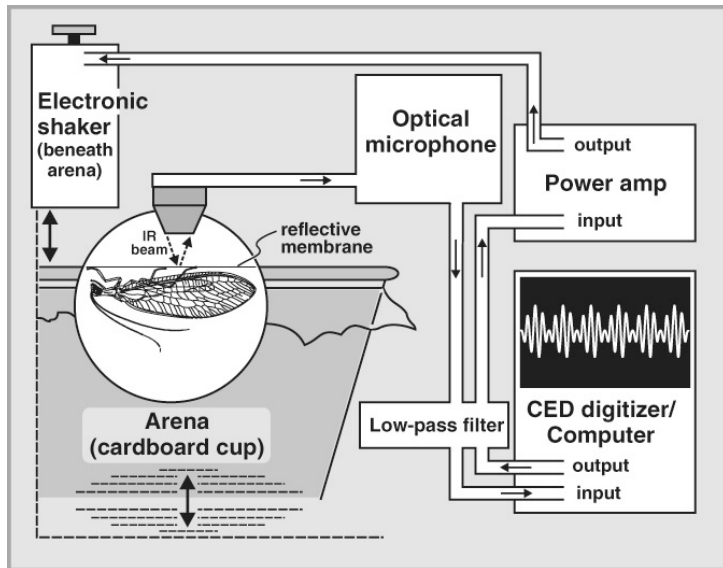
Chrysoperla carnea group

- *Chrysoperla carnea*
- *Chrysoperla lucasina*
- *Chrysoperla pallida*

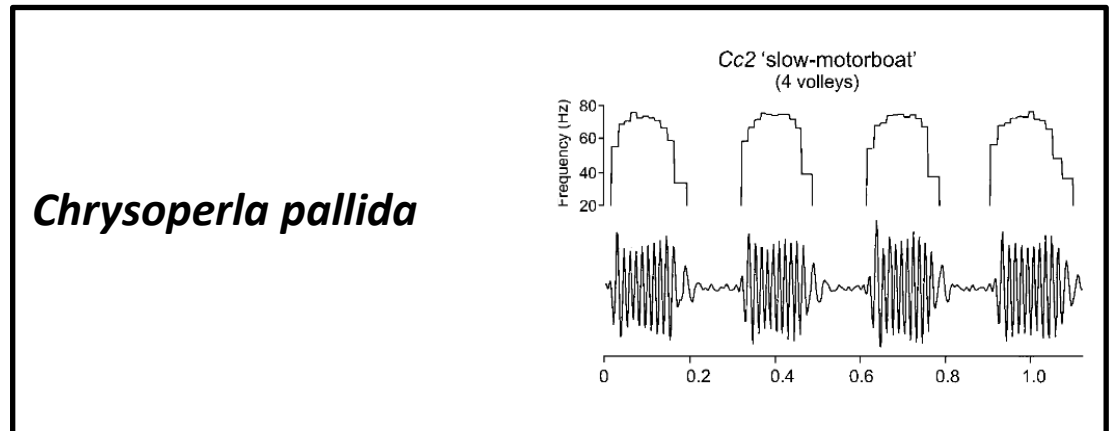
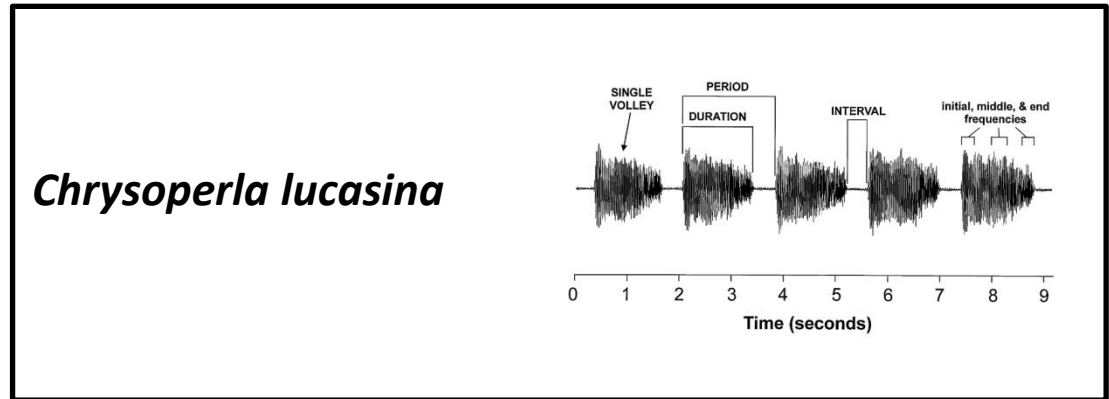
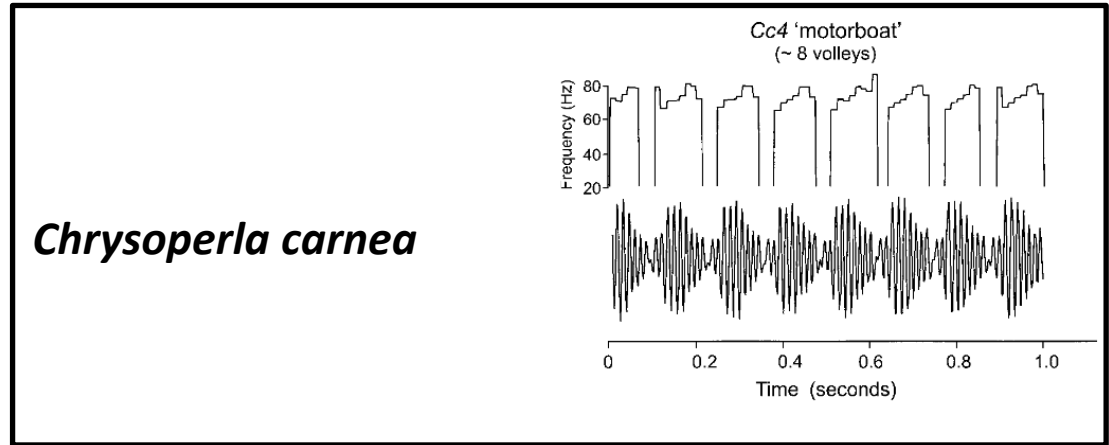


Songs

- Substrate based song by abdominal vibration to attract a mate
- The abdomen doesn't strike the substrate – insect shakes the leaf or conifer needle which its standing on
- Travels around a metre at most



Henry et al. 2013



Henry et al. 2002

Chrysoperla carnea

T.Rintala & T. Lehto (CC BY 4.0)



More rounded wing tip

Become brown for
Autumn/Winter
hibernation

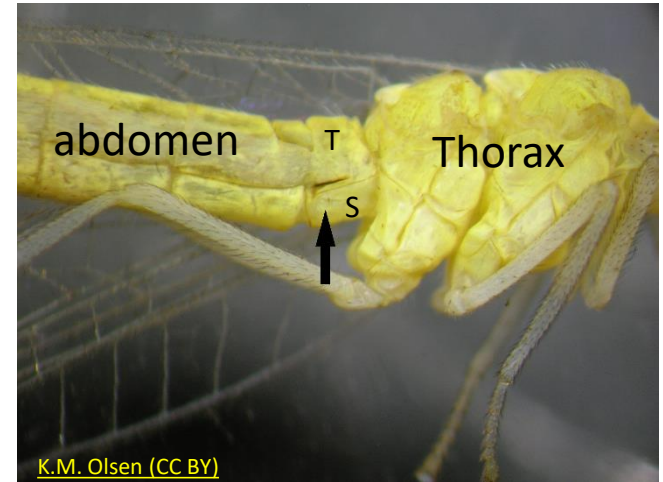
O. Fogh Nielsen (CC BY 4.0)



Chrysoperla lucasina

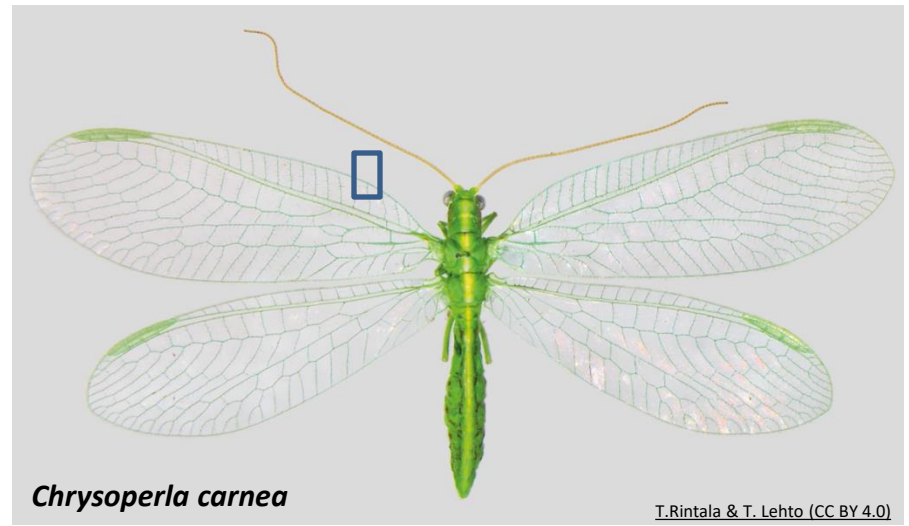
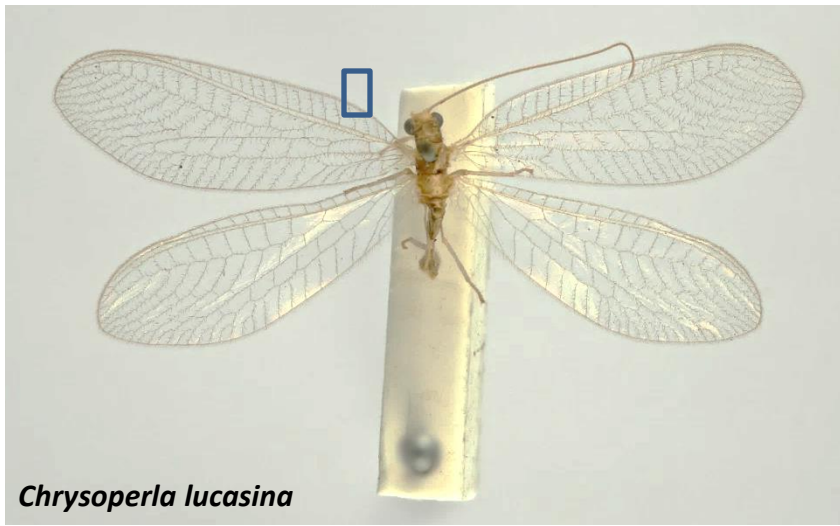


