KEYS FOR THE IDENTIFICATION OF BRYOPHYTES OCCURRING IN HUNGARY

Peter Erzberger

Belziger Str. 37, D 10823 Berlin, Germany, E-mail: erzberger.peter@gmail.com

Abstract: Keys for the identification of all bryophytes presently known to occur in Hungary are presented. The three groups: Hornworts (2 taxa), Liverworts (149 taxa), and Mosses (541 taxa) are treated separately. Bryophyte identification using these keys proceeds in two steps: 1. Artificial keys to the genera, 2. Keys from genera to species, arranged systematically according to recent taxonomy. Each species of the Hungarian bryophyte flora is assigned to one of six frequency classes (very common: cc, common: c, widespread: w, rare: r, very rare: rr, not seen: n.s.). A glossary explaining the technical terms used in the keys and an index of genera are included.

Keywords: Genera, species, glossary, mosses, liverworts, hornworts

INTRODUCTION

Identification of Hungarian bryophytes up to now had to rely on either keys in Hungarian published 68 and 38 years ago, respectively, long out of print (Boros 1953, Orbán and Vajda 1983), and out-of-date in many respects, or on keys written in various languages for other countries or regions (e.g. for liverworts and hornworts: Smith (1991), Paton (1999), Damsholt (2002), Schumacker and Váňa (2005), Frey et al. (2006), Casas et al. (2009), Atherton et al. (2010) in English, Gradstein and van Melick (1996), Siebel and During (2006) in Dutch, Müller (1905–1916, 1951–1958), Frahm and Frey (1992, 2004), Frey et al. (1995), Nebel and Philippi (2005) in German; for mosses: Smith (1978, 2004), Casas et al. (2006), Frey et al. (2006), Atherton et al. (2010) in English, Touw and Rubers (1989), Siebel and During (2006) in Dutch, Frahm and Frey (1992, 2004), Frey et al. (1995), Nebel and



Philippi (2000, 2001) in German, Brugués et al. (eds.) (2007), Brugués and Guerra (eds.) (2015), Guerra et al. (eds.) (2006, 2010, 2014, 2016) in Spanish, to name just a few). None of these treatments by itself allows the identification of all bryophytes presently known in the Hungarian bryophyte flora, except Frey et al. (2006). Therefore, time is overdue to present an accurate and updated tool for naming all bryophyte taxa of Hungary, including the many that have been discovered as new members of the Hungarian bryoflora in recent years. This need became particularly urgent during the first stages of the Hungarian bryophyte recording project (Erzberger 2012, Erzberger and Németh 2016, Erzberger 2020) when new contributors to the project had to be guided in improving their identifications skills. To remedy this, the author then compiled a preliminary version of identification keys, which was circulated among the participants of the recording project and continuously improved according to the accumulating experience.



Figure 1. *Riccia frostii* Austin, Hungary, Győr-Moson-Sopron County, Vének, bank of the Danube, 02.10.2015 photo by Csaba Németh

Taxonomy and nomenclature closely follow the most recent European and Hungarian checklists (Hodgetts *et al.* 2020, Erzberger and Papp 2020). Here authorities to all names can be found; they have been deliberately omitted in this treatment. To

facilitate the use of older works, the names used in older checklists (Erzberger and Papp 2004, Papp *et al.* 2010) are included as synonyms. To give an overwiew of the underlying taxonomy, a conspectus of classification has been included. A glossary explaining all technical terms used in the keys has also been compiled.

As a consequence of recent taxonomic developments, mainly based on molecular evidence, many species have changed their generic names and many genera have been transferred to different families. Thus the conventional approach of three steps in bryophyte identification – keys to families, keys to genera of each family, keys to species of each genus – is no longer practical, since many families lack easily observable morphological defining characters. I have therefore followed Guerra *et al.* (eds.) (2016) and Casas *et al.* (2006, 2009) in first giving a key to genera and then within each genus (containing more than one species in Hungary) a key to species.

However, for families with a \pm clear-cut morphological definition, keys are provided directly to all or nearly all species of the family (*Table 1*).

Table 1. Families for which a direct key to species is provided

Table 1. Families for which a direct key to species is provided			
Liverworts	page	Mosses	page
Aneuraceae	109	Bartramiaceae	189
Calypogeiaceae	98	Bryaceae	190
Cephaloziaceae	86	Dicranaceae	139
Lejeuneaceae	108	Ditrichaceae s.l. incl. Saelania,	143
		Flexitrichum	
Lophocoleaceae	104	Funariaceae	131
Pelliaceae	111	Grimmiaceae	175
Plagiochilaceae	106	Polytrichaceae	126
Scapaniaceae excluding	95	Rhabdoweisiaceae	142
Schistochilopsis		Seligeriaceae	174

In some cases these keys are additional to the keys from genus to species, but in others there are no separate keys from genus to species. This applies in particular where genera are keyed out collectively (often along the delimitations of taxa formerly in use), not separately, because their morphological delimitation is not useful for a simple key. Some of the newly established genera have been included in collective keys to species (*Table 2*).

Table 2. Groups of genera treated collectively in keys to species

	Liverworts	page
Barbilophozia group	Barbilophozia, Neoorthocaulis	85
Jungermanniaceae	Endogemma, Jungermannia, Liochlaena,	101
group	Nardia, Solenostoma, Syzygiella	
Lophoziaceae group	Barbilophozia, Isopaches, Lophozia,	89
	Lophoziopsis, Mesoptychia, Neoorthocaulis,	
	Obtusifolium, Schistochilopsis, Trilophozia,	
	Tritomaria	
Marchantiales pp.	Asterella, Clevea, Conocephalum, Lunularia,	112
	Mannia, Marchantia, Reboulia	
Riccia group	Oxymitra, Riccia, Ricciocarpos	115
	Mosses	page
Amblystegium group	Amblystegium, Hygroamblystegium,	216
	Pseudoamblystegium, Pseudocampylium,	
	Serpoleskea	
Anomodon group	Anomodon, Claopodium, Pseudanomodon	230
Bryum s.l.	Bryum, Imbribryum, Ptychostomum	190
Brachythecium s.l.	Brachytheciastrum, Brachythecium,	223
	Sciuro-hypnum	
Calliergon group	Calliergon, Straminergon	218
Gymnostomum group	Gymnostomum, Gyroweisia	170
Hypnum s.l.	Buckia, Hypnum	227
Mnium group	Mnium, Plagiomnium, Rhizomnium	201
Neckera s.l.	Alleniella, Exsertotheca, Neckera	228
Pottia s.l.	Hennediella, Microbryum (excl. M.	153
	curvicollum, M. floerkeanum), Tortula caucasica, T. lindbergii, T. protobryoides,	
	T. truncata	
Tortula s.l.	Hilpertia, Syntrichia, Tortula (excl. T.acaulon,	160
	T. caucasica, T. lindbergii, T. protobryoides,	
	T. truncata)	

This approach, although at first glance seeming to lack systematic rigour, should provide the possibility to enter the process of identification at various taxonomic levels (family, group of morphologically similar genera, genus) and thus enable a comparison among taxa that could easily be confounded, although in some cases not related taxonomically. A similar purpose is intended by various notes mentioning differences between related or unrelated taxa.

The keys have been adapted from the references mentioned above and others (e.g. Watson 1981), and from additional treatments of special groups which are indicated at the families or

genera, including personal experience of the author. In general, only taxa from the most recent Hungarian checklist (Erzberger and Papp 2020) have been considered, plus three species added to the Hungarian bryoflora after the publication of the checklist (Hydrogonium croceum, Orthothecium rufescens, Rhytidiadelphus loreus), their names are printed in **bold**, but some taxa that might be expected (or have been doubtfully recorded) have in a few cases been included (printed in normal case).

As an additional information the frequency of recent occurrences of taxa (based on records since 1974, i.e. after the era of Á. Boros) is included in the following categories: very common: cc, common: c, widespread: w, rare: r, very rare: rr, not seen: n.s. (i.e. no record after 1973). These estimates are based on a preliminary evaluation of the recording project (Erzberger and Németh 2016, Erzberger 2020) which will be published elsewhere, and their accuracy should not be overestimated, since coverage of the country is far from complete; in particular the lowlands are sorrowfully under-represented. The frequency may help mainly beginners to check the result of their identification, excluding very rare and rare species, which are unlikely to be found without experience.

Acknowledgements – I am indebted to Péter Szűcs for editing the keys, to Csaba Németh for providing the photograph of *Riccia frostii* (Fig. 1.) and to the participants of the recording project for feedback on the preliminary keys and their contributions to the dataset. Special thanks are due to Christian Berg (Graz), Frank Müller (Dresden) and Tamás Pócs (Eger) for their constructive criticism of the manuscript resulting in many essential improvements.

REFERENCES

- ATHERTON, I., BOSANQUET, S.D.S. & LAWLEY, M. (eds.) (2010). *Mosses and Liverworts of Britain and Ireland*. A Field Guide. British Bryological Society, Plymouth, 848 pp.
- Bakalin, V.A. (2016). Notes on *Lophozia* VIII. The Lectotypification of *Lophozia longiflora* (Nees) Schiffn. (Lophoziaceae, Hepaticae). *Herzogia* **29**: 635–643. https://doi.org/10.13158/heia.29.2.2016.635
- BARÁTH, K. & ERZBERGER, P. (2019). *Heterocladium heteropterum*, a new member of the Hungarian bryophyte flora. *Studia Botanica Hungarica* **50**(2): 323–329. https://doi.org/10.17110/StudBot.2019.50.2.323
- BARÁTH, K., ERZBERGER, P., KOVÁCS, A. & PAPP, B. (2016). *Heterocladium dimorphum* (Brid.) Schimp. (Heterocladiaceae) an old element of the Hungarian bryophyte flora rediscovered. *Studia Botanica Hungarica* **47**: 269–278.

- http://dx.doi.org/10.17110/StudBot.2016.47.2.269
- Bergamini, A. (2001). Provisorischer Schlüssel zur Unterscheidung steriler Philonotis-Proben. http://www.bryolich.ch/pdfs/Philonotis_key.pdf
- BLOCKEEL, T.L. (2017). The *Ulota crispa* group in Britain and Ireland, with notes on other species of the genus. *Field Bryology* **117**: 8–19.
- BLOCKEEL, T.L., OCHYRA, R. & Gos, L. (2000). *Seligeria campylopoda* Kindb. in the British Isles. *Journal of Bryology* **22**: 29–33. https://doi.org/10.1179/jbr.2000.22.1.29
- BLOM, H. (1996). *A revision of the Schistidium apocarpum complex in Norway and Sweden*. Bryophytorum Bibliotheca 49, I. Cramer, Borntraeger, Berlin, 333 pp.
- Boros, Á. (1953). *Magyarország mohái (Bryophyta Hungariae*). Akadémiai Kiadó, Budapest, 360 pp.
- Boros, Á. (1968). *Bryogeographie und Bryoflora Ungarns*. Akadémiai Kiadó, Budapest, 466 pp.
- Brugués, M., Cros, R.M. & Guerra, J. (eds.) (2007). Flora Briofítica Ibérica. Vol. I, Sphagnales, Andreaeales, Polytrichales, Tetraphidales, Buxbaumiales, Diphysciales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 183 pp.
- Brugués, M. & Guerra, J. (eds.) (2015). Flora Briofítica Ibérica. Vol. II, Archidiales, Dicranales, Fissidentales, Seligeriales, Grimmiales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 355 pp.
- CAPARRÓS, R., LARA, F., DRAPER, I., MAZIMPAKA, V. & GARILLETI, R. (2016). Integrative taxonomy sheds light on an old problem: the *Ulota crispa* complex (Orthotrichaceae, Musci). *Botanical Journal of the Linnean Society* **180**: 427–451. https://doi.org/10.1111/boj.12397
- Casas, C., Brugués, M., Cros, R.M. & Sérgio, C. (2006). *Handbook of Mosses of the Iberian Peninsula and the Balearic Islands*. Institut d'Estudis Catalans, Barcelona, 349 pp.
- CASAS, C., BRUGUÉS, M., CROS, R.M., SÉRGIO, C. & INFANTE, M. (2009). *Handbook of Liverworts and Hornworts of the Iberian Peninsula and the Balearic Islands*. Institut d'Estudis Catalans, Barcelona, 177 pp.
- Caspari, S., Dürhammer, O., Sauer, M. & Schmidt, C. (2018). Rote Liste und Gesamtartenliste der Moose (*Anthocerotophyta, Marchantiophyta* und *Bryophyta*) Deutschlands. In: Metzing, D., Hofbauer, N., Ludwig, G. & Matzke-Hajek, G. (eds.): *Rote Liste gefährdeter Tiere, Pflanzen und Pilze Deutschlands. Band 7: Pflanzen*. Landwirtschaftsverlag, Münster. *Naturschutz und Biologische Vielfalt* 70(7): 361–489.
- CSIKY, J., ERZBERGER, P., KOVÁCS, D. & DEME, J. (2014). *Campylopus pyriformis* (Schultz) Brid. in the Western Mecsek Mts. (South Transdanubia, Hungary) / *Campylopus pyriformis* (Schultz) Brid. a Ny-Mecsekben. *Kitaibelia* 19: 366–367.
- CSIKY, J., ERZBERGER, P., KOVÁCS, D. & DEME, J. (2015). *Campylopus flexuosus* (Hedw.) Brid. in the Western Mecsek Mts. (South Transdanubia, Hungary). / *Campylopus flexuosus* (Hedw.) Brid. a Nyugat-Mecsekben. *Kitaibelia* **20**(1): 28–37. https://doi.org/10.17542/kit.20.28
- DAMSHOLT, K. (2002). *Illustrated Flora of Nordic Liverworts and Hornworts*. Nordic Bryological Society, Lund. 837 pp.
- DEMARET, F. (1993). Bryum Hedw. In: DE SLOOVER, J.-L., DEMARET, F., DE ZUTTERE, PH. & ARTS, TH. (eds.) Flore Générale de Belgique Bryophytes Vol III. Fascicule 2.

- Ministère de l' Agriculture, Jardin Botanique National de Belgique, Meise, pp. 152–258.
- Deme, J., Erzberger, P., Kovács, D., Tóth, I. Zs. & Csiky, J. (2020). *Buxbaumia viridis* (Moug. ex Lam. & DC.) Brid. ex Moug. & Nestl. in Hungary predominantly terricolous and found in managed forests. *Cryptogamie, Bryologie* **41**: 89–103. https://doi.org/10.5252/cryptogamie-bryologie2020v41a8
- Dierssen, K. (1996). Bestimmungsschlüssel der Torfmoose in Norddeutschland. *Mitt. Arbeitsgem. Geobot. Schleswig-Holstein & Hamburg* **50**: 1–86.
- Düll-Hermanns, I. (1981). Spezielle Untersuchungen zur modernen Taxonomie von *Thuidium abietinum* und der Varietät *hystricosum*. *Journal of Bryology* **11**: 467–487.
- ELLIS, L.T. & PRICE, M.J. (2015). Review of the type specimens of species described by J. Hedwig in *Phascum* Hedw. (Pottiaceae). *Journal of Bryology* **37**: 23–41. https://doi.org/10.1179/1743282014Y.0000000116
- ERZBERGER, P. (1996). Zur Verbreitung von *Hedwigia stellata* in Europa. *Herzogia* **12**: 221–238.
- Erzberger, P. (1998). *Tortula brevissima* Schiffn. eine für die Flora Ungarns neue Moosart. *Botanikai Közlemények* **85**: 63–72.
- ERZBERGER, P. (1999). Distribution of *Dicranum viride* and *Dicranum tauricum* in Hungary. *Studia Botanica Hungarica* **29**: 35–47.
- ERZBERGER, P. (2001). *Ditrichum crispatissimum* (Muell. Hal.) Paris, a new species of the Hungarian bryoflora, and *Ditrichum flexicaule* (Schleich. ex Schwaegr.) Hampe in Hungary. *Studia Botanica Hungarica* **32**: 87–105.
- ERZBERGER, P. (2002). Funaria muhlenbergii and Funaria pulchella (Funariaceae, Bryophyta) in Hungary. Studia Botanica Hungarica 33: 47–63.
- ERZBERGER, P. (2005). The bulbilliferous species of *Pohlia* (Bryaceae, Musci) in Hungary. *Studia Botanica Hungarica* **36**: 67–75.
- ERZBERGER, P. (2009). The genera *Grimmia* and *Coscinodon* (Grimmiaceae, Musci) in Hungary. *Studia Botanica Hungarica* **40**: 37–124.
- ERZBERGER, P. (2012). *Project plan: Bryophyte mapping of Hungary*. Abstracts of 8th ECCB Conference in Budapest, April 2012, p. 6.
- Erzberger, P. (2016). The genus *Fissidens* Hedw. (Bryophyta) in Hungary. *Studia Botanica Hungarica* **47**: 41–139.
 - http://dx.doi.org/10.17110/StudBot.2016.47.1.41
- ERZBERGER, P. (2020). Bryophyte recording in Hungary in the 21st century. *Field Bryology* **123**: 21–33.
- ERZBERGER, P. & BARÁTH, K. (2017). *Plagiothecium latebricola* Schimp. a new member of the Hungarian bryoflora. *Studia Botanica Hungarica* **48**(2): 189–197. https://doi.org/10.17110/StudBot.2017.48.2.189
- ERZBERGER, P., BEDNAREK-OCHYRA, H. & OCHYRA, R. (2016). Grimmiaceae subfam. Racomitrioideae (Bryophyta) in Hungary. *Polish Botanical Journal* **61**(1): 23–51. https://doi.org/10.1515/pbj-2016-0015
- ERZBERGER, P. & NÉMETH, Cs. (2014). *Campylopus flexuosus* (Hedw.) Brid.: a moss new to the Hungarian bryophyte flora. / Új faj Magyarország mohaflórájában: *Campylopus flexuosus* (Hedw.) Brid. *Kitaibelia* **19**: 22–28.
- ERZBERGER, P. & NÉMETH, Cs. (2016). *Bryophyte recording in Hungary results 2012–2015.* 11th International Conference "Advances in research on the flora and

- vegetation of the Carpato-Pannonian region", Budapest, 12–14 February 2016 (lecture). Book of Abstracts, p. 17.
- ERZBERGER, P., NÉMETH, Cs., DEME, J. & CSIKY, J. (2018). Stomatal anatomy allows clarification of historical collections of *Buxbaumia* Hedw. in Hungary. *Studia Botanica Hungarica* **49**(1): 71–82. https://doi.org/10.17110/StudBot.2018.49.1.71
- ERZBERGER, P., NÉMETH, CS., SAUER, M., NAGY, J. & PAPP, B. (2020). *Plagiothecium platyphyllum* Moenk. a rare species in Hungary. *Studia Botanica Hungarica* **51**(1): 25–40. https://doi.org/10.17110/StudBot.2020.51.1.25
- ERZBERGER, P. & PAPP, B. (2000). *Orthotrichum sprucei* discovered in continental Central Europe. *Herzogia* **14**: 213–215.
- ERZBERGER, P. & PAPP, B. (2004). Annotated checklist of Hungarian bryophytes. *Studia Botanica Hungarica* **35**: 91–149.
- ERZBERGER, P. & PAPP, B. (2018). *Tortella fasciculata* and *T. pseudofragilis* (Pottiaceae, Bryophyta) in Hungary. *Studia Botanica Hungarica* **49**(2): 39–48. https://doi.org/10.17110/StudBot.2018.49.2.39
- Erzberger, P. & Papp, B. (2020). The checklist of Hungarian bryophytes second update. *Studia Botanica Hungarica* **51**(2): 11–76. https://doi.org/10.17110/StudBot.2020.51.2.11
- ERZBERGER, P. & SCHRÖDER, W. (2008). The genus *Schistidium* (Grimmiaceae, Musci) in Hungary. *Studia Botanica Hungarica* **39**: 27–88.
- Erzberger, P. & Schröder, W. (2013). The genus *Bryum* (Bryaceae, Musci) in Hungary. *Studia Botanica Hungarica* **44**: 5–192.
- Frahm, J.-P. & Ahmed, J. (2004). *Barbula sardoa* (Schimp.) J.-P. Frahm, a new name for *Barbula convoluta* Hedw. var. *commutata* (Jur.) Husn. *Journal of Bryology* **26**(1): 29–35.
- Frahm, J.-P. & Frey, W. (1992). Moosflora. 3rd. ed. Ulmer, Stuttgart, 528 pp.
- Frahm, J.-P. & Frey, W. (2004). *Moosflora*. 4th. ed. Ulmer, Stuttgart, 538 pp.
- FREY, W., FRAHM, J.-P., FISCHER, E. & LOBIN, W. (1995). *Die Moos- und Farnpflanzen Europas*. Fischer, Stuttgart, Jena, New York, 426 pp.
- FREY, W., FRAHM, J.-P., FISCHER, E. & LOBIN, W. (2006). *The Liverworts, Mosses and Ferns of Europe.* English edition revised and edited by T.L.Blockeel. Harley Books, Colchester, 512 pp.
- Gallego, M.T. (2005). A taxonomic study of the genus *Syntrichia* Brid. (Pottiaceae, Musci) in the Mediterranean region and Macaronesia. *Journal of Hattori Botanical Laboratory* **98**: 47–122.
- Gallego, M.T., Hugonnot, V. & Cano, M.J. (2018). Taxonomic resurrection of an awnless variety of *Syntrichia ruralis* and comparison with other European muticous taxa in this genus. *Journal of Bryology* **40**(3): 244–250. https://doi.org/10.1080/03736687.2018.1468971
- Godfrey, M. & Hill, M. (2012). Sphagnum Workshop, November 2011. Field Bryology 106: 69.
- Gradstein, S.R. & van Melick, H.M.H. (1996). *De Nederlandse Levermossen en Hauwmossen. Flora en verspreidingsatlas van de Nederlandse Hepaticae en Anthocerotae*. Stichting Uitgeverij van de Koninklijke Nederlandse Natuurhistorische Vereniging, Utrecht, 366 pp.

- GUERRA, J., BRUGUÉS, M., CANO, M.J. & CROS, R.M. (eds.) (2010). Flora Briofítica Ibérica. Vol. IV, Funariales, Splachnales, Bryales, Timmiales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 317 pp.
- GUERRA, J., CANO, M.J. & BRUGUÉS, M. (eds.) (2014). Flora Briofítica Ibérica. Vol. V, Orthotrichales, Hedwigiales, Leucodontales, Hookeriales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 261 pp.
- Guerra, J., Cano, M.J. & Brugués, M. (eds.) (2016). Flora Briofítica Ibérica. Vol. VI, Hypnales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 463 pp.
- Guerra, J., Cano, M.J. & Ros, R.M. (eds.) (2006). Flora Briofítica Ibérica. Vol. III, Pottiales, Encalyptales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 305 pp.
- HASSEL, K., KYRKJEEIDE, M.O., YOUSEFI, N., PRESTØ, T., STENØIEN, H.K., SHAW, J.A. & FLATBERG, K.I. (2018). *Sphagnum divinum* (sp. nov.) and *S. medium* Limpr. and their relationship to *S. magellanicum* Brid. *Journal of Bryology* **40**: 197–222. https://doi.org/10.1080/03736687.2018.1474424
- HEDENÄS, L. (1994). The *Hedwigia ciliata* complex in Sweden, with notes on the occurrence of taxa in Fennoscandia. *Journal of Bryology* **18**: 139–157.
- HEDENÄS, L. & BISANG, I. (2002). *Drepanocladus sordidus* und *D. stagnatus*, zwei Sippen für die Schweiz angegeben. *Meylania* 23: 15–20.
- HEDENÄS, L. & BISANG, I. (2004). Key to the European *Dicranum* species. *Herzogia* **17**: 179–197.
- Hedenäs, L., Heinrichs, J. & Gallego, M.T. (2019). The Scandinavian *Syntrichia ruralis* complex (Musci. Pottiaceae): a chaos of diversification. *Plant Systematics and Evolution* **305**: 639–661. https://doi.org/10.1007/s00606-019-01596-0
- Hodgetts, N.G., Söderström, L., Blockeel, T.L., Caspari, S., Ignatov, M.S., Konstantinova, N.A., Lockhart, N., Papp, B., Schröck, C., Sim-Sim, M., Bell, D., Bell, N.E., Blom, H.H., Bruggeman-Nannenga, M.A., Brugués, M., Enroth, J., Flatberg, K.I., Garilleti, R., Hedenäs, L., Holyoak, D.T., Hugonnot, V., Kariyawasam, I., Köckinger, H., Kučera, J., Lara, F. & Porley, R.D. (2020). An annotated checklist of bryophytes of Europe, Macaronesia and Cyprus. *Journal of Bryology* **42**(1): 1–116. https://doi.org/10.1080/03736687.2019.1694329
- HOMM, TH. (2017). Leaf-born gemmae in *Syntrichia virescens* (De Not.) Ochyra a neglected feature in bryological literature. *Frahmia* **15**: 1–4.
- Hugonnot, V. (2010). Towards an improved understanding of the taxonomy of *Riccia ciliata* Hoffm. (Marchantiopsida: Ricciaceae). *Journal of Bryology* **32**: 300–303. https://doi.org/10.1179/jbr.2010.32.4.300
- JIMÉNEZ, J.A. (2006). Didymodon Hedw. In: GUERRA, J., CANO, M.J. & ROS, R.M. (eds.) Flora Briofítica Ibérica. Vol. III, Pottiales, Encalyptales. Universidad de Murcia, Sociedad Española de Briología, Murcia. pp. 217–244.
- JOVET-AST, S. (1986). Les *Riccia* de la région méditerranée. *Cryptogamie Bryologie Lichénologie* 7 Suppl. **3**: 287–431.
- KÖCKINGER, H. (2017). Die Horn- und Lebermoose Österreichs (Anthocerotophyta und Marchantiophyta) Catalogus Florae Austriae, Teil 2. Heft 2. Biosystematic and Ecology Series 32: 1–382. https://doi.org/10.2307/j.ctt1v2xvg0
- KÖCKINGER, H. & HEDENÄS, L. (2017). A farewell to *Tortella bambergeri* (Pottiaceae) as understood over the last decades. *Journal of Bryology* **39**: 213–225. https://doi.org/10.1080/03736687.2017.1307313

- Košnar, J. & Kolář, F. (2009). A taxonomic study of selected European taxa of the *Tortula muralis* (Pottiaceae, Musci) complex: variation in morphology and ploidy level. *Preslia* 81: 399–421.
- Kučera, J. (2000). Illustrierter Bestimmungsschlüssel zu den mitteleuropäischen Arten der Gattung *Didymodon. Meylania* **19**: 1–49.
- KUČERA, J., BLOCKEEL, T.L., ERZBERGER, P., PAPP, B., SOLDÁN, Z., VELLAK, K., WERNER, O. & Ros, R.M. (2018). The *Didymodon tophaceus* Complex (Pottiaceae, Bryophyta) Revisited: New Data Support the Subspecific Rank of Currently Recognized Species. *Cryptogamie, Bryologie* 39(2): 241–257. https://doi.org/10.7872/cryb/v39.iss2.2018.241
- KUČERA, J., KOŠNAR, J. & WERNER, O. (2013). Partial generic revision of *Barbula* (Musci: Pottiaceae): Re-establishment of *Hydrogonium* and *Streblotrichum*, and the new genus *Gymnobarbula*. *Taxon* 62: 21–39. https://doi.org/10.1002/tax.621004
- Laine, J., Flatberg, K.-I., Harju, P., Timonen, T., Minkkinen, K., Laine, A., Tuittila, E.-S. & Vasander, H. (2018). *Sphagnum mosses. The Stars of European Mires*. University of Helsinki Department of Forest Sciences, Helsinki, 326 pp. + > 1000 photographs.
- Laine, J., Harju, P., Timonen, T., Laine, A., Tuittila, E.-S., Minkkinen, K. & Vasander, H. (2011). The intricate beauty of *Sphagnum* mosses a Finnish guide to identification. *University of Helsinki Department of Forest Sciences Publications* 2: 1–191.
- LANDWEHR, J. (1984). *Nieuwe Atlas Nederlandse bladmossen*. Thieme, Zutphen, 568 pp.
- LIMPRICHT, K.G. (1890). Die Laubmoose Deutschlands, Oesterreichs und der Schweiz. I. Abtheilung: Sphagnaceae, Andreaeaceae, Archidiaceae, Bryinae (Cleistocarpae, Stegocarpae [Acrocarpae]). (= Dr. L. Rabenhorst's Kryptogamenflora von Deutschland, Österrreich und der Schweiz 2. Aufl.). Kummer, Leipzig, 836 pp.
- LÜTH, M. (2019). *Mosses of Europe. A Photographic Flora*. Vol. 1–3. Michael Lüth, Freiburg, 1360 pp.
- MAIER, E. (2010). The Genus *Grimmia* Hedw. (Grimmiaceae, Bryophyta) A morphological-anatomical study. *Boissiera* **63**: 1–377.
- MAGILL, R.E. (ed.) (1990). *Glossarium Polyglottum Bryologiae*. Missouri Botanical Garden, St Louis, 297 pp.
- MALCOLM, B. & MALCOLM, N. (2000). *Mosses and Other Bryophytes. An Illustrated Glossary*. Micro-Optics Press, Nelson, New Zealand, 220 pp.
- MASTRACCI, M. (1993). Taxonomic significance of stem and leaf-sheath anatomy in *Timmia* Hedw. *Journal of Bryology* **17**: 481–487.
- MASTRACCI, M. (2003). *Thamnobryum neckeroides* (Bryopsida: Neckeraceae): lectotypification, synonymies, diagnostic characters, habitat and distribution. *Journal of Bryology* **25**(2): 115–120.
- Meinunger, L. & Schröder, W. (2007). *Verbreitungsatlas der Moose Deutschlands.* 3 vols. Regensburgische Botanische Gesellschaft, Regensburg, 636, 700, 709 pp.
- MÜLLER, F. (2017). *Didymodon sicculus* and *Tortula pallida* new for Germany from inland salt marshes in eastern Germany. *Herzogia* **30**: 387–396. https://doi.org/10.13158/heia.30.2.2017.387