Wing tip appears more pointed

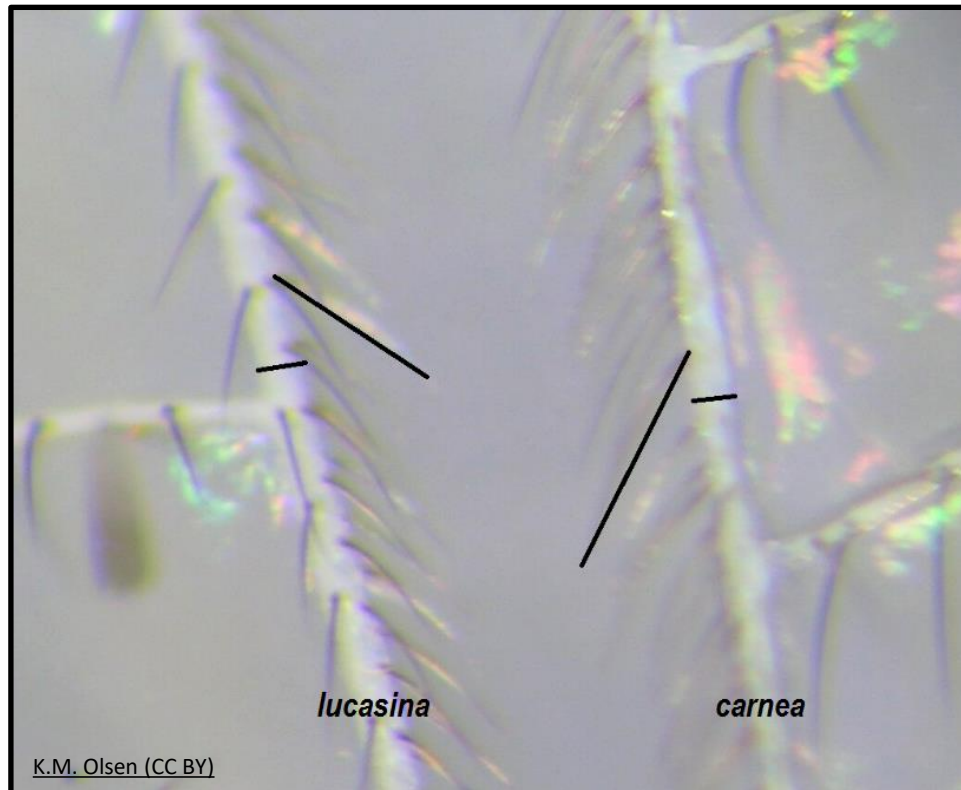


Brown line on membrane between first 1 or 2 tergite (upper) and sternite (lower).

Ignore any colour on tergite (T) and sternites (S).



Shorter hairs



Longer hairs

Chrysoperla pallida

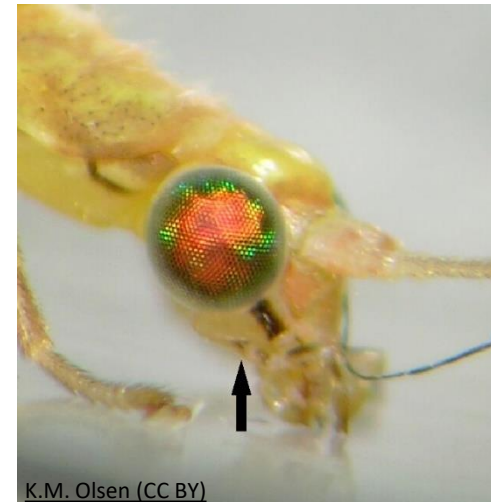
- A newly described species (described 2002)
- Identified from song and some morphological traits



K.M. Olsen (CC BY)

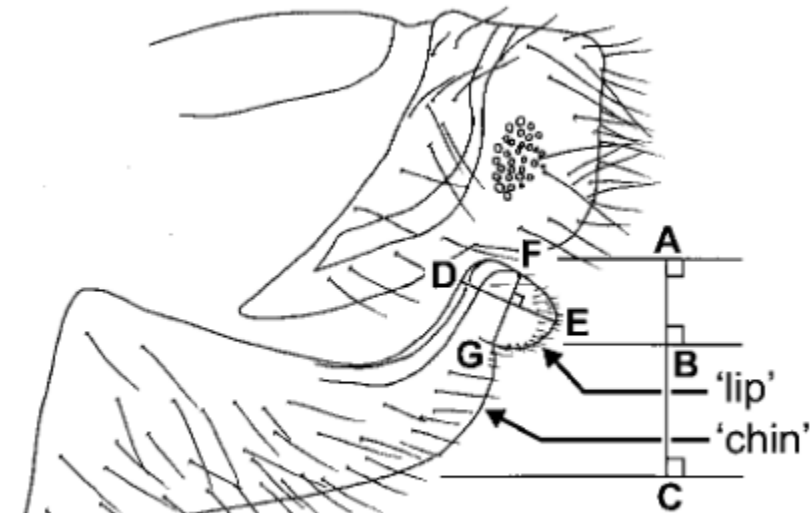


Abdomen with blond setae



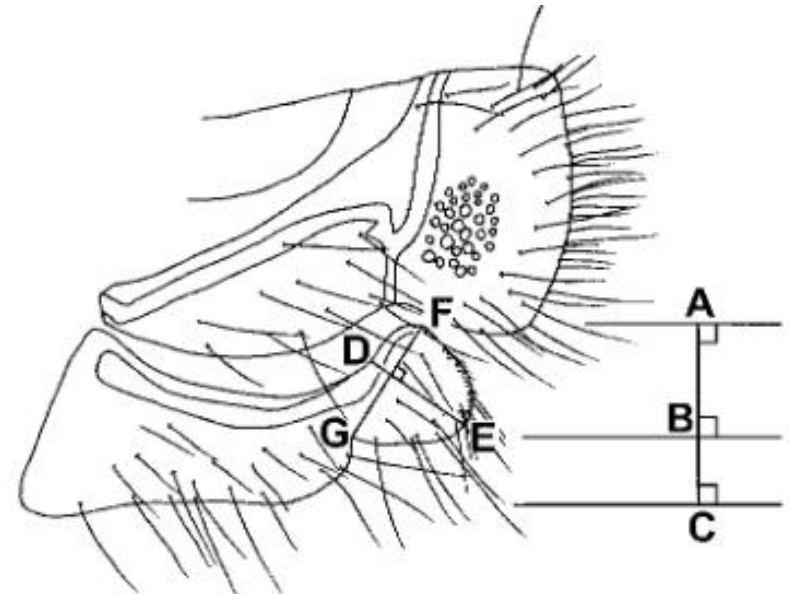
Narrow brown stripe on the cheek (gena) below the eyes

Lip short and narrow with short pale setae.
Chin broad



Chrysoperla pallida

Lip broader and more protuberant
and bears several long, coarse dark setae.
Chin narrow



Chrysoperla carnea

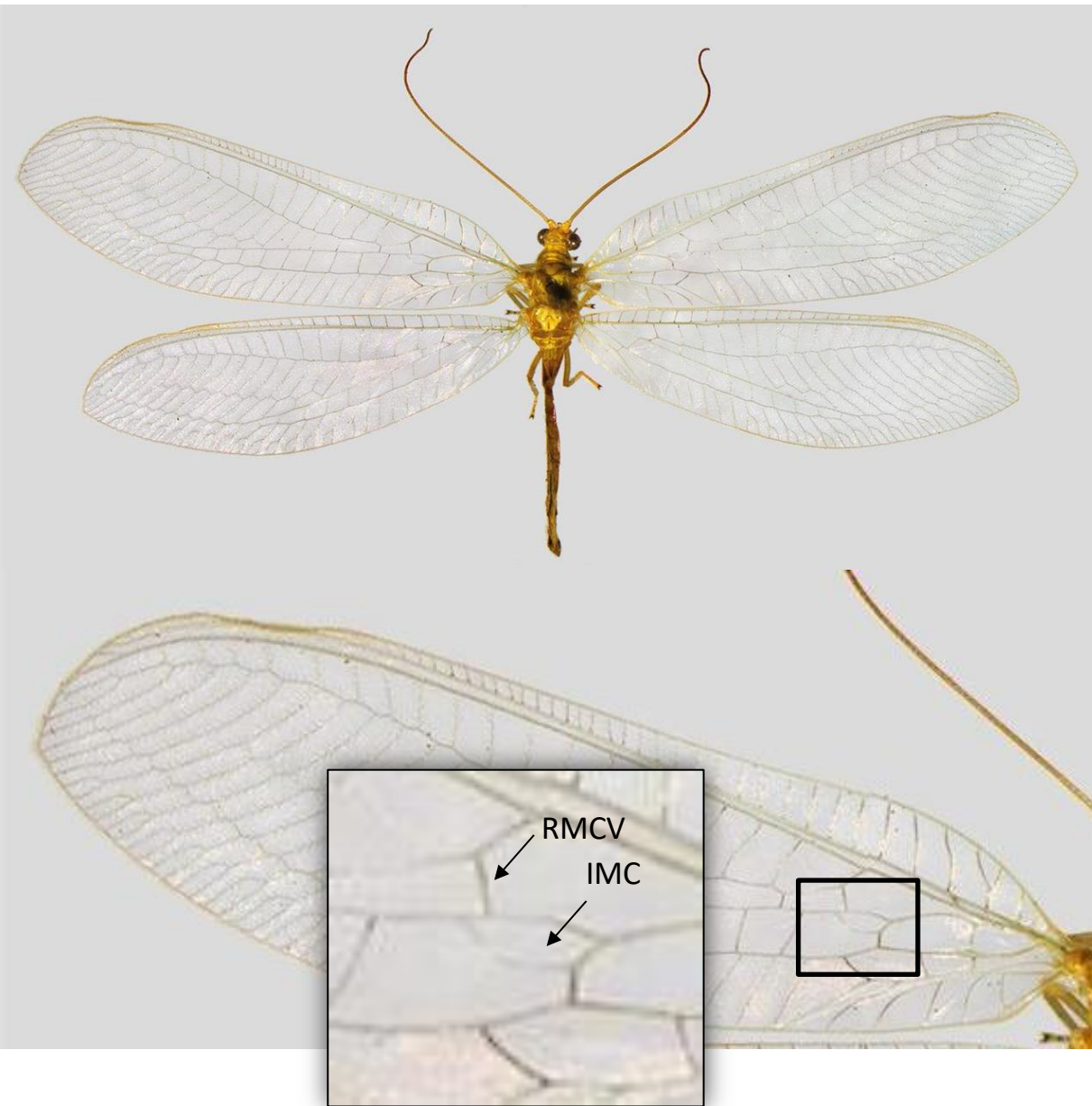
Terminus of male abdomen showing line segments AC, AB, DE, and FG used for calculating the ratios describing the shape of the genital “lip” and “chin”.

Narrow lip = narrow and not protruding if $DE > FG$

Broad chin = if $AB < BC$

If in doubt

Record as *Chrysoperla carnea* group (agg.)