- Müller (Frib.), K. (1905–1916). *Die Lebermoose Deutschlands, Oesterreichs und der Schweiz*. In: Rabenhorst, G.L. (founder), *Kryptogamenflora* (ed. 2) Vol. 6 part 1, 871 pp., part 2, 947 pp., Kummer, Leipzig.
- MÜLLER (FRIB.), K. (1951–1958). *Die Lebermoose Europas*. In: RABENHORST, G.L. (founder), *Kryptogamenflora* (ed. 3) Vol. 6 Geest & Portig, Leipzig, 1365 pp.
- Murray, B.M. (1987). Andreaeaceae. In: Mogensen, G.S. (ed.): Illustrated moss flora of arctic North America and Greenland 3. Andreaeobryaceae Tetraphidaceae. *Meddel. Grønland, Biosci.* 23: 1–36.
- Murray, B.M. (1988). The genus *Andreaea* in Britain and Ireland. *Journal of Bryology* **15**(1): 17–82.
- Nebel, M. & Philippi, G. (eds.) (2000). *Die Moose Baden-Württembergs, I.* (*Andreaeales bis Funariales*). Ulmer, Stuttgart, 512 pp.
- Nebel, M. & Philippi, G. (eds.) (2001). *Die Moose Baden-Württembergs, II.* (*Schistostegales bis Hypnobryales*). Ulmer, Stuttgart, 529 pp.
- Nebel, M. & Philippi, G. (eds.) (2005). *Die Moose Baden-Württembergs, III.* (*Sphagnopsida, Marchantiopsida, Anthocerotophyta*). Ulmer, Stuttgart, 487 pp.
- NÉMETH, Cs. & ERZBERGER, P. (2015). *Anacamptodon splachnoides* (Amblystegiaceae): Hungarian populations of a moss species with a peculiar habitat. *Studia Botanica Hungarica* **46**(1): 61–75. https://doi.org/10.17110/StudBot.2015.46.1.61
- Nyholm, E. (1960). *Illustrated Moss Flora of Fennoscandia II. Musci. Fasc. 4.* Swedish Natural Science Research Council, Lund, pp. 284–408.
- Nyholm, E. (1979). *Illustrated Moss Flora of Fennoscandia II. Musci. Fasc. 5, ed. 2*. Swedish Natural Science Research Council, Stockholm, pp. 407–647.
- Nyholm, E. (1981). *Illustrated Moss Flora of Fennoscandia II. Musci. Fasc. 6, ed. 2*. Swedish Natural Science Research Council, Stockholm, pp. 647–799.
- Nyholm, E. (1987). Illustrated Flora of Nordic Mosses. Fasc. 1. Fissidentaceae Seligeriaceae. Nordic Bryological Society, Copenhagen and Lund, pp. 1–72.
- Nyholm, E. (1990). *Illustrated Flora of Nordic Mosses. Fasc. 2. Pottiaceae Splachnaceae Schistostegaceae*. Nordic Bryological Society, Copenhagen and Lund. pp. 75–141.
- Nyholm, E. (1993). *Illustrated Flora of Nordic Mosses. Fasc. 3. Bryaceae Rhodobryaceae Mniaceae Cinclidiaceae Plagiomniaceae.* Nordic Bryological Society, Copenhagen and Lund, pp. 145–244.
- Nyholm, E. (1998). Illustrated Flora of Nordic Mosses, Fasc. 4 (Aulacomniaceae Meesiaceae Catascopiaceae Bartramiaceae Timmiaceae Encalyptaceae Grimmiaceae Ptychomitraceae Hedwigiaceae Orthotrichaceae). Nordic Bryological Society, Copenhagen and Lund, pp. 249–404.
- Orbán, S. & Vajda, L. (1983). *Magyarország mohaflórájának kézikönyve (Handbook of the Hungarian bryophyte flora)*. Akadémiai Kiadó, Budapest, 518 pp.
- PAPP, B., ERZBERGER, P., ÓDOR, P., HOCK, Zs., SZÖVÉNYI, P., SZURDOKI, E. & TÓTH, Z. (2010). Updated checklist and red list of Hungarian bryophytes. *Studia Botanica Hungarica* **41**: 31–59.
- PATON, J.A. (1999). *The Liverwort Flora of the British Isles*. Harley Books, Great Horkesley, 626 pp.
- PLÁŠEK, V., SAWICKI, J., TRÁVNIČKOVÁ, V., PASEČNÁ, M. (2009). *Orthotrichum moravicum* (Orthotrichaceae), a new moss species from the Czech Republic. *The Bryologist* **112**: 329–336. https://doi.org/10.1639/0007-2745-112.2.329

- Schlüsslmayr, G. (2005). Soziologische Moosflora des Südlichen Oberösterreich. *Stapfia* **84**: 1–695.
- Schumacker, R. & Váňa, J. (2005). *Identification keys to the liverworts and hornworts of Europe and Macaronesia (distribution & status). Second edition fully revised and updated.* Sorus, Poznań, 209 pp.
- SIEBEL, H.N. & DURING, H.J. (2006). *Beknopte Mosflora van Nederland en België*. KNNV Uitgeverij, Utrecht, 559 pp.
- SMITH, A.J.E. (1978). *The Moss Flora of Britain and Ireland*. Cambridge University Press, Cambridge, New York, Port Chester, Melbourne, Sidney, 706 pp.
- SMITH, A.J.E. (1991). *The Liverworts of Britain & Ireland*. Cambridge University Press, Cambridge, 362 pp.
- SMITH, A.J.E. (2004). *The Moss Flora of Britain and Ireland, 2nd ed.* University Press, Cambridge, 1012 pp.
- Szurdoki, E. (2000). *Tőzegmohás élőhelyek Magyarországon: kutatás, kezelés, védelem*. CEEWEB Munkacsoport, Miskolc, 184 pp.
- SZURDOKI, E. (2003). Peatmosses of North Hungary. *Studia Botanica Hungarica* **34**: 55–79.
- SZURDOKI, E., MÁRTON, O. & SZÖVÉNYI, P. (2014). Genetic and morphological diversity of *Sphagnum angustifolium, S. flexuosum, S. fallax* in Europe. *Taxon* **63**(2): 237–248. https://doi.org/10.12705/632.6
- SZURDOKI, E. & NAGY, J. (2002). *Sphagnum* dominated mires and *Sphagnum* occurrences of North-Hungary. *Folia Historico-Naturalia Musei Matraensis* **26**: 67–84.
- SZURDOKI, E., TÓTH, Z. & PELLES, G. (1999–2000). The *Sphagnum* populations of the Zemplén Mts, NE Hungary. *Studia Botanica Hungarica* **30–31**: 113–125.
- Szweykowski, J. & Mendelak, M. (1964). Experimental investigations of the variability of *Riccia gougetiana* and *Riccia ciliifera* from Czechoslovakia. *Acta Societatis Botanicorum Poloniae* **33**: 359–369.
- Touw, A. & Rubers, W.V. (1989). *De Nederlandse Bladmossen*. Stichting Uitgeverij Koninklijke Nederlandse Natuurhistorische Vereniging, Utrecht, 532 pp.
- VIGALONDO, B., DRAPER, I., MAZIMPAKA, V., CALLEJA, J.A., LARA, F. & GARILLETI, R. (2020). The *Lewinskya affinis* complex (Orthotrichaceae) revisited: species description and differentiation. *The Bryologist* **123**(3): 454–481. https://doi.org/10.1639/0007-2745-123.3.454
- WATSON, E.V. (1981). *British Mosses and Liverworts*. Cambridge University Press, Cambridge, 537 pp.
- Wolski, G.J. & Krawczyk, P. (2020). Resurrection of the *Plagiothecium longisetum* Lindb. and proposal of the new species *P. angusticellum. PLoS ONE* **15**(3): e0230237. https://doi.org/10.1371/journal.pone.0230237
- ZÜNDORF, H.-J. (1988). Moose Mecklenburgs II: Leucobryum glaucum und Leucobryum juniperoideum. Botanischer Rundbrief für den Bezirk Neubrandenburg 20: 55–60.

TABLE OF CONTENTS

CONSPECTUS OF CLASSIFICATION	20
Anthocerotophyta (Hornworts)	20
Marchantiophyta (Liverworts)	20
Bryophyta (Mosses)	23
PRELIMINARY REMARKS – HOW TO USE THESE KEYS	30
ABBREVIATIONS	30
FREQUENCY CLASSES	31
RECOMMENDED LITERATURE	31
PART I: GENERAL KEYS	36
1. Key to the main groups of bryophytes	36
2. Keys to genera of thalloid bryophytes	37
Hornworts	37
Thalloid liverworts	37
Group 1 Thallus with internal differentiation; dorsal epidermis with	
pores; rhizoids smooth or warty	37
Group 2 Thallus without internal differentiation, dorsal epidermis	
without pores, rhizoids smooth	38
3. Keys to genera of foliose liverworts	39
Group 3 Leaves simple, succubous	40
Group 4 Leaves 3–5-lobed	41
Group 5 Leaves conduplicate, underleaves lacking	41
Group 6 Leaves conduplicate, underleaves present	42
Group 7 Leaves bilobed, transversely or subtransversely inserted	42
Group 8 Leaves bilobed, longitudinally or obliquely inserted,	
succubous	44
4. Keys to genera of mosses	44
Group 9 Leaves distichous or complanate	47
Group 10 Costa or ventral leaf surface with filaments or lamellae	48
Group 11 Leaves with hyaline point or costa excurrent in hair point	48
Group 12 Leaves bordered with narrow cells or margin pluristratose	50
Group 13 Acrocarps with indehiscent capsule	51
Group 14 Acrocarps with immersed or emergent capsule	51
Group 15 Acrocarps with exserted globose or subglobose capsule	52
Group 16 Acrocarps with exserted strumose capsule	53
Group 17 Acrocarps with exserted capsule and peristome lacking or	
rudimentary	53
Group 18 Acrocarps with exserted capsule, striate or sulcate when dry	
and peristome well developed	54
Group 19 Acrocarps with exserted erect capsule, peristome teeth 16,	
entire or slightly and irregularly divided	56
Group 20 Acrocarps with exserted erect capsule, peristome teeth 16,	
divided halfway or to the base (32 teeth)	57
Group 21 Acrocarps with exserted capsule, inclined or pendulous,	
peristome simple	59

Group 2	2 Acrocarps with exserted capsule, inclined to pendulous,	
peris	stome double	59
Group 2	3 Acrocarps with propagules on stems, leaves or in	
rece	ptacles	60
Group 2	4 Acrocarps with costa 1/3 or more of leaf base	62
	5 Acrocarps with lamina cells 18 µm wide or more	63
	6 Acrocarps with alar cells differentiated	66
	7 Acrocarps with lamina cells isodiametric and leaf margins	
	iculate or dentate, at least near apex or base	66
	8 Acrocarps with lamina cells isodiametric and excurrent	00
costa		67
	9 Acrocarps with isodiametric lamina cells, leaf apex obtuse or	07
	ided, apiculate or not and costa not excurrent	69
	0 Acrocarps with isodiametric cells, apex acute, subacute or	0)
	ninate, margins recurved at least on one side, costa not	
		70
	rrent or lacking	70
	1 Acrocarps with isodiametric cells, apex acute, subacute or	
	ninate, margins plane or recurved at base only, costa not	71
	rrent	71
	2 Acrocarps with lamina cells elongate, leaves acuminate or	
	late and apex consisting largely or entirely of costa	72
	3 Acrocarps with lamina cells elongate, leaf apex obtuse to	
	ninate, costa percurrent or excurrent, short or lacking	73
_	4 Pleurocarps with long costa and lamina cells short, at least	
	argins	74
	5 Pleurocarps with longitudinally plicate leaves, costa long	
	lamina cells elongate	75
	6 Pleurocarps with squarrose or falcate leaves, long costa and	
	gated lamina cells	76
	7 Pleurocarps with long costa, elongated lamina cells and	
	ided, obtuse, or obtuse and apiculate apex	77
Group 3	8 Pleurocarps with long costa, elongated lamina cells and	
	e or acuminate apex	78
Group 3	9 Pleurocarps with short or lacking costa, short lamina cells at	
least	at margin	80
Group 4	0 Pleurocarps with short or lacking costa, elongated lamina	
cells	and rounded, obtuse or apiculate apex	80
Group 4	1 Pleurocarps with distinctly falcate or squarrose leaves, short	
	cking costa, elongated lamina cells and acute or acuminate	
apex		81
	2 Pleurocarps with straight, slightly falcate or squarrose	
	es, costa single, short, long and double, or lacking, elongated	
	na cells and acute or acuminate apex	82
PART II: SPI	<u> </u>	85
_	pecies of Liverworts	85
	pecies of Barbilophozia group, incl. Neoorthocaulis floerkii	85
	pecies of Cephaloziaceae	86
	pecies of Gephaloziella	87

Key to Lophoziaceae group. (incl. <i>Barbilophozia, Isopaches, Lophozia,</i>	
Lophoziopsis, Mesoptychia, Neoorthocaulis, Obtusifolium,	
Schistochilopsis, Trilophozia and Tritomaria)	89
Key to species of Diplophyllum and the genus Scapania	94
Key to species of <i>Scapania</i>	95
Supplementary 'key' to species of <i>Scapania</i> according to habitats	98
Key to species of <i>Calypogeia</i>	98
Key to species of Marsupella	100
Key to species of <i>Nardia</i>	100
Key to Jungermanniaceae group (incl. Endogemma, Jungermannia,	
Liochlaena, Nardia, Solenostoma, Syzygiella)	101
Key to species of <i>Mesoptychia</i>	103
Key to species of Lophocoleaceae	104
Key to species of Plagiochilaceae	106
Key to species of <i>Frullania</i>	106
Key to species of Lejeuneaceae	108
Key to species of <i>Porella</i>	108
Key to species of <i>Radula</i>	109
Key to species of Aneuraceae	109
Key to species of <i>Metzgeria</i>	110
Key to species of <i>Fossombronia</i>	111
Key to species of Pelliaceae	111
Key to Marchantiales pp. (incl. Asterella, Clevea, Conocephalum,	
Lunularia, Mannia, Marchantia, Reboulia)	112
Key to species of <i>Riccia</i> group (incl. <i>Oxymitra, Riccia,</i> and <i>Ricciocarpos</i>)	115
6. Keys to species of Mosses	122
Key to sections and species of <i>Sphagnum</i>	122
Key to the varieties of <i>Andreaea rupestris</i>	126
Key to species of Polytrichaceae	126
Auxiliary key for sterile plants of <i>Polytrichum, Polytrichastrum</i> and	
Pogonatum	129
Key to species of <i>Buxbaumia</i>	130
Key to species of <i>Timmia</i>	130
Key to species of <i>Encalypta</i>	130
Key to species of Funariaceae	131
Key to species of <i>Flexitrichum</i>	133
Key to species of <i>Campylopus</i>	133
Key to species of Leucobryum	135
Key to species of <i>Dicranella</i>	135
Key to species of <i>Fissidens</i>	137
Key to species of Dicranaceae	139
Key to species of Rhabdoweisiaceae	142
Key to species of Ditrichaceae s.l. incl. <i>Saelania, Flexitrichum</i>	143
Key to species of <i>Acaulon</i>	145
Key to species of <i>Aloina</i>	145
Key to species of <i>Cinclidotus</i> s.l. incl. <i>Dialytrichia</i>	146
Key to species of <i>Crossidium</i>	147
Key to species of <i>Didymodon</i>	147

Key to infraspecific taxa of <i>Didymodon tophaceus</i>	151
Key to species of <i>Microbryum</i>	151
Key to Pottia s.l., incl. Hennediella heimii, Tortula caucasica, T.	
lindbergii, T. protobryoides, T. truncata, but without species	
formerly in Phascum: Microbryum curvicollum, M. floerkeanum	153
Key to species of <i>Pseudocrossidium</i>	155
Key to species of <i>Pterygoneurum</i>	155
Key to species of <i>Syntrichia</i>	156
Key to species of <i>Tortula</i> s.l. (excl. <i>T. acaulon, T. caucasica, T. lindbergii,</i>	
T. protobryoides, T. truncata, incl. Syntrichia, Tortula cernua,	
Hilpertia velenovskyi)	160
Key to species of <i>Tortula</i>	166
Key to varieties of <i>Streblotrichum convolutum</i>	169
Key to species of <i>Ephemerum</i>	169
Key to species of <i>Gymnostomum</i> group	170
Key to species of <i>Hydrogonium</i>	171
Key to species of <i>Trytal agontum</i>	171
Key to species of <i>Trichostomum</i>	172
Key to species of Weissia	173
Key to species of Weissia Key to species of Seligeriaceae	174
Key to species of Grimmiaceae	175
Key to species of <i>Hedwigia</i>	188
Key to species of Bartramiaceae	189
Key to species of Bryaceae	190
Key to species of <i>Pohlia</i>	198
Key to species of Mnium, Plagiomnium and Rhizomnium	201
Key to species of Orthotrichaceae	203
Key to species of Aulacomnium	211
Key to species of <i>Funtacommum</i> Key to species of <i>Fontinalis</i>	211
Key to species of <i>Politinans</i> Key to species of <i>Plagiothecium</i>	212
Key to species of <i>Orthothecium</i>	214
Key to species of <i>Fabronia</i>	214
Key to species of <i>Palustriella</i>	214
Key to species of <i>Campyliadelphus</i>	215
Key to species of <i>Campylium</i>	215
Key to species of <i>Campyhum</i> Key to species of <i>Drepanocladus</i>	215
Key to species of <i>Brepanochauas</i> Key to species of <i>Amblystegium</i> group (incl. <i>Amblystegium</i> ,	213
Hygroamblystegium, Pseudoamblystegium, Pseudocampylium and	
Serpoleskea)	216
Key to species of <i>Calliergon</i> s.l. incl. <i>Straminergon</i>	218
Key to species of <i>Camergon</i> s.i. mci. <i>Strammergon</i> Key to species of <i>Scorpidium</i>	218
Key to species of <i>Scorplanam</i> Key to species of <i>Pseudoleskeella</i>	219
Key to varieties of Abietinella abietina	219
· ·	219
Key to species of <i>Thuidium</i>	
Key to species of Eurhynchium	220 220
Key to species of Rhynchostegium	
Key to species of Cirriphyllum	221
Key to species of <i>Oxyrrhynchium</i>	221

Erzberger (2021): Identification keys for Hungarian bryophytes

Key to species of Rhynchostegiella	222
Key to species of <i>Homalothecium</i>	222
Key to species of Brachythecium s.l. (incl. Brachytheciastrum,	
Brachythecium and Sciuro-hypnum)	223
Key to species of <i>Hypnum</i> s.l. incl. <i>Buckia</i>	227
Key to species of <i>Taxiphyllum</i>	228
Key to species of Calliergonella	228
Key to species of Rhytidiadelphus	228
Key to species of Neckera s.l. incl. Alleniella, Exsertotheca	228
Key to species of <i>Thamnobryum</i>	229
Key to species of <i>Isothecium</i>	229
Key to species of Anomodon s.l. incl. Pseudanomodon, Claopodium	230
GLOSSARY	231
INDEX OF GENERA	253

CONSPECTUS OF CLASSIFICATION

The taxonomic hierarchy presented below is based on the most recent European checklist (Hodgetts *et al.* 2020), and the most recent Hungarian checklist for the set of taxa considered (Erzberger and Papp 2020). Although it represents the most up-to-date classification, it will probably be outdated very soon since current concepts are in a state of flux due to ongoing molecular work. Only orders, families and genera are listed in this conspectus.

Anthocerotophyta (Hornworts)

Order	1.	Anthocerotales
Family	1.	Anthocerotaceae
	1.	Anthoceros
Order	2.	Notothyladales
Family	2.	Notothyladaceae
	2.	Phaeoceros

Marchantiophyta (Liverworts)

Order Family	3. 3.	Jungermanniales Adelanthaceae
Tumny	3.	Syzygiella
Family	4.	Anastrophyllaceae
	4.	Anastrophyllum
	5.	Barbilophozia
	6.	Crossocalyx
	7.	Gymnocolea
	8.	Isopaches
	9.	Neoorthocaulis
	10.	Sphenolobus
Family	5.	Cephaloziaceae
	11.	Cephalozia
	12.	Fuscocephaloziopsis
	13.	Nowellia
Family	6.	Cephaloziellaceae
	14.	Cephaloziella
	15.	Obtusifolium

Family	7.	Lophoziaceae
1 unity	16.	Lophozia
	17.	Lophoziopsis
	18.	Trilophozia
	19.	Tritomaria
Family	8.	Scapaniaceae
1 unity	20.	Diplophyllum
	21.	Scapania
	22.	Schistochilopsis
Family	9.	Calypogeiaceae
Tunniy	23.	Calypogeia
Family	10.	Endogemmataceae
Taning	24.	Endogemma
Family	11.	Gymnomitriaceae
Taniny	25.	Marsupella
	26.	Nardia
Family	12.	Jungermanniaceae
ranniy	27.	Liochlaena
	28.	
	26. 29.	Jungermannia Masantushia
Eamily	29. 13.	<i>Mesoptychia</i> Solenostomataceae
Family		
P 1	30.	Solenostoma
Family	14.	Blepharostomataceae
P 1	31.	Blepharostoma
Family	15.	Lepidoziaceae
	32.	Bazzania
	33.	Lepidozia
Family	16.	Lophocoleaceae
	34.	Chiloscyphus
_	35.	Lophocolea
Family	17.	Plagiochilaceae
	36.	Pedinophyllum
	37.	Plagiochila
Family	18.	Trichocoleaceae
	38.	Trichocolea
Order	4.	Porellales
Family	19.	Frullaniaceae
	39.	Frullania
Family	20.	Lejeuneaceae
	40.	Cololejeunea

	4.4	7
T	41.	Lejeunea
Family	21	Porellaceae
	42.	Porella
Family	22.	Radulaceae
	43.	Radula
Order	5.	Ptilidiales
Family	23.	Ptilidiaceae
	44.	Ptilidium
Order	6.	Metzgeriales
Family	24.	Aneuraceae
	45.	Aneura
	46.	Riccardia
Family	25.	Metzgeriaceae
	47.	Metzgeria
Order	7.	Fossombroniales
Family	26.	Fossombroniaceae
	48.	Fossombronia
Order	8.	Pelliales
Family	27.	Pelliaceae
	49.	Apopellia
	50.	Pellia
Order	9.	Blasiales
Family	28.	Blasiaceae
	51.	Blasia
Order	10.	Lunulariales
Family	29.	Lunulariaceae
5	52.	Lunularia
Order	11.	Marchantiales
Family	30.	Aytoniaceae
5	53.	Asterella
	54.	Mannia
	55.	Reboulia
Family	31.	Cleveaceae
y	56.	Clevea
Family	32.	Conocephalaceae
2 4	57.	Conocephalum
Family	33.	Marchantiaceae
- y	58.	Marchantia
Family	34.	Oxymitraceae
	59.	Oxymitra
	٠,٠	

Family	35. 60.	Ricciaceae <i>Riccia</i>
	61.	Ricciocarpos
Order	12 .	Sphaerocarpales
Family	36.	Sphaerocarpaceae
	62.	Sphaerocarpos

Bryophyta (Mosses)

Order	13.	Sphagnales
Family	37.	Sphagnaceae
	63.	Sphagnum
Order	14.	Andreaeales
Family	38.	Andreaeaceae
	64.	Andreaea
Order	15.	Tetraphidales
Family	39.	Tetraphidaceae
	65.	Tetraphis
Order	16.	Polytrichales
Family	40.	Polytrichaceae
•	66.	Atrichum
	67.	Pogonatum
	68.	Polytrichastrum
	69.	Polytrichum
Order	17.	Buxbaumiales
Family	41.	Buxbaumiaceae
•	70.	Buxbaumia
Order	18.	Diphysciales
Family	42.	Diphysciaceae
•	71.	Diphyscium
Order	19.	Timmiales
Family	43.	Timmiaceae
•	72.	Timmia
Order	20.	Encalyptales
Family	44.	Encalyptaceae
•	73.	Encalypta
Order	21.	Funariales
Family	45.	Funariaceae
•	74.	Pyramidula
	75.	Entosthodon

	76.	Funaria
	77.	Physcomitrium
Order	22.	Dicranales
Family	46.	Distichiaceae
,	78.	Distichium
Family	47.	Flexitrichaceae
· ,	79.	Flexitrichum
Family	48.	Archidiaceae
,	80.	Archidium
Family	49.	Leucobryaceae
,	81.	Campylopus
	82.	Dicranodontium
	83.	Leucobryum
Family	50.	Amphidiaceae
,	84.	Amphidium
Family	51.	Aongstroemiaceae
, and the second	85.	Dichodontium
Family	52.	Dicranellaceae
, and the second	86.	Dicranella
Family	53.	Fissidentaceae
, and the second	87.	Fissidens
Family	54.	Dicranaceae
-	88.	Dicranum
	89.	Paraleucobryum
Family	55.	Rhabdoweisiaceae
	90.	Cnestrum
	91.	Cynodontium
	92.	Dicranoweisia
	93.	Rhabdoweisia
Family	56.	Bruchiaceae
	94.	Bruchia
Family	57.	Ditrichaceae
	95.	Ceratodon
	96.	Ditrichum
	97.	Pleuridium
	98.	Pseudephemerum
	99.	Trichodon
Family	58.	Pottiaceae
	100.	Acaulon
	101.	Aloina

	102.	Barbula
	103.	Bryoerythrophyllum
	104.	Chenia Cinalidatas
	105.	Cinclidotus
	106.	Crossidium
	107.	Didymodon
	108.	Hennediella
	109.	Hilpertia
	110.	Microbryum
	111.	Pseudocrossidium
	112.	Pterygoneurum
	113.	Syntrichia
	114.	Tortula
	115.	Streblotrichum
	116.	Chionoloma
	117.	Ephemerum
	118.	Eucladium
	119.	Gymnostomum
	120.	Gyroweisia
	121.	Hydrogonium
	122.	Hymenostylium
	123.	Splachnobryum
	124.	Tortella
	125.	Trichostomum
	126.	Weissia
Order	23.	Grimmiales
Family	59.	Saelaniaceae
. J	127.	Saelania
Family	60.	Seligeriaceae
	128.	Blindia
	129.	Blindiadelphus
	130.	Seligeria
Family	61.	Ptychomitriaceae
1 dilliny	131.	Brachydontium
	132.	Campylostelium
Family	62.	Grimmiaceae
1 diffily	133.	Racomitrium
	134.	Coscinodon
	13 4 . 135.	Grimmia
	135. 136.	Schistidium
	130.	SCHISCIUIUIII

Order	24.	Hedwigiales
Family	63.	Hedwigiaceae
	137.	Hedwigia
Order	25.	Bartramiales
Family	64.	Bartramiaceae
	138.	Bartramia
	139.	Plagiopus
	140.	Philonotis
Family	65.	Meesiaceae
	141.	Amblyodon
	142.	Leptobryum
	143.	Meesia
Order	26.	Bryales
Family	66.	Bryaceae
	144.	Bryum
	145.	Imbribryum
	146.	Ptychostomum
	147.	Rhodobryum
Family	67.	Mniaceae
	148.	Pohlia
	149.	Mnium
	150.	Plagiomnium
	151.	Rhizomnium
Order	27.	Orthotrichales
Family	68.	Orthotrichaceae
	152.	Codonoblepharon
	153.	Lewinskya
	154.	Nyholmiella
	155.	Orthotrichum
	156.	Pulvigera
	157.	Ulota
	158.	Zygodon
Order	28.	Orthodontiales
Family	69.	Orthodontiaceae
	159.	Orthodontium
Order	29.	Aulacomniales
Family	70.	Aulacomniaceae
	160.	Aulacomnium
Order	30.	Hookeriales
Family	71.	Hookeriaceae