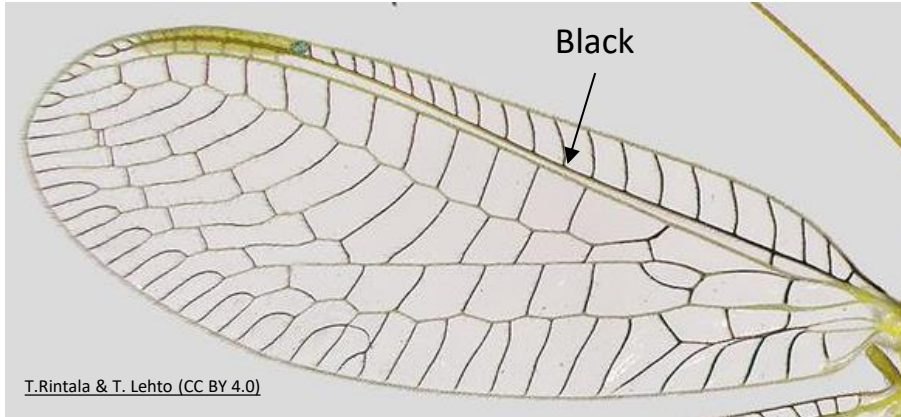


- *Chrysopa dorsalis*
- *Chrysopa perla*
- *Chrysopa commata*
- *Chrysopa abbreviata*
- *Chrysopa phyllochroma*
- *Chrysopa pallens*
- *Apertochrysa ventralis*
- *Apertochrysa prasina*
- *Apertochrysa flavifrons*
- *Nineta vittata*
- *Nineta flava*
- *Nineta inpunctata*
- *Nineta pallida*
- *Chrysopidia ciliata*
- *Cunctochrysa albolineta*
- *Cunctochrysa cosmia*

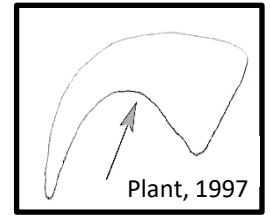
Vein Sc black or green

Black:

Chrysopa dorsalis



Second antennal segment (pedicel) black



Tarsal claw simple

Head with extensive black markings
Small pale elongate area on top of head

Green:

- Chrysopa perla*
- Chrysopa commata*
- Chrysopa abbreviata*
- Chrysopa phyllochroma*
- Chrysopa pallens*
- Apertochrysa ventralis*
- Apertochrysa prasina*
- Apertochrysa flavifrons*
- Nineta vittata*
- Nineta flava*
- Nineta inpunctata*
- Nineta pallida*
- Chrysopidia ciliata*
- Cunctochrysa albolineta*
- Cunctochrysa cosmia*



Green

Second antennal segment: black or green



Second antennal segment black

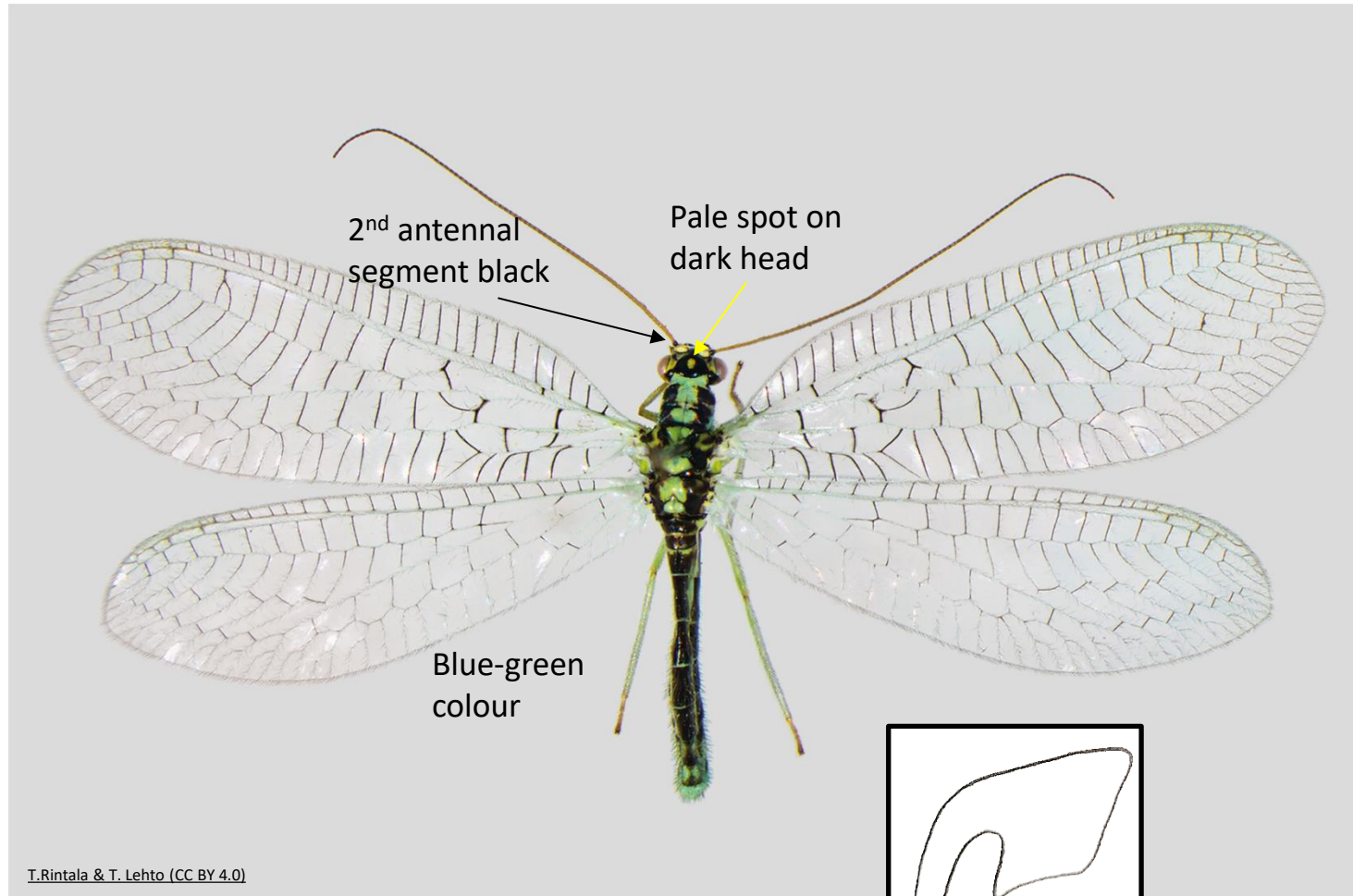
Chrysopa perla
Chrysopa commata
Chrysopa abbreviata
Chrysopa phyllochroma



Second antennal segment green

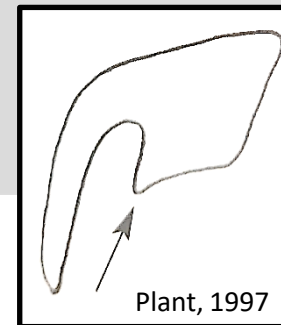
Chrysopa pallens
Apertochrysa ventralis
Apertochrysa prasina
Apertochrysa flavifrons
Nineta vittata
Nineta flava
Nineta inpunctata
Nineta pallida
Chrysopidia ciliata
Cunctochrysa albolineta
Cunctochrysa cosmia

Chrysopa perla



T.Rintala & T. Lehto (CC BY 4.0)

Tarsal claw with tooth



Plant, 1997

Chrysopa commata



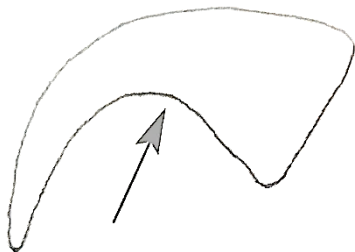
O. Fogh Nielsen (CC BY 4.0)



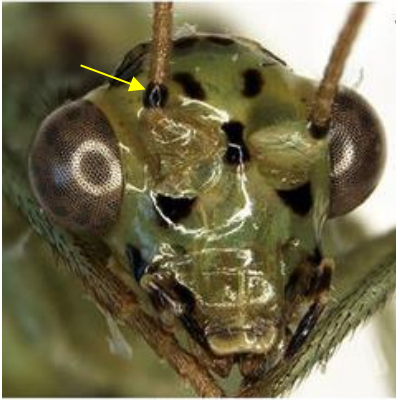
T. Rintala & T. Lehto (CC BY 4.0)