	161.	Hookeria
Order	31.	Hypnales
Family	72.	Fontinalaceae
	162.	Fontinalis
Family	73.	Plagiotheciaceae
	163.	Herzogiella
	164.	Isopterygiopsis
	165.	Plagiothecium
	166.	Myurella
	167.	Orthothecium
	168.	Platydictya
	169.	Pseudotaxiphyllum
Family	74.	Fabroniaceae
	170.	Fabronia
Family	75.	Pterigynandraceae
-	171.	Pterigynandrum
Family	76.	Climaciaceae
-	172.	Climacium
Family	77.	Amblystegiaceae
	173.	Cratoneuron
	174.	Palustriella
	175.	Amblystegium
	176.	Anacamptodon
	177.	Campyliadelphus
	178.	Campylium
	179.	Campylophyllopsis
	180.	Conardia
	181.	Drepanocladus
	182.	Hygroamblystegium
	183.	Hygrohypnum
	184.	Leptodictyum
	185.	Pseudoamblystegium
	186.	Pseudocampylium
	187.	Serpoleskea
	188.	Tomentypnum
Family	78.	Calliergonaceae
-	189.	Calliergon
	190.	Sarmentypnum
	191.	Straminergon
Family	79.	Scorpidiaceae

	192.	Hamatocaulis
	193.	Sanionia
	194.	Scorpidium
Family	80.	Leskeaceae
,	195.	Claopodium
	196.	Leskea
Family	81.	Pseudoleskeaceae
,	197.	Lescuraea
Family	82.	Pseudoleskeellaceae
5	198.	Pseudoleskeella
Family	83.	Thuidiaceae
,	199.	Abietinella
	200.	Helodium
	201.	Thuidium
Family	84.	Brachytheciaceae
	202.	Eurhynchium
	203.	Plasteurhynchium
	204.	Pseudoscleropodium
	205.	Rhynchostegium
	206.	Cirriphyllum
	207.	Microeurhynchium
	208.	Oxyrrhynchium
	209.	Rhynchostegiella
	210.	Brachytheciastrum
	211.	Brachythecium
	212.	Eurhynchiastrum
	213.	Homalothecium
	214.	Kindbergia
	215.	Sciuro-hypnum
Family	85.	Hypnaceae
-	216.	Нурпит
Family	86.	Callicladiaceae
•	217.	Callicladium
Family	87.	Taxiphyllaceae
·	218.	Taxiphyllum
Family	88.	Pylaisiadelphaceae
-	219.	Platygyrium
Family	89.	Pylaisiaceae
-	220.	Buckia
	221.	Calliergonella

	222. 223.	Homomallium Ptilium
P 4	224.	Pylaisia
Family	90.	Sematophyllaceae
	225.	Sematophyllum
Family	91.	Hylocomiaceae
	226.	Hylocomiadelphus
	227.	Hylocomium
	228.	Loeskeobryum
	229.	Pleurozium
	230.	Rhytidiadelphus
Family	92.	Rhytidiaceae
	231.	Rhytidium
Family	93.	Entodontaceae
	232.	Entodon
Family	94.	Leucodontaceae
	233.	Leucodon
	234.	Nogopterium
Family	95.	Antitrichiaceae
-	235.	Antitrichia
Family	96.	Neckeraceae
•	236.	Alleniella
	237.	Exsertotheca
	238.	Homalia
	239.	Leptodon
	240.	Neckera
	241.	Pseudanomodon
	242.	Thamnobryum
Family	97.	Heterocladiellaceae
· J	243.	Heterocladiella
Family	98.	Lembophyllaceae
y	244.	Heterocladium
	245.	Isothecium
Family	99.	Myuriaceae
- 	246.	Ctenidium
Family	100.	Anomodontaceae
	247.	Anomodon
	- 17.	11110111011011

PRELIMINARY REMARKS - HOW TO USE THESE KEYS

The keys presented below require the use of a compound microscope and ideally also of a stereomicroscope, preferably equipped with micrometer eyepieces to enable measurement of small distances. Some preparation skills are also useful.

Following the directions in the keys, beginning with the key to the main groups of bryophytes (1.), the user will eventually arrive at the name of a genus or a group of genera, which can then be looked up in the second part, keys to species of liverworts and hornworts (5.), and keys to species of mosses (6.), to obtain the name of the species in question. If there is only one species to a genus, the species name is given already in part one.

After arriving at a species name, it is essential that the result should be carefully compared with illustrations and descriptions of the species including remarks about its ecological demands. Unfortunately, in this respect the user must consult some of the treatments listed below, since an up-to-date bryophyte flora of Hungary is still missing. Since numerous bryophyte taxa that were not hitherto known to occur in Hungary have been detected during the last years, it is always possible that the user might try to identify a plant that is not treated in the present keys. It is only by consulting detailed descriptions and illustrations that such a situation can be noticed. Also, the keys cannot take account of every possibility given that certain taxa show a wide range of variability. If the user cannot decide between the alternatives in a given couplet of the key, both paths should be followed. Often taxa that may display both character states of a given couplet have been keyed out under both alternatives.

ABBREVIATIONS

- agg. aggregate, a group of related species (or microspecies) which are difficult to distinguish from one another
- pp. pro parte, meaning part of a taxon, e.g. only some species of a genus, not all
- s.l. sensu lato, meaning in the wider sense, when a name may denote different circumscriptions of a taxon, e.g. *Ulota crispa* s.l. (*U. crispa* s.str., *U. crispula*, *U. intermedia*)
- s.str. sensu stricto, meaning in the narrow sense

FREQUENCY CLASSES

cc very common

c common

w widespread

r rare

rr very rare

n.s. not seen, i.e. without records after 1973

RECOMMENDED LITERATURE

(References for particular groups can be found in the second part: Keys to species. Without aiming at completeness, these have often been selected because of some reference to Hungary. They usually will guide the user to other important references.)

Hornworts and liverworts

Illustrations and descriptions

PATON, J.A. (1999). *The Liverwort Flora of the British Isles*. Harley Books, Great Horkesley, 626 pp. – Line drawings of many details, often showing variability of characters.

Does not treat the following liverworts: Anastrophyllum michauxii, Asterella saccata, Cephalozia lacinulata, Cephaloziella divaricata var. scabra, C. varians, Clevea hyalina, Conocephalum salebrosum, Frullania cleistostoma, F. jackii, Liochlaena subulata, Lophocolea minor, Lophozia ascendens, Mannia fragrans, M. triandra, Oxymitra incrassata, Porella baueri, Riccia ciliata, R. ciliifera, R. frostii, R. gougetiana, R. papillosa, R. warnstorfii, Scapania apiculata, S. mucronata. The treatment of Conocephalum conicum is out-dated. The plants described and illustrated under Riccia bifurca belong to R. subbifurca agg. (Meinunger and Schröder 2007).

DAMSHOLT, K. (2002). *Illustrated Flora of Nordic Liverworts and Hornworts*. Nordic Bryological Society, Lund. 837 pp. – Very precise drawings, taxonomically up to date.

Does not treat the following liverworts: Calypogeia suecica, Cephaloziella divaricata var. scabra, Cololejeunea rossettiana, Conocephalum salebrosum, Frullania cleistostoma, Mannia triandra, Oxymitra incrassata, Pedinophyllum interruptum, Riccia crozalsii, R.

- frostii, R. gougetiana, R. papillosa, Scapania parvifolia, Sphaerocarpos europaeus.
- CASAS, C., BRUGUÉS, M., CROS, R.M., SÉRGIO, C. & INFANTE, M. (2009). Handbook of Liverworts and Hornworts of the Iberian Peninsula and the Balearic Islands. Institut d'Estudis Catalans, Barcelona, 177 pp. – Keys, short descriptions, illustrations of some essential features.
 - Does not treat the following liverworts: Anastrophyllum michauxii, Asterella saccata, Cephalozia lacinulata, Cephaloziella divaricata var. scabra, C. varians, Endogemma caespiticia, Frullania cleistostoma, F. jackii, Fuscocephaloziopsis macrostachya, Liochlaena subulata, Mannia triandra, Riccia rhenana, Scapania lingulata, S. parvifolia.
- SMITH, A.J.E. (1991). *The Liverworts of Britain & Ireland*. Cambridge University Press, Cambridge, 362 pp. Second choice after the publication of Paton (1999) as concerns the illustrations as well as the descriptions. Missing taxa are the same as in Paton (1999).
- FREY, W., FRAHM, J.-P., FISCHER, E. & LOBIN, W. (2006). *The Liverworts, Mosses and Ferns of Europe. English edition revised and edited by T.L. Blockeel.* Harley Books, Colchester, 512 pp. Treats all species occurring in Hungary, but mostly very briefly, and not all are illustrated. Illustrations for most taxa missing in other accounts can usually be found.
- SCHUMACKER, R. & VÁŇA, J. (2005). *Identification keys to the liverworts and hornworts of Europe and Macaronesia (distribution & status). Second edition fully revised and updated.* Sorus, Poznań, 209 pp. only keys, no descriptions, no illustrations, but covering all of Europe.

Mosses

Illustrations

LÜTH, M. (2019). Mosses of Europe. A Photographic Flora. Vol. 1–3. Michael Lüth, Freiburg, 1360 pp. – Nearly all mosses occurring in Hungary are shown in very attractive photographs, including many microscopic details! Available for purchase on www.mosses-of-europe.com. Many of Michael Lüth's brilliant moss photographs can also be found / downloaded on his website: www.milueth.de, www.bildatlas-moose.de

- LANDWEHR, J. (1984). *Nieuwe Atlas Nederlandse bladmossen*. Thieme, Zutphen, 568 pp. Excellent drawings of the species occurring in Netherlands; most of the drawings (including also those of the corresponding atlas on liverworts) have been included in the following:
- SIEBEL, H.N. & DURING, H.J. (2006). *Beknopte Mosflora van Nederland en België*. KNNV Uitgeverij, Utrecht, 559 pp. with keys and descriptions in Dutch.

Illustrations and descriptions

- ATHERTON, I., BOSANQUET, S.D.S. & LAWLEY, M. (eds.) (2010). *Mosses and Liverworts of Britain and Ireland*. A Field Guide. British Bryological Society, Plymouth, 848 pp. Many of the species occurring in Hungary can be found in this useful book, with many helpful photographs, but without reference to microscopic details, only field characters (visible with a hand-lens) are treated.
- CASAS, C., BRUGUÉS, M., CROS, R.M. & SÉRGIO, C. (2006). Handbook of Mosses of the Iberian Peninsula and the Balearic Islands. Institut d'Estudis Catalans, Barcelona, 349 pp. Mainly keys, short descriptions and line drawings of important characters. For the list of taxa missing in the Iberian Peninsula, see the next reference.
- GUERRA, J. & CROS, R.M. (coord.) (2006–2016). Flora Briofítica Ibérica. 6 Volumes. Universidad de Murcia, Sociedad Española de Briología, Murcia. (In Spanish). Very detailed line drawings and extensive descriptions.
 - Does not treat the following mosses: Anacamptodon splachnoides, Anomodon rugelii, Blindiadelphus campylopodus, Brachythecium capillaceum, Brachythecium geheebii, Bruchia flexuosa, Bryum barnesii, Bryum stirtonii, Bryum violaceum, Bryum warneum, Callicladium haldanianum, Cinclidotus danubicus, Cnestrum schisti, Crossidium laxefilamentosum, Cynodontium tenellum, Dicranella crispa, Dicranella humilis, Dicranum fulvum, Didymodon glaucus, Ditrichum pallidum, Drepanocladus lycopodioides, Drepanocladus sendtneri, Drepanocladus sordidus, Fissidens arnoldii, Grimmia plagiopodia, Grimmia teretinervis, Helodium blandowii, Hilpertia velenovskyi, Hydrogonium consanguineum, Hypnum pallescens var. reptile, Lescuraea saviana, Microbryum muticum, Neckera pennata, Nyholmiella gymnostoma, Orthodontium lineare, Orthotrichum urnigerum, Physcomitrium

eurystomum, Physcomitrium sphaericum, Plagiothecium latebricola, Pohlia lutescens, Pohlia nutans subsp. schimperi, Polytrichum perigoniale, Pseudocampylium radicale, Racomitrium microcarpon, Rhynchostegium rotundifolium, Schistidium confusum, Schistidium lancifolium, Sematophyllum adnatum, Sphagnum inundatum, Sphagnum obtusum, Sphagnum riparium, Splachnobryum obtusum, Taxiphyllum densifolium, Thamnobryum neckeroides, Tortula cernua, Tortula muralis subsp. obtusifolia, Weissia rostellata.

- Reference to the six volumes of Flora Briofitica Ibérica and their content
- Brugués, M., Cros, R.M. & Guerra, J. (eds.) (2007). Flora Briofítica Ibérica. Vol. I, Sphagnales, Andreaeales, Polytrichales, Tetraphidales, Buxbaumiales, Diphysciales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 183 pp.
- BRUGUÉS, M.& GUERRA, J. (eds.) (2015). Flora Briofítica Ibérica. Vol. II, Archidiales, Dicranales, Fissidentales, Seligeriales, Grimmiales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 355 pp.
- GUERRA, J., CANO, M.J. & ROS, R.M. (eds.) (2006). *Flora Briofítica Ibérica. Vol. III, Pottiales, Encalyptales*. Universidad de Murcia, Sociedad Española de Briología, Murcia, 305 pp.
- GUERRA, J., BRUGUÉS, M., CANO, M.J. & CROS, R.M. (eds.) (2010). Flora Briofítica Ibérica. Vol. IV, Funariales, Splachnales, Bryales, Timmiales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 317 pp.
- GUERRA, J., CANO, M.J. & BRUGUÉS, M. (eds.) (2014). Flora Briofítica Ibérica. Vol. V, Orthotrichales, Hedwigiales, Leucodontales, Hookeriales. Universidad de Murcia, Sociedad Española de Briología, Murcia, 261 pp.
- GUERRA, J., CANO, M.J. & BRUGUÉS, M. (eds.) (2016). *Flora Briofítica Ibérica. Vol. VI, Hypnales.* Universidad de Murcia, Sociedad Española de Briología, Murcia, 463 pp.
- Keys and detailed descriptions (in German), some photographs
- NEBEL, M. & PHILIPPI, G. (eds.) (2000). *Die Moose Baden-Württembergs, I.* (*Andreaeales bis Funariales*). Ulmer, Stuttgart, 512 pp.