Sutures on thorax black



Tarsal claw simple



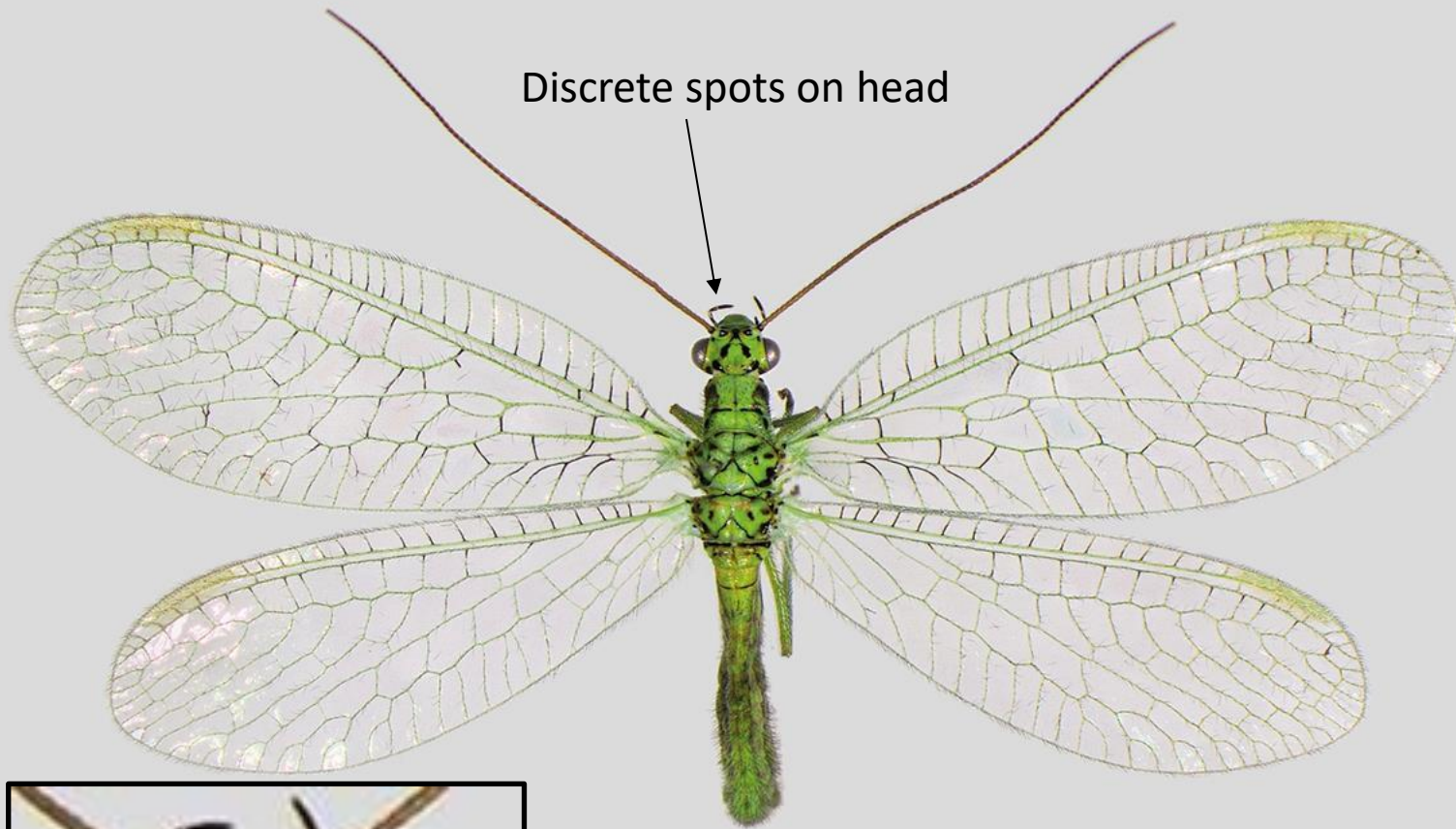
Second antennal segment black



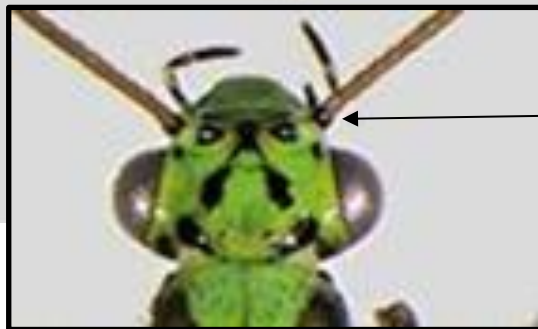
Discrete spots on the head

Chrysopa abbreviata

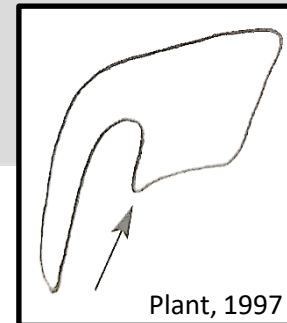
T.Rintala & T. Lehto (CC BY 4.0)



Discrete spots on head



Second antennal segment black



Tarsal claw with tooth

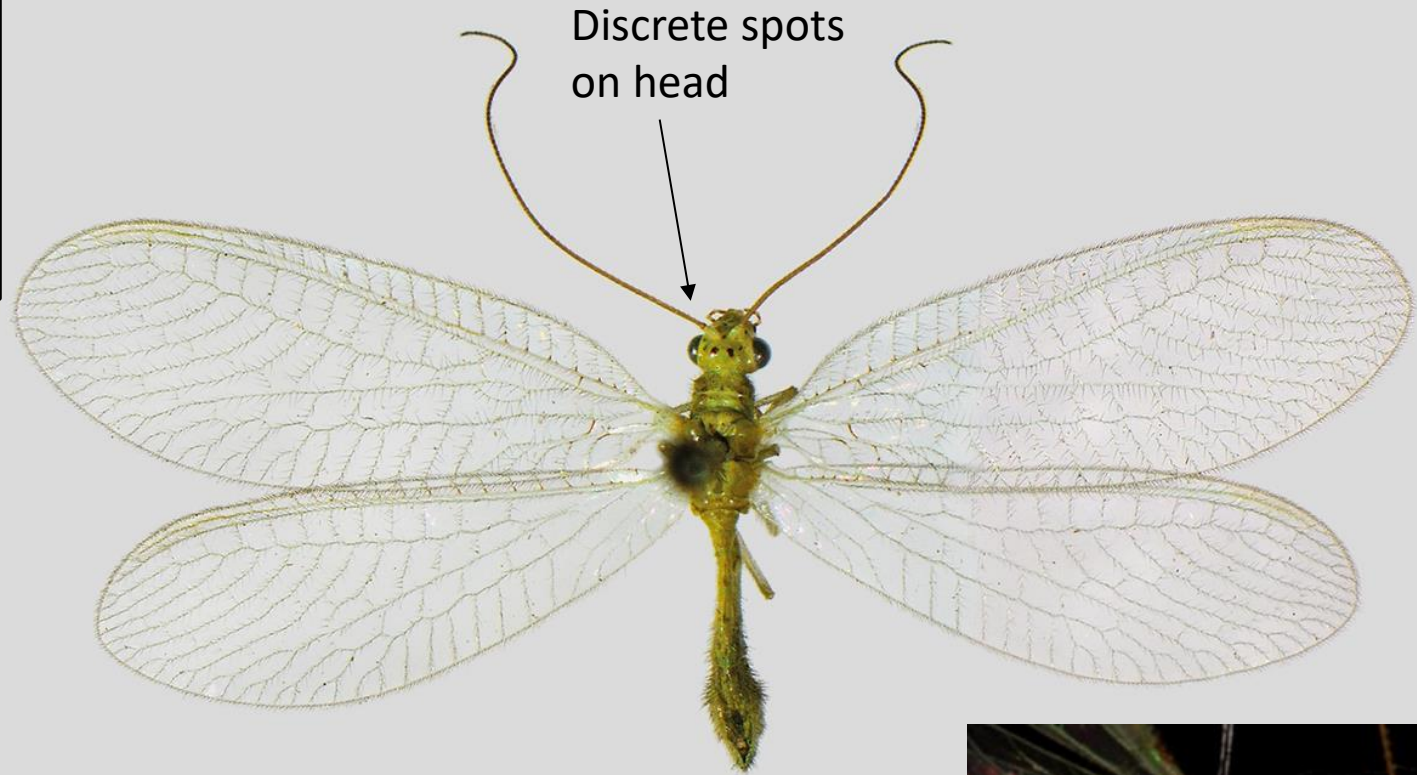
Plant, 1997

Chrysopa phyllochroma

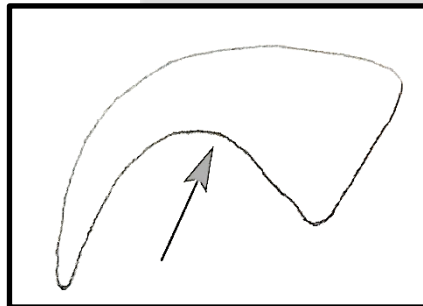
T. Rintala & T. Lehto (CC BY 4.0)



2nd antennal segment black



Discrete spots on head



Tarsal claw simple

Side of thorax green
no dark lines



Duelli et al. 2016



Second antennal segment green

Chrysopa pallens

Apertochrysa ventralis

Apertochrysa prasina

Apertochrysa flavifrons

Nineta vittata

Nineta flava

Nineta inpunctata

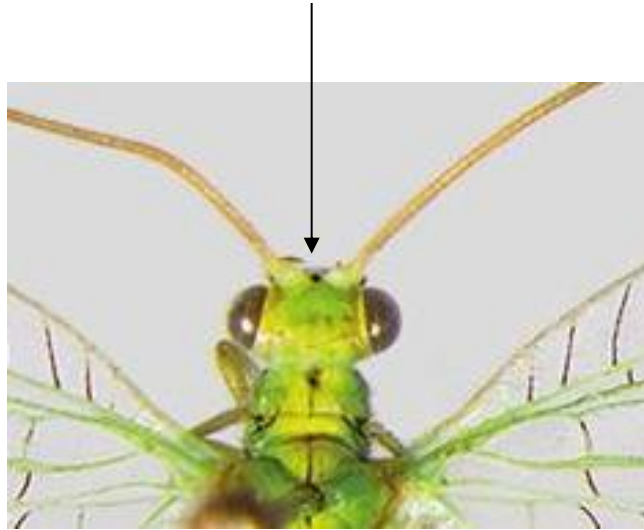
Nineta pallida

Chrysopidia ciliata

Cunctochrysa albolineta

Cunctochrysa cosmia

Black spot between the antennae



T. Rintala & T. Lehto (CC BY 4.0)

Apertochrysa ventralis

Apertochrysa prasina

Chrysopa pallens

Apertochrysa ventralis (= *Dichochrysa ventralis*)

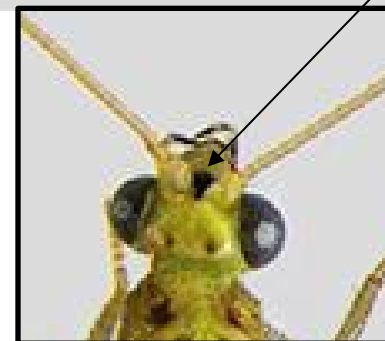
T.Rintala & T. Lehto (CC BY 4.0)



Black spot
Between
antennae

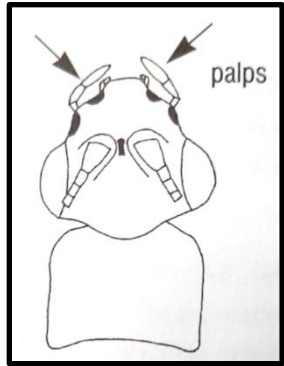


Under surface of
abdomen glossy
black



T.Rintala & T. Lehto (CC BY 4.0)

Apertochrysa prasina (= *Dichochrysa prasina*)



3-5 spots on head

T.Rintala & T. Lehto (CC BY 4.0)

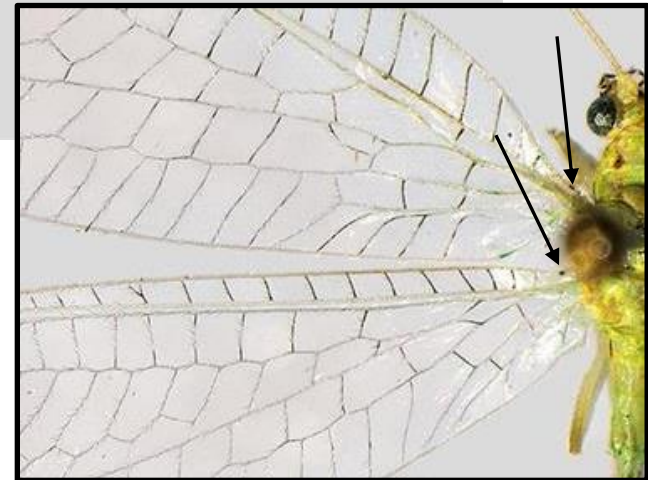
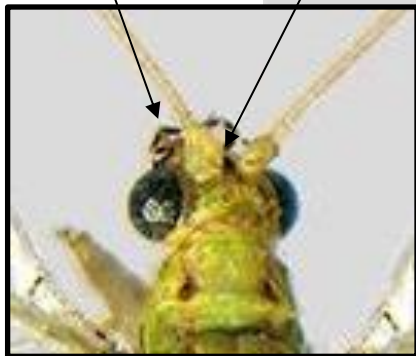


Dark basal costal crossvein

Dark spot at base of Costa

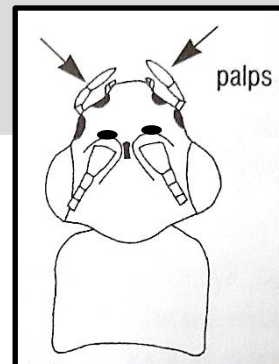
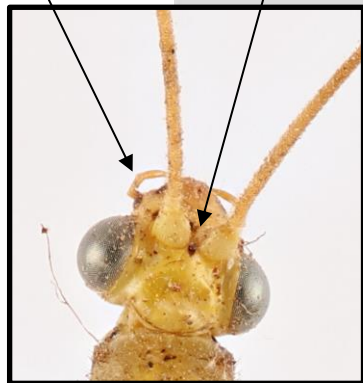
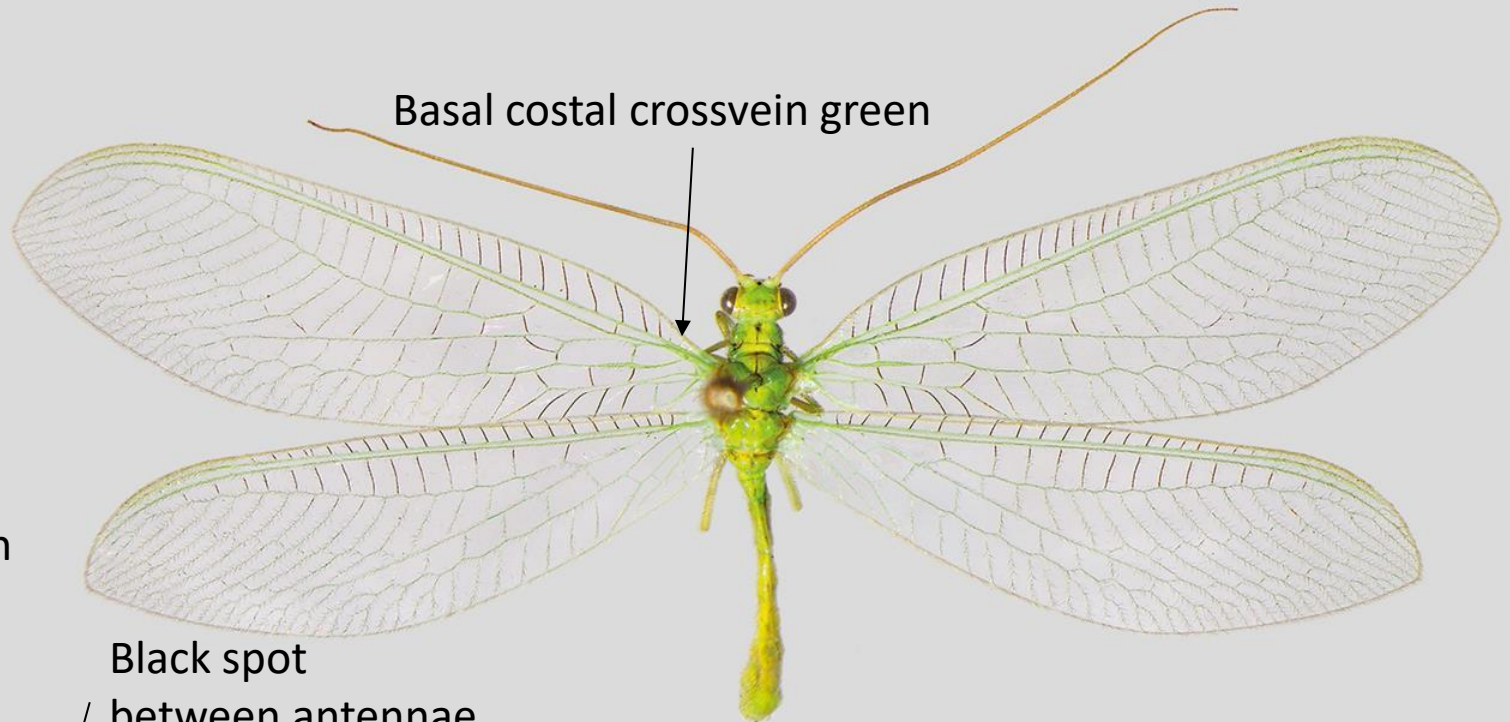
Palps ringed with black
(appear entirely black in dried specimens)

Black spot between antennae



Chrysopa pallens

T.Rintala & T. Lehto (CC BY 4.0)



7 spots on head

No black spot between the antennae



T.Rintala & T. Lehto (CC BY 4.0)

Apertochrysa flavifrons

Nineta vittata

Nineta flava

Nineta inpunctata

Nineta pallida

Chrysopidia ciliata

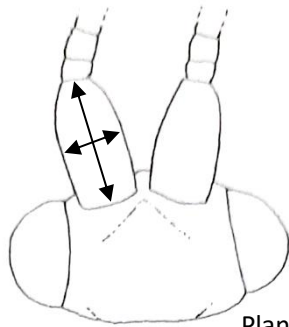
Cunctochrysa albolineta

Cunctochrysa cosmia

Antennal scape



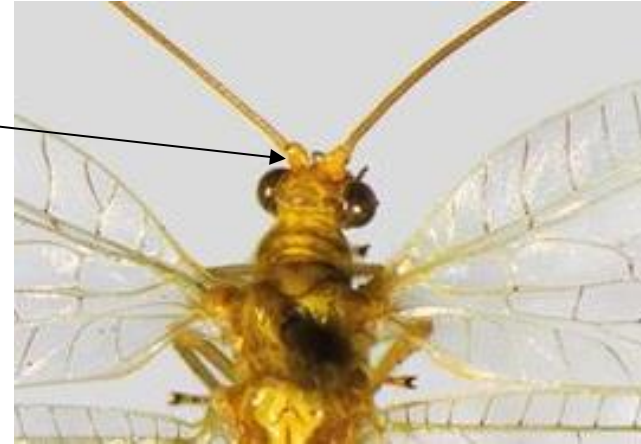
T.Rintala & T. Lehto (CC BY 4.0)



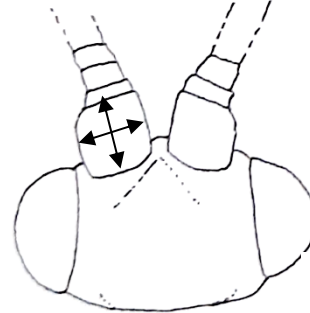
Plant, 1997

Twice as long as wide

Nineta vittata



T.Rintala & T. Lehto (CC BY 4.0)



Plant, 1997

More or less square Slightly longer than wide

Nineta flava

Nineta inpunctata

Apertochrysa flavifrons

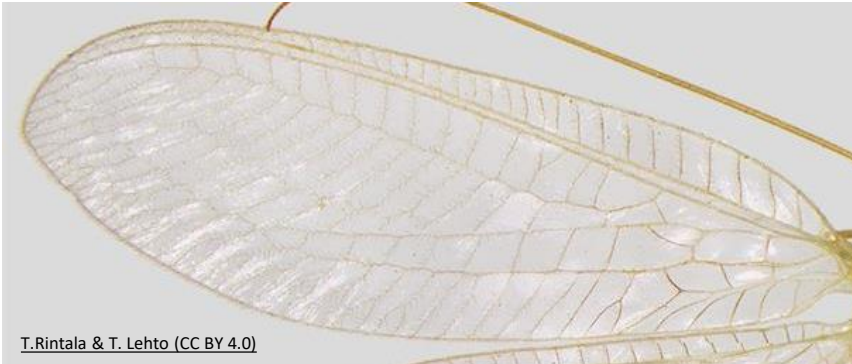
Chrysopidia ciliata

Cunctochrysa albolineata

Cunctochrysa bellifontensis

Wing length and costal margin

Straight



Forewing *less than* 16 mm
Costal margin straight *NEVER* concave

Apertochrysa flavifrons
Cunctochrysa cosmia
Cunctochrysa albolineata
Chrysopidia ciliata

Concave or straight



Forewing 16 mm or *greater*
Costal margin straight or concave

Nineta flava
Nineta pallida
Nineta inpunctata

Colour of basal costal crossvein and SCRCV



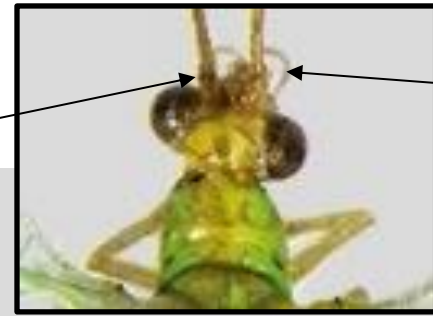
Cunctochrysa albolineata
Chrysopidia ciliata



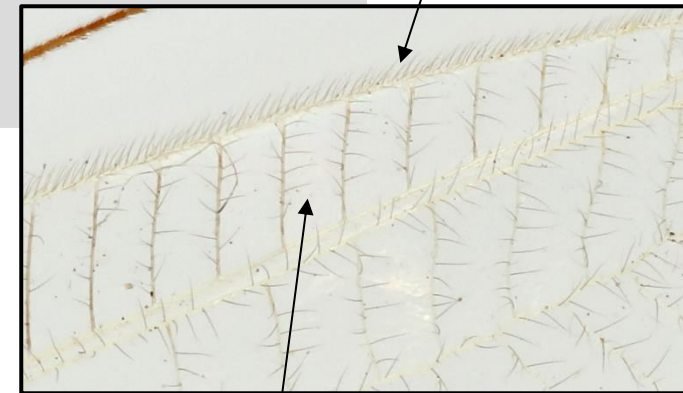
Apertochrysa flavifrons
Cunctochrysa cosmia

Chrysopidia ciliata

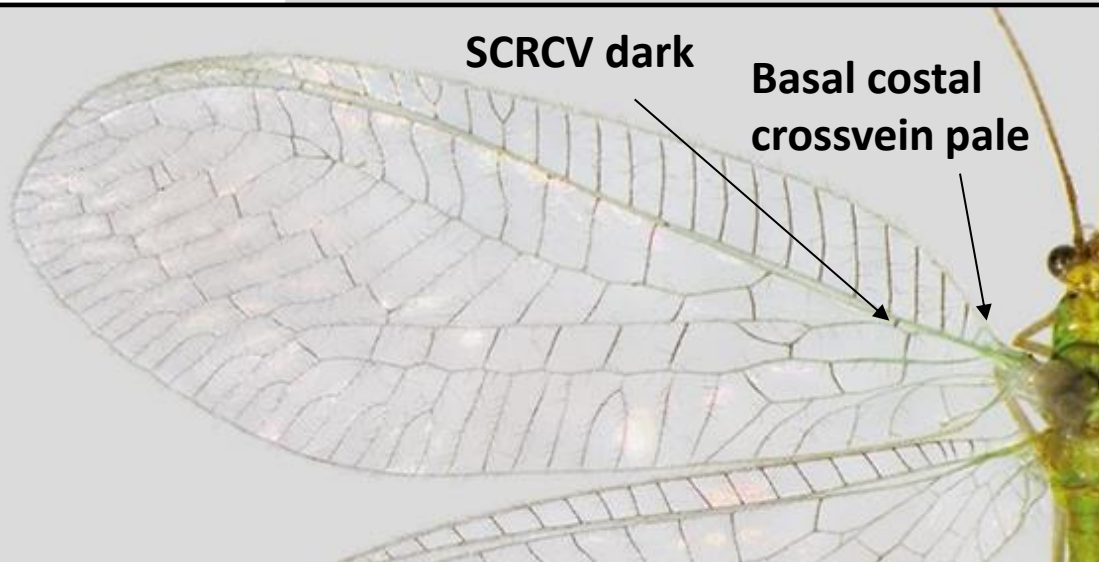
T. Rintala & T. Lehto (CC BY 4.0)