- NEBEL, M. & PHILIPPI, G. (eds.) (2001). *Die Moose Baden-Württembergs, II.* (*Schistostegales bis Hypnobryales*) Ulmer, Stuttgart, 529 pp.
- NEBEL, M. & PHILIPPI, G. (eds.) (2005). *Die Moose Baden-Württembergs, III.* (*Sphagnopsida, Marchantiopsida, Anthocerotophyta*) Ulmer, Stuttgart, 487 pp.
- Short descriptions (in English) and original drawings mainly by the author, often showing very useful details
- Nyholm, E. (1960). *Illustrated Moss Flora of Fennoscandia II. Musci. Fasc. 4.* Swedish Natural Science Research Council, Lund, pp. 284–408.
- Nyholm, E. (1979). *Illustrated Moss Flora of Fennoscandia II. Musci. Fasc. 5, ed. 2.* Swedish Natural Science Research Council, Stockholm, pp. 407–647.
- NYHOLM, E. (1981). *Illustrated Moss Flora of Fennoscandia II. Musci. Fasc. 6, ed. 2.* Swedish Natural Science Research Council, Stockholm, pp. 647–799.
- NYHOLM, E. (1987). *Illustrated Flora of Nordic Mosses. Fasc. 1. Fissidentaceae Seligeriaceae.* Nordic Bryological Society, Copenhagen and Lund, pp. 1–72.
- NYHOLM, E. (1990). *Illustrated Flora of Nordic Mosses. Fasc. 2. Pottiaceae Splachnaceae Schistostegaceae.* Nordic Bryological Society, Copenhagen and Lund. pp. 75–141.
- Nyholm, E. (1993). *Illustrated Flora of Nordic Mosses. Fasc. 3.*Bryaceae Rhodobryaceae Mniaceae Cinclidiaceae Plagomniaceae. Nordic Bryological Society, Copenhagen and Lund, pp. 145–244.
- Nyholm, E. (1998). *Illustrated Flora of Nordic Mosses, Fasc.* 4 (Aulacomniaceae Meesiaceae Catascopiaceae Bartramiaceae Timmiaceae Encalyptaceae Grimmiaceae Ptychomitraceae Hedwigiaceae Orthotrichaceae). Nordic Bryological Society, Copenhagen and Lund, pp. 249–404.

PART I: GENERAL KEYS

1. KEY TO THE MAIN GROUPS OF BRYOPHYTES

1	Plants thallose, i.e. a more or less flat gametophyte, not
	differentiated into stem and leaves2
-	Plants not thallose, differentiated into stems and leaves3
2	Thallus cells usually with a single, large chloroplast; capsule
	cylindrical, dehiscing by 2 valves
-	Thallus cells with numerous chloroplasts; capsule globose, ovoid, ellipsoid or cylindrical, dehiscing by 4 valves or
	irregular
3	Leafy liverworts
	Mosses

Differences between liverworts and mosses

liverworts

gametophyte a thallus leaves deeply lobed or segmented

leaves never with costa (but beware of vitta in *Diplophyllum albicans*) leaves without costa, cells isodiametric, never prosenchymatic leaves arranged in 3 ranks, 2 ranks of lateral leaves and 1 rank of (smaller) underleaves, which may be lacking rhizoids consisting of a single cell each

seta weak, hyaline, elongating rapidly and dying after a few days

spore capsule containing spores and elaters, but without peristomium

capsule globose or ovoid, without lid, splitting into 4 valves capsule blackish, without chloloplasts and wall without stomata

mosses

gametophyte never a thallus leaves never deeply lobed or segmented leaves with or without costa

leaves without costa, cells prosenchymatic, never isodiametric leaves not in 3 or 2 ranks (but beware of Fissidens, Distichium: leaves in 2 ranks, but leaves costate) rhizoids made up of more than one cell, with oblique cross-walls seta stout, brown, red, yellow, elongating slowly and long persisting, usually visible for a year spore capsule containing spores only, with peristomium in the majority of cases

capsule pear-shaped to cylindrical, typically with lid and peristome capsule green with chloroplasts when young, usually brown when ripe, wall often with stomata

2. KEYS TO GENERA OF THALLOID BRYOPHYTES

Н	or	'n	LA7	n	rt	c
	u		w	.,		

-	Thallus variously lobed, lobes with crispate margins; spores dark brown or blackish; mature antheridia 50–90 µm long
-	Thallus lobed, but margins only rarely crispate; spores yellowish, rarely yellowish brown; proximal face of spores with weak, centrally placed papillae
Tha	lloid liverworts
1	Thallus circular, completely covered by clavate involucres on dorsal side
-	Thallus not as above, not covered completely by clavate involucres
2	Dorsal epidermis reticulate, with pores or irregular openings, and usually with air chambers visible in cross section; rhizoids dimorphic, smooth or warty, respectively; thallus with chlorenchyma and parenchyma differentiated
-	Dorsal epidermis not reticulate, without pores or air chambers; rhizoids smooth; thallus not differentiated into chlorenchyma and parenchyma
	<i>up 1</i> Thallus with internal differentiation; dorsal epidermis h pores; rhizoids smooth or warty
1	Dorsal side of thallus with gemma receptacles2
-	Dorsal side of thallus without gemma receptacles
2	Gemma receptacles semilunar
3	Gemma receptacles cup-shaped

-	Plants not in rosettes; chlorenchyma with air chambers with simple or barrel-shaped pores; capsule not immersed in thallus.
4	Pores barrel-shaped (cross section), conspicuous
_	Pores simple, conspicuous or not
5	Epidermis conspicuously reticulate, with elongate areolae 1
	mm or longer
-	Epidermis conspicuously reticulate, with areolae less than 1
_	mm long, or reticule lacking
6	Thallus surface indistinctly reticulate, when dry parchment-
	like, smooth except for the slightly elevated, simple pores;
	ventral scales metallic purple, with 2 filiform appendages;
	pores prominent, surrounded by 4–5 concentric rings of cells
	Thallus distinctly reticulate; ventral scales coloured or
-	hyaline, with 1–2 lanceolate appendages; pores prominent
	or not, surrounded by 1–5 concentric rings of cells7
7	Ventral scales lacking marginal slime hairs, triangular,
,	hyaline, with a lanceolate appendage; air chambers high,
	obliquely in 2–3 layers
_	Ventral scales with marginal 1-celled slime hairs; air
	chambers low, vertically in 1–3 layers8
8	Archegonia and sporophytes not surrounded by laciniate
	pseudoperianth; crushed thallus with or without smell of
	cedar oil; dorsal thallus epidermis with or without trigones
-	Archegonia and sporophytes surrounded by laciniate
	pseudoperianth; crushed thallus with a faint smell of rotten
	freshwater fish; dorsal thallus epidermis without trigones
Grou	p 2 Thallus without internal differentiation, dorsal
	ermis without pores, rhizoids smooth
1	Thallus with obliquely inserted crisped or undulate leaf-like
1	lobes
-	Thallus without leaf-like lobes 3

2	Gemmae dimorphic, (i) stellate, about 400 μm , (ii) globose or ellipsoidal, about 100 μm , produced in flask-like receptacles
	on the dorsal surface of the thallus
-	Gemmae lacking
3	Thallus furcate, to 2 mm wide, with unicellular hairs at margins, on ventral or on both sides, sometimes glabrous; midrib narrow, clearly distinct on the dorsal side; gametangia and sporophytes on small branches on ventral
-	side of midrib
	4
4	Thallus furcate; gametangia on dorsal side of thallus5
-	Thallus simple, pinnate, palmate or irregularly branched; gametangia on small lateral branches6
5	Slime papillae restricted to ventral side of costa near apex, on stalk 1–5 (6) cells long; thalli with miniature apical branches proliferating in autumn; dioicous; involucres subhorizontal to erect, tubular, with dentate-ciliate lobes; male tubercles <i>ca</i> 200 µm wide, cells surrounding aperture not papilliform; calcicole
_	Slime papillae on stalk 1 cell long, on both ventral and dorsal
	sides of costa near apex; caducous branches absent; dioicous or monoicous; calcifuge
6	Thallus simple or irregularly branched, 3 mm wide or more; more than 6 oil bodies in each epidermal cell
-	Thallus 1–4-pinnately branched or palmate, lobes < 2 mm wide; 0–2 oil bodies in each epidermal cell <i>Riccardia</i> (p.109)
3. KE	YS TO GENERA OF FOLIOSE LIVERWORTS
1	Leaves laciniate, divided almost to base into filiform segments, or leaves with longly ciliate or laciniate margins
-	Leaves simple or lobed, without laciniate or ciliate margins
	4

2	Leaves laciniate, divided almost to base into 3–4 uniseriate segments		
	Leaves divided into 2–5 lobes that are not uniser		
-	longly ciliate or laciniate margins		
3	Leaf lamina very reduced		
.	Leaf lamina well developed <i>Ptilidium pulcherri</i>		
4 (1)			
* (1)	Leaves lobed		
5	Leaves incubous		
-	Leaves succubous		
6	Leaf apex rounded to bilobed; underleaves bilobe	ed, retuse,	
	emarginate or rounded	eia (p.98)	
-	Leaf apex truncate, usually with 1–3 teeth; un orbicular quadrate, irregularly 4–5-lobed		
	Bazzania tri		
7(4)		. ,	
_	Leaves bilobed	-	
8	Leaves conduplicate, ventral lobe sometimes sac-		
	helmet-shaped	-	
_	Leaves not conduplicate	10	
9	Underleaves lacking		
-	Underleaves present	Group 6	
10	Leaves transversely or subtransversely inserted	Group 7	
-	Leaves obliquely or longitudinally inserted, succub	ous	
		Group 8	
Grou	up 3 Leaves simple, succubous		
urou	ip 5 Leaves simple, succubous		
1	Underleaves present, conspicuous, bilobed		
-	Underleaves lacking, minute or small, rarely bilobed		
2	All leaves simple, with rounded or retuse apex		
	Chiloscyphu		
-	At least some lower leaves bilobed		
3	Leaf margin toothed, at least at apex		
-	Leaf margin entire		
4	Underleaves present, at least on young plants		
_	Underleaves lacking or caducous		
5	Lower leaves spreading, apical leaves imbricate, face to face; perianth mouth laciniate		
	Svzvaiella autumnalis (lamesoniella autum		

-	Leaves not as above; perianth mouth at most crenulate		
_			
6	Leaves strongly obliquely to longitudinally inserted		
-	Leaves transversely to obliquely inserted		
7	Plants prostrate; rhizoids only at stem apex; leaves plane		
	Pedinophyllum interruptum (w)		
-	Plants procumbent or ascending; rhizoids lacking or scarce		
0	on underground stems; leaves convex <i>Plagiochila</i> (p.106)		
8	Lower leaves spreading, apical leaves imbricate, pressed		
	face to face; perianth mouth laciniate		
-	Leaves not as above; perianth mouth at most crenulate		
	Jungermanniaceae group (incl. Endogemma,		
	Jungermannia, Liochlaena, Solenostoma) (p.101)		
Grou	p 4 Leaves 3–5-lobed		
arou	p i beaves 5 5 lobea		
1	Leaves succubous		
-	Leaves incubous or nearly transverse3		
2	Lower leaves bilobed, upper leaves sometimes irregularly 3-		
	5-lobed Lophoziaceae group (p.89)		
-	All leaves 3-4-lobed		
	Barbilophozia group (incl. Neoorthocaulis floerkei) (p.85)		
3	Underleaves lacking		
	<i>Tritomaria</i> s.l. (incl. <i>Trilophozia quinquedentata</i>) (p.89)		
-	Underleaves present, similar to leaves, although smaller;		
	leaves incubous		
_			
Grou	p 5 Leaves conduplicate, underleaves lacking		
1	Dorsal lobe larger than ventral lobe2		
_	Dorsal lobe similar in size to ventral lobe or smaller		
2	Plants medium-sized, shoots <i>ca</i> 2 mm wide; rhizoids		
_	originate from the lobule		
_	Plants minute, shoots < 0.5 mm wide; rhizoids originate		
	from the stem		
3	Ventral lobe narrowly lingulate, twice as long as wide;		
-	gemmae when present angulate		
	6.5.1)		

-	Ventral lobe orbicular to widely lingulate, less than twice as long as wide; gemmae when present ovoid to ellipsoidal
Gro	up 6 Leaves conduplicate, underleaves present
1	Ventral lobe with spinosely toothed margin
_	Ventral lobe with entire margin
2	Lobule parallel or nearly parallel to stem, almost free from stem; keel very short; shoots mostly large, 2.5–5 mm; cells with few or numerous oil bodies
-	Lobule parallel to ventral leaf margin; keel long; shoots 0.9–1.3 mm wide; cells with numerous small oil bodies
_	Lejeunea cavifolia (w)
3	Ventral lobe mostly sac-shaped or helmet-shaped (but evolute in <i>Frullania cleistostoma</i>); ocelli (cells with one large oil body) present in dorsal lobe or not, cells with few oil bodies
-	Ventral lobe plane or convex; ocelli absent, cells with numerous (15–40) oil bodies
	up 7 Leaves bilobed, transversely or subtransversely or transversely
1	Underleaves present2
_	Underleaves absent or very small 3
2	Plants filiform or very small, leaves to 0.3 mm long
-	Plants not filiform, small to large, leaves to 1.5 mm long
3	Leaves appressed; branches julaceous or dorsiventrally compressed, often clavate
-	Leaves not appressed; branches not as above4
4	Leaves asymmetrical, with ventral margin widely incurved and saccate
-	Leaves not or only slightly asymmetrical; ventral margin not saccate

5	Plants filiform, very small; usually ventrally branched
-	Plants not filiform, small to medium-sized; laterally branched
6	Leaves channelled or strongly concave7
_	Leaves not channelled, ± concave
7	Usually with reddish gemmae at margins of upper leaves8
_	Gemmae lacking
8	Minute species, plants only 2–10 (12) mm long and 0.2 mm
	wide; lateral leaves slighty concave, ± transversely inserted,
	2-lobed to 1/3-1/2, sometimes 3-lobed, lobes lanceolate,
	acute; cell walls uniformly thickened or trigones
	occasionally present; underleaves absent or sparse, minute,
	subulate; perianths rare; on rotting wood and bark; stolons
	absent
	Crossocalyx hellerianus (Anastrophyllum hellerianum) (rr)
-	Larger species; if shoots only 2-6 mm long, then plants
	wider than 0.2 mm or lateral leaves lobed to 2/3-3/4;
	(compare Lophozia ascendens on rotting wood)9
9	Lateral leaves conduplicate-concave (boat-shaped), nearly
	transversely inserted, 0.3–0.7 mm long; shoots 0.7–1.5 mm
	wide, leaves with regular pectinate arrangement; walls of
	leaf cells almost equally thickened, trigones very small or
	absent; mid-leaf cells (12) 14-18 (20) µm wide, arranged in
	± concentric circles near lobe apex; gemmae angulate, 2-
	celled
-	Lateral leaves as long as wide, transversely inserted on
	antical side of stem, obliquely on postical side, (0.5) 0.7–1.8
	mm long, 2-lobed to 1/3, lobes acute, equal, slightly
	squarrose; plants 2.5 cm long and 1–2 mm wide; walls of
	leaf cells thick, trigones small to large, not bulging; shoots
	brownish green; gemmae ovoid to angulate
10(()	Anastrophyllum michauxii (n.s.)
10 (6)	Usually with gemmae at apex and margins of upper leaves;
	cortex not translucent, not revealing the medulla; living
	plants with oil bodies Lophoziaceae group (p.89) Usually without gemmae; cortex translucent, revealing the
_	medulla; living plants without oil bodies
	Cephaloziaceae pp. (p.86)
	Cephaloziaceae pp. (p.00)