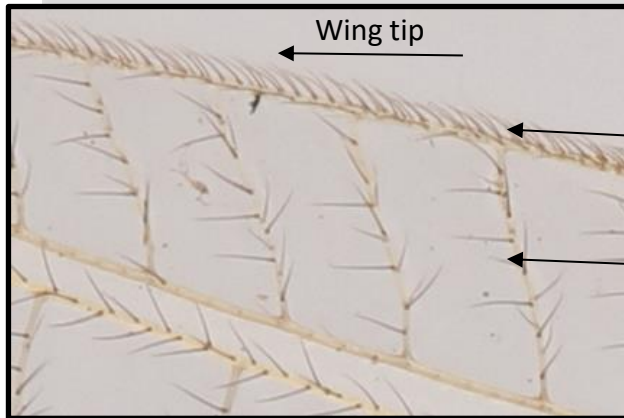
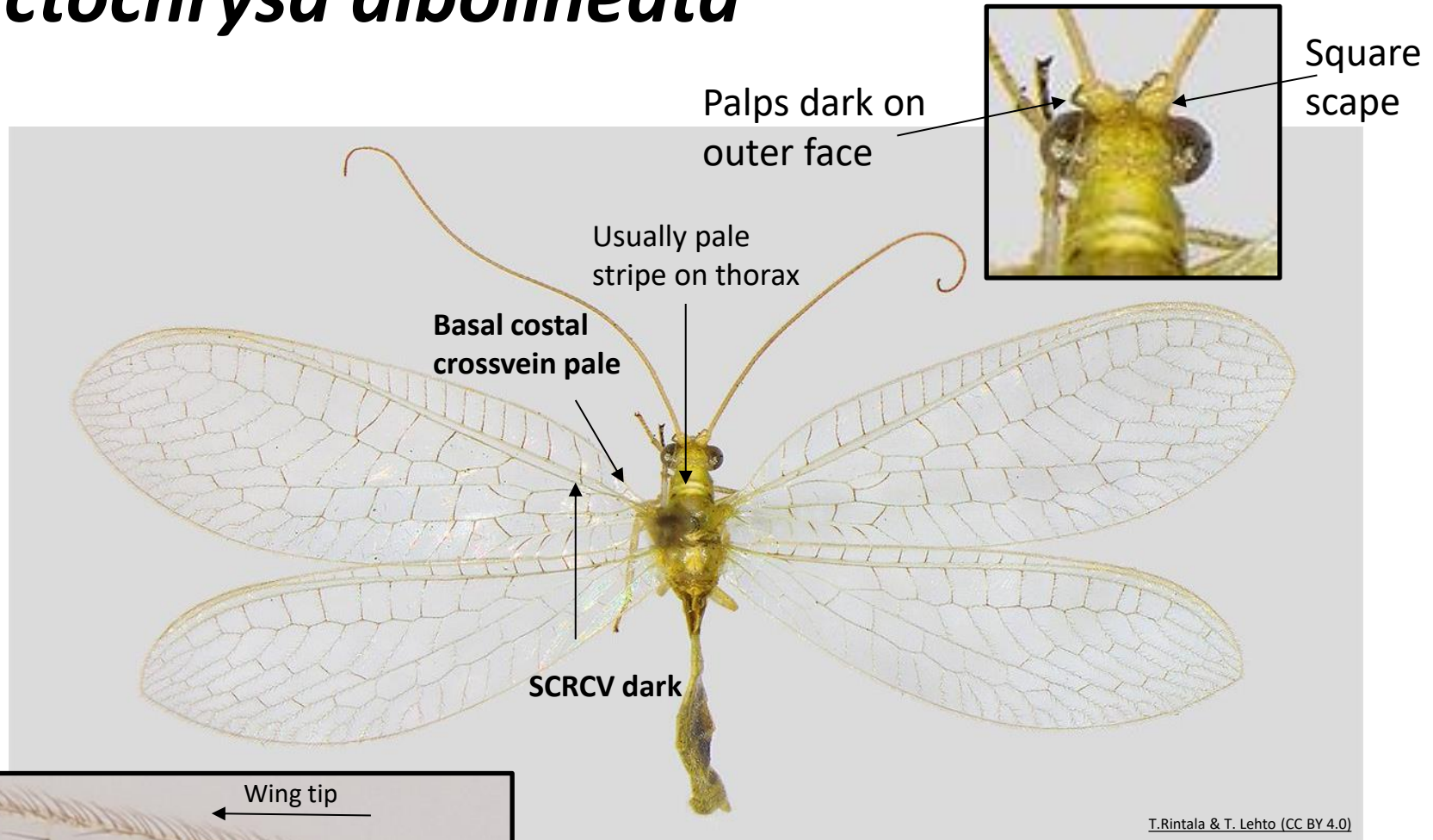
Hairs on Costa
7 or 8 times
long as width
of Costa



Hairs on Costal crossveins often
longer than $\frac{1}{2}$ width of cell pointing
in both directions, overlapping



Cunctochrysa albolineata



Hairs on Costa 2 or 3 times long as width of Costa

Hairs on costal veins pointing towards wing tip
Around 1/3 length of cell

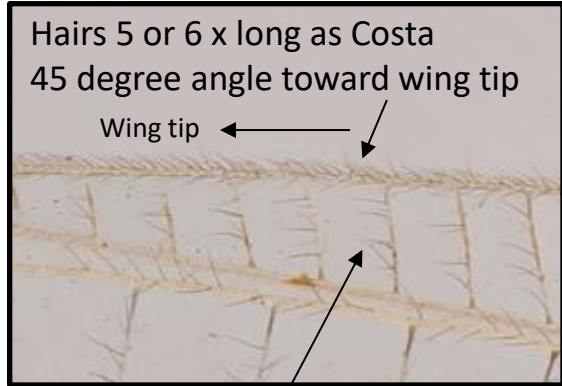
Colour of basal costal crossvein and SCRCV



Apertochrysa flavifrons
Cunctochrysa cosmia

Apertochrysa flavifrons (= *Dichochrysa flavifrons*)

T. Rintala & T. Lehto (CC BY 4.0)



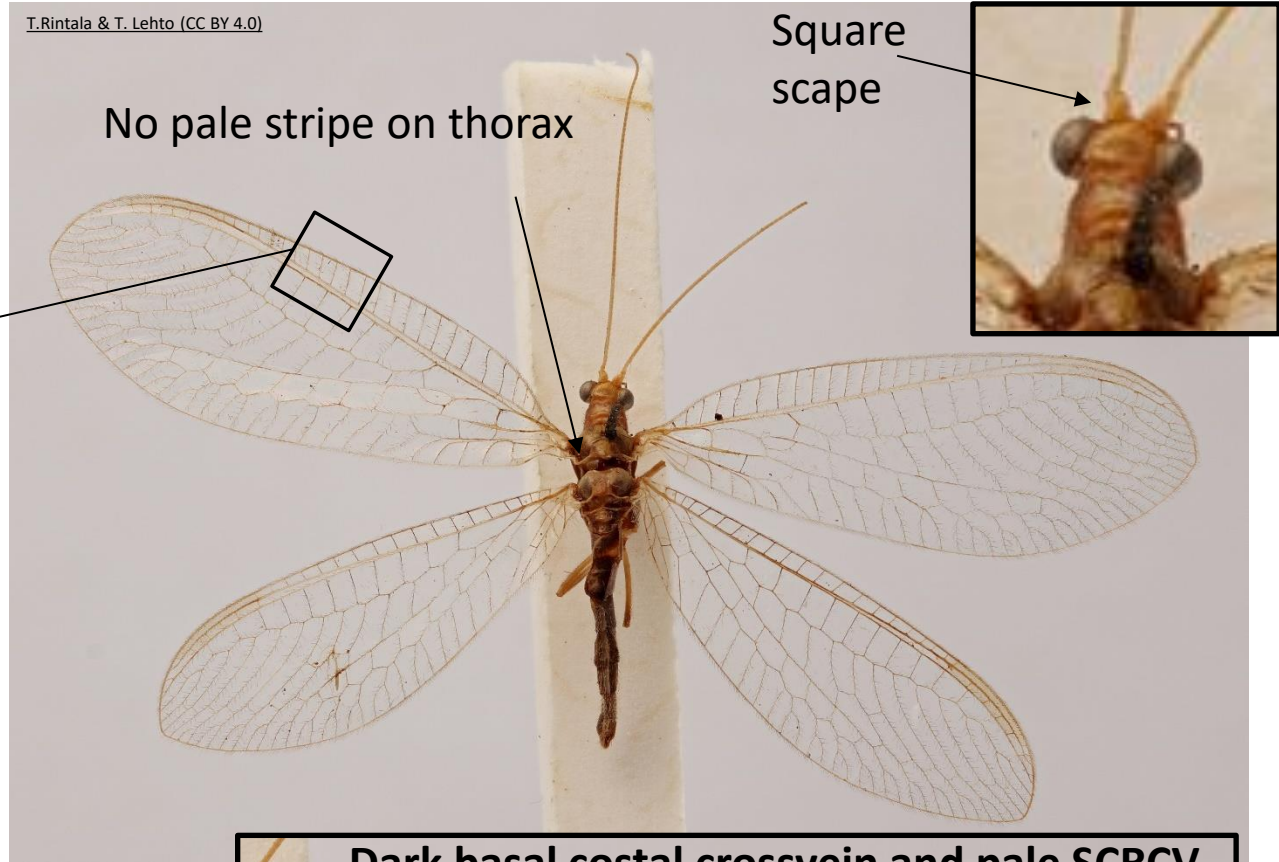
Hairs on costal crossveins
do not quite reach half way
Most point towards wing tip

This text describes the characteristics of the hairs on the costal crossveins, noting that they do not reach halfway and most point towards the wing tip.

Palps ringed black



Red-Brown spots on side of face



Dark spot
on base
of Costa

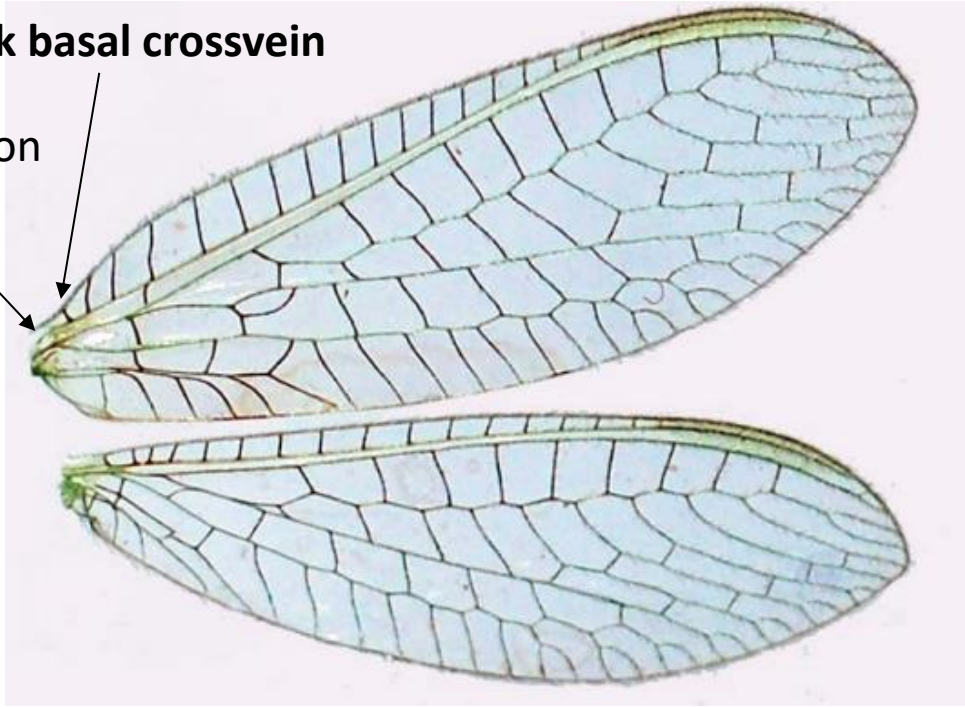
Dark basal costal crossvein and pale SCRCV

This image shows a close-up of the wing base, highlighting the dark spot on the base of the Costa and the dark basal costal crossvein and pale SCRCV.

Cunctochrysa cosmia

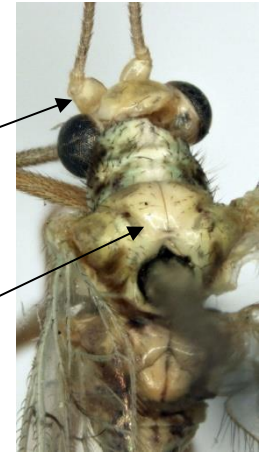
Dark basal crossvein

No dark spot on base of Costa



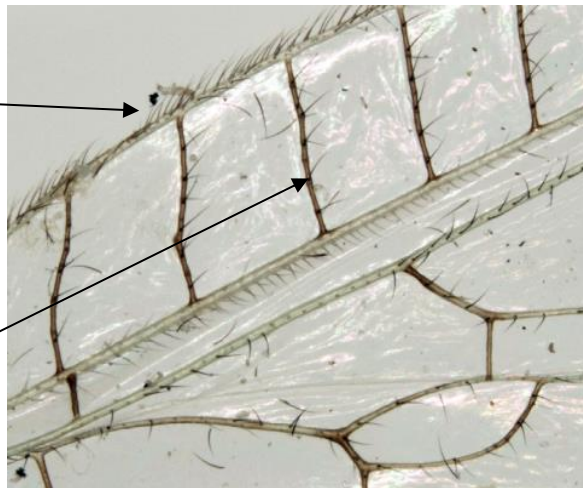
Square scape

Pale stripe on thorax



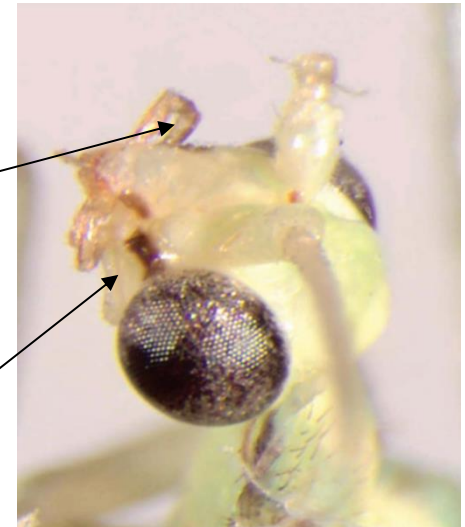
Hairs on Costa
2 or 3x as long
as Costa

Hairs on costal
crossveins about
1/3 length of cell



Palps pale
on inner face

Black face
markings



Concave or straight



T.Rintala & T. Lehto (CC BY 4.0)