${\it Group~8}$ Leaves bilobed, longitudinally or obliquely inserted, succubous

1	Underleaves conspicuous
-	Underleaves lacking or underleaves small, subulate or
	lanceolate3
2	Rhizoids restricted to base of underleaves; underleaves
	bilobed 1/2-3/4, with teeth on each side. <i>Lophocolea</i> (p.104)
_	Rhizoids scattered along the ventral surface of stem
	underleaves simple, lanceolate, usually with lateral teeth
3	Plants very small, filiform
_	Plants small to medium-sized4
4	Cortex translucent, revealing the medulla; living plants
т	without oil bodies Cephaloziaceae pp. (p.86)
	Cortex not translucent, not revealing the medulla; living
_	plants with oil bodies5
5	Plants with gemmaeLophoziaceae group (p.89)
3	Plants without gemmae
6	9
0	Underleaves small 7
- 7	Underleaves lacking or rudimentary
/	Cuticle smooth Nardia (p.100)
-	Cuticle papillose
8	Plants acidophilous, perianths usually caducous; underleaves
	absent
-	Plants calcicolous, perianths not caducous; underleaves
	present (except in <i>M. badensis</i>)
4. KI	EYS TO GENERA OF MOSSES
1	Leaf lamina consisting of a network of narrow green cells
	enclosing large, inflated, hyaline cells; plants with branches
	in fascicles
-	Plants lacking above combination of characters2
2	Plants dark brownish, reddish or blackish; saxicolous;
	fragile when dry; capsule opening with 4 longitudinal slits
	giving rise to 4 valves united at the top; lamina cells thick-
	walled reddish to brownish Andregeg runestris (r) (n 126)

-	Plants lacking above combination of characters	
3	Basal lamina cells elongate, thick-walled, sinuos	
	Racomit	
-	Basal lamina cells different	4
4	Capsule with peristome of 4 triangular teeth, en	ect when dry
	and moist	
-	Capsule with peristome of more than 4 tee	th, or plants
	without capsules	
5	Stem leaves arranged in 2 rows, distichous, ± co	mplanate
_	Stem leaves arranged in 3 or more rows, not co	mplanate6
6	Ventral (adaxial) surface of costa or lamina with	h filaments or
	lamellae	
_	Ventral surface of costa or lamina without	
	lamellae	
7	At least upper leaves with a hyaline tip or costa	
	hair point	
_	Leaves without hyaline tip or hair point	
8	Leaves bordered with several rows of elongate	
	or leaf margin pluristratose	
_	Leaves unbordered, leaf margin 1–2-stratose	
9	Plants acrocarpous	
_	Plants pleurocarpous	
10	Capsule present	
_	Capsule lacking	
11	Capsule indehiscent	
_	Capsule dehiscent	
12	Capsule immersed or emergent	
_	Capsule exserted	
13	Plants minute, capsule larger than plant, ovoice	
	asymmetric, on warty stalk	
_	Plants lacking this combination of characters	
14	Capsule globose or sub-globose	
_	Capsule ovoid, oblong, cylindrical, ellipsoid or p	
15	Capsule strumose	
_	Capsule not strumose	
16	Peristome absent or rudimentary	
_	Peristome well developed	-
17	Capsule striate or sulcate when dry	
_	Capsule smooth or only very slightly striate wh	
	1	, ,

18	Capsule erect or nearly so; seta straight, rarely curved19	
-	Capsule inclined or pendulous; seta straight or curved20	
19	Peristome teeth 16, entire or divided at the tip only, o	
	slightly and irregularly divided	
-	Peristome teeth 16, divided longitudinally to halfway o	
	more (32 teeth)Group 20	
20	O Peristome single	
_	Peristome doubleGroup 22	
21 (10)	Propagules present on stem or leaves or in receptacles	
	Group 23	
_	Propagules absent on stem or leaves or in receptacles22	
22	2 Leaves without costa	
_	Leaves with costa 23	
23	Group 24 Costa wide, 1/3 or more of leaf baseGroup 24	
-	Costa narrow, less than 1/3 of leaf base24	
2 4	4 Lamina cells 18 μm wide or moreGroup 25	
_	Lamina cells less than 18 µm wide25	
25	Alar cells differentiated from basal cells	
-	Alar cells not differentiated	
26	5 Lamina cells ± isodiametric27	
-	Lamina cells longer than wide31	
27	7 Leaf margin denticulate or dentate, at least near apex of	
	baseGroup 27	
-	Leaf margin entire, crenulate or papillose-crenulate28	
28	Group 28	
-	Costa not excurrent	
29	· · · · · · · · · · · · · · · · · · ·	
-	Leaf apex acute or acuminate30	
30	1	
-	Leaf margin plane or recurved only at baseGroup 31	
31 (26)		
	by costa	
-	Leaf apex obtuse to acuminate, costa percurrent to	
	excurrent, short or lackingGroup 33	
32 (9)		
-	Costa single, extending less than 1/2 way up leaf, or i	
	double, short or long, or lacking	
33		
	elongate and marginal cells short	
_	Lamina cells elongate more than twice as long as wide 34	

34	Leaves longitudinally plicate, at least at base	5
-	Leaves plane or only slightly longitudinally plicate3!	5
35	Leaves squarrose, falcate or circinate	6
-	Leaves straight or nearly so	6
36	Leaf apex rounded, obtuse or apiculateGroup 3	7
_	Leaf apex acute or acuminate Group 3	8
37 (32)	Lamina cells short, to twice as long as wide, or median cell	ls
	elongate and marginal cells short	9
_	Lamina cells elongate, more than twice as long as wide38	8
38	Leaf apex rounded, obtuse or apiculateGroup 4	0
_	Leaf apex acute or acuminate	
39	Leaves distinctly falcate or squarrose	
_	Leaves straight or very slightly falcate or squarrose	
	Group 42	2
Gro	up 9 Leaves distichous or complanate	
1	Plants acrocarpous	2
-	Plants pleurocarpous	3
2	Lower part of leaf conduplicate	7)
-	Leaf not conduplicate, narrowed to subulate point	
	Distichium capillaceum (w)
3	Median lamina cells 60-80 μm wide	.)
-	Median lamina cells 4–45 μm wide	
4	Alar cells differentiated, hyaline, decurrent	
)
-	Alar cells greenish or yellowish, differentiated or not	5
5	Costa single	6
-	Costa double or short and lacking	7
6	Leaves oblong to spathulate, costa extending 1/2 - 3/4 u	p
	leaf	7)
-	Leaves lanceolate or ovate-lanceolate, costa extending 2/	3
	up leaf to almost to apex	:)
7	Leaves mostly oblong-ovate to oblong or lingulate	8
_	Leaves mostly lanceolate, ovate or ovate-lanceolate	9
8	Median lamina cells 8–80 μm long	
)
_	Median lamina cells 45–160 μm long	
	Tavinhyllum wiccarillii (w	

9	Flagelliform branchlets in leaf axils		
-	Flagelliform branchlets absent, leaves only slightly complanate		
	up 10 Costa or ventral leaf surface with filaments or ellae		
1	Costa with simple or branched filaments, at least in distal part2		
-	Costa with lamellae, at least in distal part, sometimes with a few filaments		
2	Leaf margin widely incurved, leaves rigid		
3	Plants to 0.5 (1) cm tall; costa with 2–4 lamellae; peristome rudimentary or lacking		
-	Plants 0.5–6 cm tall; costa with 4 or more lamellae; peristome well developed, with 32 or 64 teeth (Polytrichaceae)		
4	Costa narrow, with 4–7 ventral lamellae; leaves with a border of long narrow cells		
-	Costa broad, with more than 7 lamellae on the ventral surface; leaves unbordered		
5	Capsule not angled, without apophysis and stomata, peristome teeth 32; leaves muticous		
-	Capsule angled or not, with apophysis and stomata; peristome teeth 64; leaves muticous or with hair point		
6	Capsule not angled; end cell of lamellae papillose, higher than wide. <i>Polytrichastrum alpinum</i> (<i>Polytrichum alpinum</i>) (n.s.)		
-	Capsule angled; end cell of lamellae smooth (p.126)		
<i>Gro</i> i poir	up 11 Leaves with hyaline point or costa excurrent in hair nt		
1	Pleurocarps with papillose cells, hyaline point only visible in microscope		
-	Acrocarps, hyaline point visible to the naked eye or with a handlens3		

2	Plants regularly 2–3-pinnate; stem leaves with a long fine		
	acumen, consisting of 3–4 uniseriate cells		
	Thuidium assimile (Thuidium philibertii) (w)		
-	Plants irregularly branched, stem leaves with a long,		
	uniseriate hyaline point		
_			
3	Costa lacking		
-	Costa present4		
4	Plants julaceous, silvery when dry Bryum argenteum (cc),		
	Ptychostomum zieri (Plagiobryum zieri) (r)		
-	Plants not julaceous, not silvery when dry5		
5	Capsule pale yellow, ± smooth, emergent; exostome of 16		
	free teeth, reflexed when dry Orthotrichum diaphanum (cc)		
-	Capsule and peristome different6		
6	Costa wide, 1/2–1/3 of width of leaf base, hair point reflexed		
	when dry		
_	Costa narrow, less than 1/3 of width of leaf base7		
7	Lamina cells smooth or finely papillose8		
-	Lamina cells strongly papillose12		
8	Capsule immersed or emergent9		
-	Capsule exserted10		
9	Capsule indehiscent, calyptra cucullate		
	(<i>Tortula acaulon</i> pp.) (p.166)		
-	Capsule dehiscent, calyptra cucullate, mitrate or campanulate		
	Grimmiaceae pp. (p.175)		
10	Lamina cells smooth in median part of leaf, papillose on		
	strongly revolute margins		
_	Lamina cells smooth throughout		
11	At least some lamina cells sinuoseGrimmiaceae pp. (p.175)		
_	Lamina cells not sinuose		
12 (7)			
	cells on both sides of costa		
_	Basal cells of leaf not forming a distinct ovate group on both		
	sides of costa		
13	Peristome teeth not spirally twisted or lacking; calyptra		
	covering the whole capsule; basal hyaline cells with pale		
	orange cross walls distinctly thickened		
_	Peristome teeth spirally twisted; calyptra partially covering		
	the capsule; basal cells different		
	· • • • • • • • • • • • • • • • • • • •		

14	Capsule immersed, indehiscent	<i>Microbryum</i> pp. (p.151),
	<i>Tortula</i> pp. (<i>To</i>	o rtula acaulon pp.) (p.166)
-	Capsule exserted or emergent, del	niscent <i>Tortula</i> pp. (p.166)

Group 12 Leaves bordered with narrow cells or margin pluristratose

1	Marginal cells short; margin 2–5-stratose
_	Marginal cells longer and narrower than the rest of lamina
	cells; margin uni- to pluristratose
2	Leaf margin recurved; lamina cells strongly papillose;
	central strand well developedDialytrichia mucronata
_	Leaf margin plane; lamina cells smooth or faintly papillose;
	central strand absent
3	Leaf margin unistratose
_	Leaf margin bi- to pluristratose
4	Upper and median lamina cells papillose
_	Upper and median lamina cells smooth
5	Plants rhizomatous; leaves crowded in terminal rosette
_	Plants not rhizomatous; leaves crowded in terminal rosette
	or not6
6	Sterile shoots often creeping; leaf margin dentate or
	(sometimes only faintly) denticulate <i>Plagiomnium</i> (p.201)
_	Sterile shoots erect; leaf margin ± denticulate, rarely entire
	Bryum s.l. (incl. Imbribryum, Ptychostomum) (p.190)
7 (3)	Leaf margin entire; leaves less than twice as long as wide
	Rhizomnium punctatum (w)
-	Leaf margin entire below, denticulate towards apex, or
	dentate ± all round; leaves more than twice as long as wide
	8
8	Lamina cells papillose, sometimes only faintly so; leaf
	margin entire below, denticulate towards apex
-	Lamina cells smooth; leaf margin dentate9
9	Leaf margin with simple teeth
-	Leaf margin with geminate teeth

Group 13 Acrocarps with indehiscent capsule

1	Capsule exserted
	(Pottia bryoides, Protobryum bryoides) (w)
-	Capsule immersed or emergent 2
2	Capsule with conspicuous neck, $1/3-1/2$ of capsule length;
	capsule emergent
-	Capsule without perceptible neck
3	Plants with persistent protonema
-	Plants without persistent protonema4
4	Lamina cells smooth, more than 25 μm wide
	Physcomitrium patens
	(Aphanorrhegma patens, Physcomitrella patens) (w)
-	Lamina cells smooth or papillose, less than 25 μm wide5
5	Capsule with ± differentiated lid; leaves incurved, crisped
	when dry
-	Capsule without lid; leaves not crisped when dry6
6	Capsule globose, with translucent walls, spores mostly 16-
	20 in number, 100–200 μm <i>Archidium alternifolium</i> (n.s.)
-	Capsule globose or ± apiculate, with opaque wall, spores
	numerous, < 60 μm
7	Lamina cells rectangular or linear8
-	Lamina cells quadrate, rounded, hexagonal or elliptical9
8	Leaves subulate, at least the upper ones; lamina cells 8 μm
	wide
-	Leaves acuminate; lamina cells 9–13 µm wide
	Pseudephemerum nitidum (w)
9	Plants bulbiform; costa with stereids only; capsule not or
	only minutely apiculate
-	Plants not bulbiform; costa with stereids and guide cells;
	capsule apiculate
	<i>Tortula</i> pp. (p.166)
Grou	p 14 Acrocarps with immersed or emergent capsule
1	Capsule large, ovoid, asymmetrical; perichaetial leaves
	ciliate
-	Capsule and perichaetial leaves not as above2
2	Leaves without costa
-	Leaves costate