Forewing 16 mm or greater
Costal margin straight or concave

Nineta flava

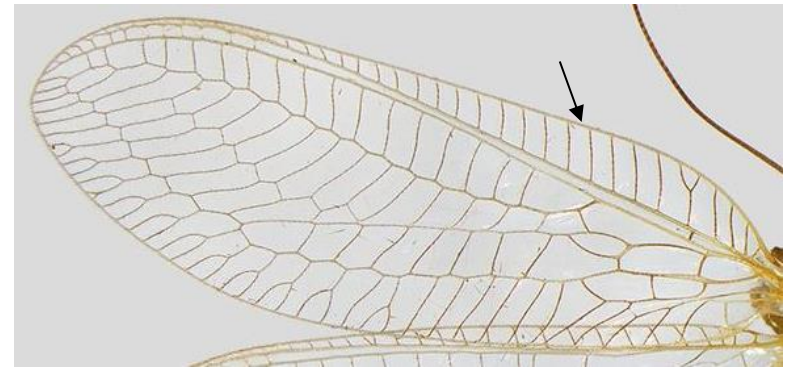
Nineta pallida

Nineta inpunctata

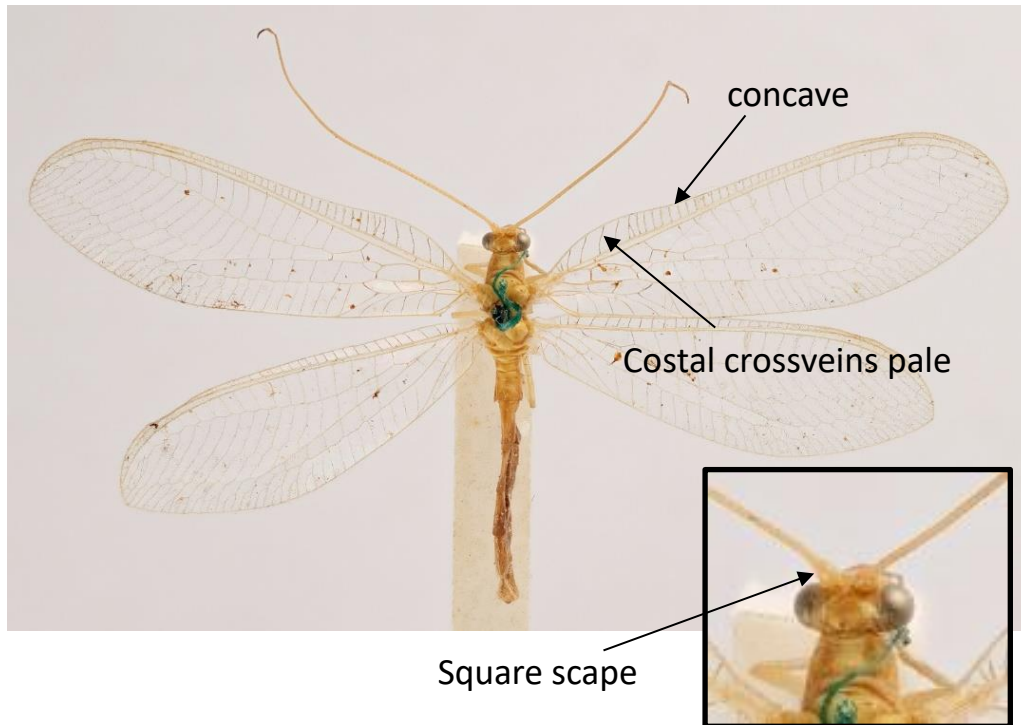
Concave



Straight

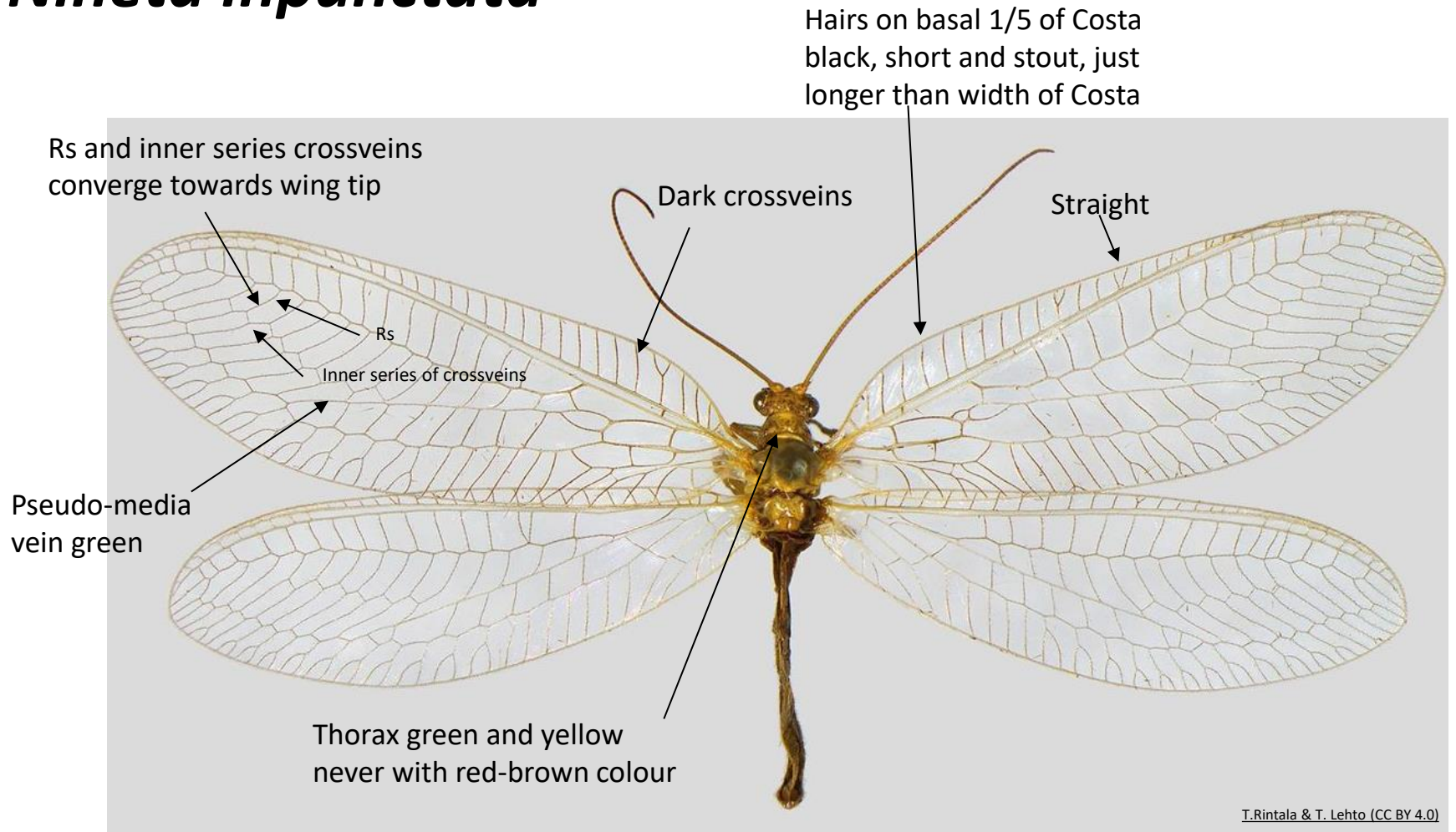


Nineta flava



Nineta inpunctata
Nineta pallida

Nineta inpunctata



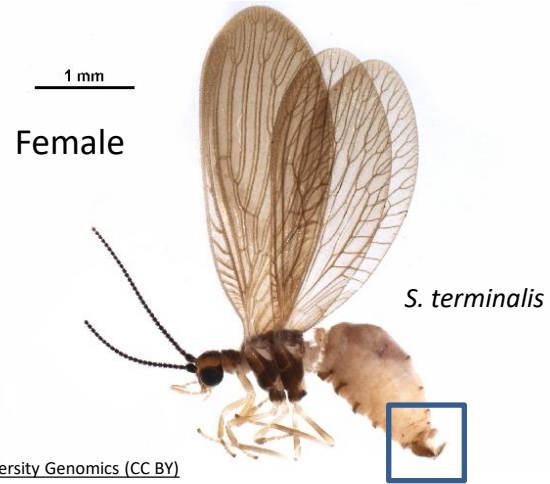
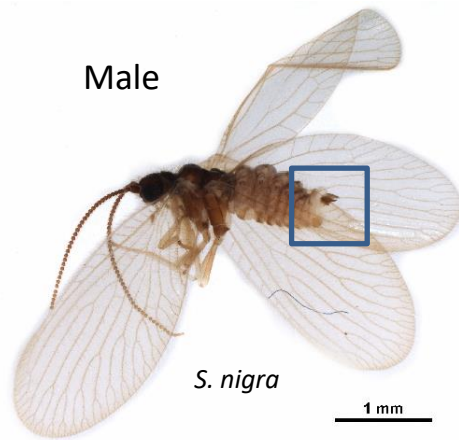
Only one British record from Essex (1996)
Retain a voucher specimen

Sisyridae (Sponge Flies)

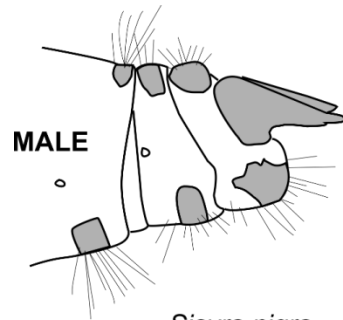
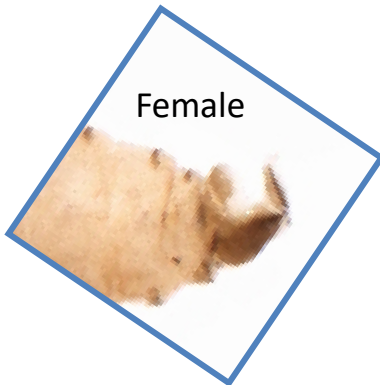
- Can be preserved pinned or in alcohol
- Can identify from photographs but genitalia is important for confirmation



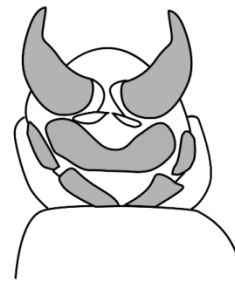
Sisyridae genitalia



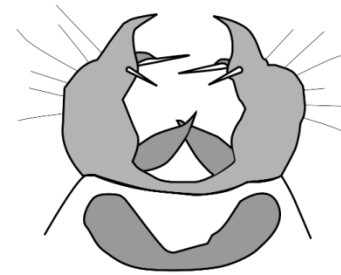
CBG Photography Group, Centre for Biodiversity Genomics (CC BY)



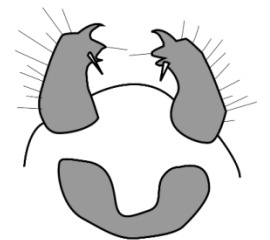
Sisyra nigra



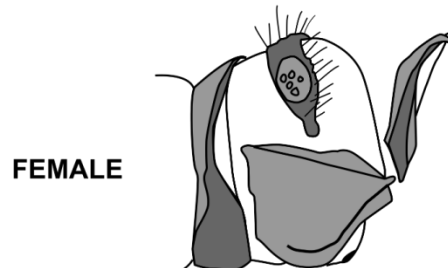
Sisyra nigra



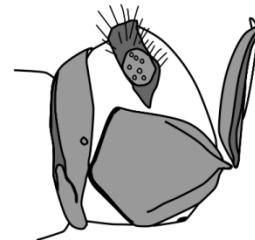
Sisyra terminalis



Sisyra dalii



Sisyra nigra



Sisyra terminalis



Sisyra dalii

Mecoptera (Scorpionflies)

- Can be preserved pinned or in alcohol
- Males can be identified in the field from genital capsule
- Females need to be dissected to identify species confidently

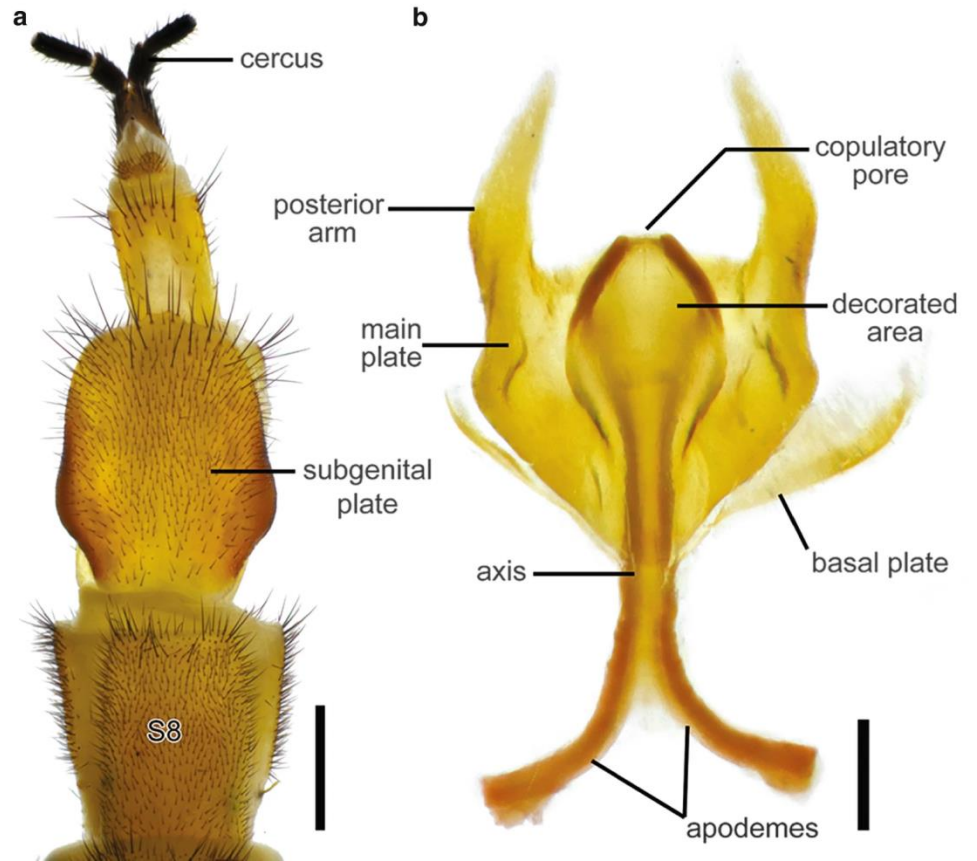


Male



Female

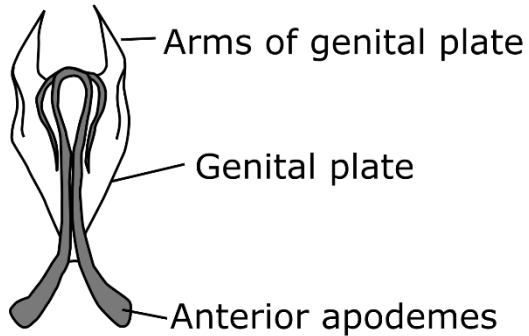
Mecoptera Females



Wang & Hua, 2022

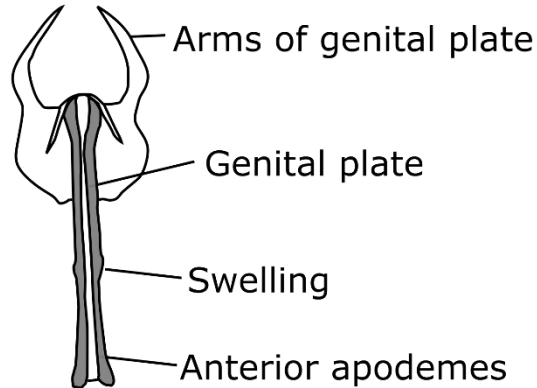
Panorpa communis

Panorpa communis



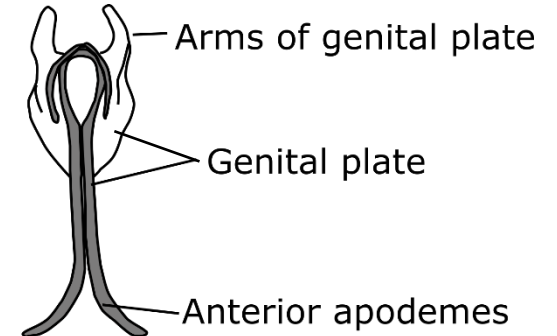
- Arms of genital plate taper to a point
- Anterior apodemes, short or flattened, or broad diverging most of length

Panorpa germanica



- Arms as long as genital plate
- Long straight apodemes with swelling

Panorpa cognata



- Arms blunt and shorter than genital plate
- Apodemes long, narrow, diverging towards tip



British Isles Lacewing and Allies Recording Scheme

Submitting your records

Email them to:

LacewingRS@gmail.com or colinwplant@gmail.com

Upload records onto iRecord



British Lacewing and Allies Recording Scheme Website:



<https://www.laars.jamesjepson.com>

Useful websites:

Neuropterida of Norway and the Nordic Countries:

https://www.artsdatabanken.no/Pages/223140/Nettvinger__mudderfluer_og_kamelhalsfluer_i

Apple Wildlife:

<http://applewildlife.co.uk/photos-lacewings/lacewings.php>

Lacewing Digital Library:

<https://lacewing.tamu.edu/>