3	Lid coming off attached to columella; capsule smooth
-	Lid not coming off attached to columella; capsule smooth or striate.
4	Plants minute; growing on calcareous rocks; capsule emergent to exserted
-	Plants medium-sized; growing on calcareous or acidic rocks or on tree trunks
5	Leaf cells mamillose; capsule globose
	Bartramia halleriana (rr)
6	Leaf cells papillose or smooth; capsule shortly cylindrical6 Stomata phaneropore
-	Stomata cryptopore
7	Leaves concave, with erect-incurved or involute margins
	and obtuse or rounded apices, gemmae abundant, in particular on the ventral surface of leaves; capsules rare
-	Leaves not as above, with recurved margins and acute to
	acuminate apices; gemmae occasional; capsules frequent
<i>Grou</i> caps	p 15 Acrocarps with exserted globose or subglobose
caps	p 15 Acrocarps with exserted globose or subglobose ule
	<i>p 15</i> Acrocarps with exserted globose or subglobose ule Peristome lacking (Funariaceae pp.)2
caps	p 15 Acrocarps with exserted globose or subglobose ule
caps	Lewinskya (p.203) p 15 Acrocarps with exserted globose or subglobose ule Peristome lacking (Funariaceae pp.) 2 Peristome single or double (Bartramiaceae) 3 Calyptra inflated, contracted at base, cucullate, covering all of the capsule Pyramidula tetragona (r)
caps	Peristome lacking (Funariaceae pp.) 2 Peristome single or double (Bartramiaceae) 3 Calyptra inflated, contracted at base, cucullate, covering all of the capsule Pyramidula tetragona (r) Calyptra small, mitriform, not covering all of the
1 - 2 -	Peristome lacking (Funariaceae pp.) 2 Peristome single or double (Bartramiaceae) 3 Calyptra inflated, contracted at base, cucullate, covering all of the capsule Pyramidula tetragona (r) Calyptra small, mitriform, not covering all of the capsule Physcomitrium (eurystomum, sphaericum) (p.131)
caps	Peristome lacking (Funariaceae pp.)
1 - 2 -	Peristome lacking (Funariaceae pp.) 2 Peristome single or double (Bartramiaceae) 3 Calyptra inflated, contracted at base, cucullate, covering all of the capsule Pyramidula tetragona (r) Calyptra small, mitriform, not covering all of the capsule Physcomitrium (eurystomum, sphaericum) (p.131) Leaves lanceolate, not sheathing at base Philonotis (p.189) Leaves narrowly lanceolate to linear lanceolate, sheathing at
1 - 2 -	Peristome lacking (Funariaceae pp.) 2 Peristome single or double (Bartramiaceae) 3 Calyptra inflated, contracted at base, cucullate, covering all of the capsule Pyramidula tetragona (r) Calyptra small, mitriform, not covering all of the capsule Physcomitrium (eurystomum, sphaericum) (p.131) Leaves lanceolate, not sheathing at base Philonotis (p.189) Leaves narrowly lanceolate to linear lanceolate, sheathing at base 4
caps 1 - 2 - 3 -	Peristome lacking (Funariaceae pp.) 2 Peristome single or double (Bartramiaceae) 3 Calyptra inflated, contracted at base, cucullate, covering all of the capsule Pyramidula tetragona (r) Calyptra small, mitriform, not covering all of the capsule Physcomitrium (eurystomum, sphaericum) (p.131) Leaves lanceolate, not sheathing at base Philonotis (p.189) Leaves narrowly lanceolate to linear lanceolate, sheathing at

Group 16 Acrocarps with exserted strumose capsule

1	Costa with a central layer of green cells surrounded by hyaline cells (cross section); plants whitish or glaucous
_	Costa not as above; plants not whitish or glaucous
2	Peristome teeth divided near to base, reddish, with a paler border or not
-	Peristome teeth divided to halfway, reddish, striate in the lower part
3	Lamina cells longly rectangular, smooth; margin plane
-	At least upper lamina cells quadrate or oblate, mamillose; margin recurved
	up 17 Acrocarps with exserted capsule and peristome ing or rudimentary
1	Calyptra inflated, 4-angled; spores 50–65 μm
	Pyramidula tetragona (r)
-	Calyptra not inflated or angled; spores smaller2
2	Capsule striate or sulcate when dry3
-	Capsule smooth when dry4
3	Leaves linear-lanceolate; gemmae lacking
	Amphidium mougeotii (r)
-	Leaves lanceolate to oblong-lanceolate; plants with ovoid
	gemmae; lamina cells papillose
4 (2)	Median lamina cells 20–50 μm wide, thin-walled
	(Funariaceae pp.)5
-	Median lamina cells less than 20 µm wide, thin-walled or thick-walled
5	Exothecial cells longer than wide
	Entosthodon hungaricus (w)
_	Exothecial cells isodiametric6
6	Calyptra mitriform
-	Calyptra cucullate
7(4)	Plants minute; lamina cells smooth <i>Seligeria donniana</i> (r)
_	Plants small to robust; lamina cells papillose8
8	Calyptra cylindrical, covering capsule; basal leaf cells
	forming a well-delimited group

	-	Calyptra cucullate, not covering capsule; basal leaf cells not
		forming a well-delimited group9
	9	Costa excurrent 10
	-	Costa not excurrent 12
	10	Leaves linear-lanceolate or lanceolate; margin incurved to
		strongly involute, rarely plane
	-	Leaves ovate-lanceolate, ovate or obovate; margin plane or
		recurved11
	11	Lid conical <i>Microbryum</i> (davallianum, muticum) (p.151)
	-	Lid rostrate
12	(9)	Lid attached to columella after dehiscence
	_	Lid not attached to columella after dehiscence
	13	Plants 1-4 (10) cm tall; leaves carinate; margin finely
		crenulated
	_	Plants to 1 cm tall; leaves not carinate; margin dentate at
		apex
	14	Annulus of capsule persistent, of large cells; stems rarely
		branched <i>Gyroweisia tenuis</i> (rr)
	_	Annulus of capsule persistent or caducous, of small cells;
		stems branched <i>Gymnostomum</i> (p.170)
		ų ,
	Grou	p 18 Acrocarps with exserted capsule, striate or sulcate
		n dry and peristome well developed
	1	Costa with a central layer of green cells surrounded by
		hyaline cells (cross section); plants whitish or glaucous
		<i>Leucobryum</i> (p.135)
	_	Costa not as above; plants glaucous or not
	2	Plants very small, to 3.5 mm tall
		Brachydontium trichodes (rr)
		Di actiyaditatii a ichdaes (11)
	-	Plants more than 3.5 mm tall
	- 3	Plants more than 3.5 mm tall
	- 3 -	Plants more than 3.5 mm tall 3 Seta curved 4
	- 3 - 4	Plants more than 3.5 mm tall
	-	Plants more than 3.5 mm tall
	-	Plants more than 3.5 mm tall 3 Seta curved 4 Seta straight 6 Costa occupying 1/3 or more of leaf base Campylopus (p.133)
	-	Plants more than 3.5 mm tall 3 Seta curved 4 Seta straight 6 Costa occupying 1/3 or more of leaf base Campylopus (p.133) Costa occupying less than 1/3 of leaf base 5
	- 4 -	Plants more than 3.5 mm tall 3 Seta curved 4 Seta straight 6 Costa occupying 1/3 or more of leaf base [p.133] Costa occupying less than 1/3 of leaf base 5 Leaves ovate-lanceolate; upper lamina cells rectangular or
	- 4 -	Plants more than 3.5 mm tall

-		lmina cells quadrate or rounded, L
6		groups of 4 (Orthotrichaceae pp.)
		7
-		rming groups12
7		8
-		9
8		Orthotrichum (p.203)
_		Lewinskya (p.203)
9		se rectangular, with thickened
		up margin, basal cells linear
		<i>Ulota</i> (p.203)
-	0	e, similar to the rest of basal cells
10	Lamina colle emooth, calunt	10 ra cucullate, glabrous
10		ron forsteri (Zygodon forsteri) (r)
	_	se; calyptra mitrate, often hairy
_		<i>ichum</i> s.l. (incl. <i>Lewinskya</i>) pp 11
11		
11		Lewinskya (p.203)
- 12 (6)		Lewiiiskyu (p.203)
		20
13		ntire or slightly and irregularly
13		14
_		ed to halfway or more16
14	The state of the s	completely by the calyptra
1-1		
_		lindrical, not completely covered
		15
15		
_	0	
16		pase17
_		nalfway18
17		Saelania glaucescens (rr)
_	0	Ceratodon (p.143)
18		Dicranum (p.139)
_		19
19		
_		Dicranella (p.135)

20 (12)	Median cells of lamina linear, smooth
	Orthodontium lineare (rr)
-	Median cells of lamina rounded, quadrate or hexagonal, mammilose or papillose21
21	Leaves with dentate margins and sheathing base; marginal cells narrower and longer than median cells; plants without gemmae
-	Leaves with entire or denticulate margins and the base not sheathing; marginal cells similar to median cells; gemmae in groups at the end of pseudopodiaAulacomnium (p.211)
	up 19 Acrocarps with exserted erect capsule, peristome th 16, entire or slightly and irregularly divided
1	Lamina cells more than 20 μm wide, smooth
_	Lamina cells less than 20 µm wide, smooth or papillose2
2	Leaf apex acuminate or subulate, very rarely obtuse
_	Leaf apex obtuse, acute or rounded, rarely acuminate
3	Alar cells differentiated, orange, brownish red
_	Alar cells not differentiated4
4	Leaves fragile; margin sinuose-undulate, notched
-	Leaves not fragile; margin not sinuose, not notched5
5	Plants minute; growing on calcareous or siliceous rocks6
-	Plants medium-sized; growing on acid, volcanic rocks or tree trunks
6	Seta arcuate when moist, straight when dry; plants growing on siliceous rocks or small calcareous stones
	Blindiadelphus (p.174)
-	Seta straight when moist and dry; plants growing on
7(2)	calcareous rocks
7 (2)	Leaves lingulate or spathulate; calyptra covering completely
	the capsule
_	Leaves not as above; calyptra not completely covering the capsule
8	Costa excurrent 9
0	Costa excurrent
_	costa percurrent(p.147)

9	Leaf margin with reflexed teeth at base
_	Leaf margin without reflexed teeth at base
10	Lamina bistratose or pluristratose in the upper part
_	Lamina unistratose in the upper part11
11	Capsule longly cylindrical or ellipsoidal
_	Capsule shortly cylindrical or ovoid
12	Leaves linear-lanceolate or lanceolate, strongly incurved at
	the apex
_	Leaves ovate-lanceolate, ovate, obovate or lingulate
13	Lid conical
-	Lid rostrate
	up 20 Acrocarps with exserted erect capsule, peristome
tee	th 16, divided halfway or to the base (32 teeth)
1	Peristome teeth slightly or strongly spirally twisted2
_	Peristome teeth straight or curved 10
2	Basal lamina cells hyaline, forming a well-delimited group
	3
-	Basal lamina cells hyaline or not, not forming a well-
	delimited group4
3	Basal hyaline cells of leaf ascending up margin in a V-
	shape
-	Basal hyaline cells of leaf forming an ovate group, not
	ascending up margin <i>Syntrichia</i> (p.156)
4	Peristome with well-developed basal membrane5
-	Peristome without or with low basal membrane6
5	Leaf margin conspicuously revolute; basal membrane of
	peristome to 45 μm
-	Leaf margin not revolute; basal membrane more than 300
	μm
	Tortula (inermis, mucronifolia, schimperi, subulata) (p.166)
6	Axillary hairs with 1–2(3) brown basal cells
	Didymodon (p.147)
	Axillary hairs hyaline throughout

7	Margin plane or recurved; costa with elongate cells on ventral side8
_	Margin recurved to strongly revolute; ventral cells of costa
	elongate or quadrate to shortly rectangular9
8	Costa excurrent in a thick point; leaf margin recurved from
	base to apex; perichaetial leaves not much different from
	vegetative leaves
_	Leaf apex at most with some hyaline cells forming a short
	apiculus, costa not excurrent; leaf margin plane or slightly
	recurved at leaf base; inner perichaetial leaves with
	sheathing base, conspicuously different from vegetative
	leaves
9	Ventral cells of costa long rectangular, smooth; margin
	conspicuously revolute; costa excurrent in hyaline awn
	Hilpertia velenovskyi (r)
-	Ventral cells of costa quadrate to shortly rectangular;
	margin recurved to strongly revolute; costa percurrent or
	excurrent in a short mucro
10 (1)	Lamina cells longer than wide11
-	Lamina cells ± quadrate
11	Peristome teeth divided to base
	Ditrichum s.l. (incl. Flexitrichum) (p.143)
-	Peristome teeth divided halfway
12	Leaves squarrose when moist
	Dichodontium pellucidum (w)
12(10)	Leaves not squarrose
13 (10)	Peristome teeth irregularly divided to base or nearly so14
- 14	Peristome teeth regularly divided to base or nearly so17 Lamina cells 15–20 µm wide
14	Lamina cells to 14 um wide
- 15	Costa excurrent in apiculus
-	Costa ending in or below apex16
16	Plants to 5 mm tall; lamina cells smooth
10	
_	Plants more than 5 mm tall; lamina cells mamillose on both
	surfaces(p.142)
17 (13)	Leaf margin entire or crenulate or slightly dentate near
(±0)	apex; lamina cells pluripapillose

-	Leaf margin strongly dentate in upper third; lamina cells mamillose
	p 21 Acrocarps with exserted capsule, inclined or ulous, peristome simple
1 - 2	Seta curved at least when moist 2 Seta straight 3 Plants minute; leaves not deciduous; growing on siliceous rocks or calcareous small stones Blindiadelphus (p.174) Plants medium-sized to robust; leaves deciduous, plants growing on humus-rich soil or dead wood Dicranodontium denudatum (rr)
3	Alar cells differentiated
_	Alar cells not differentiated
4	Median lamina cells rectagular, long and narrow, smooth
-	Median lamina cells quadrate, papillose or mamillose5
5	Lamina cells papillose, sometimes only faintly so; costa
-	excurrent <i>Tortula cernua</i> (<i>Desmatodon cernuus</i>) (rr) Lamina cells mamillose; costa percurrent <i>Dichodontium pellucidum</i> (w)
	p 22 Acrocarps with exserted capsule, inclined to ulous, peristome double
1	Plants bluish when dry (and dead) or with bluish to golden metallic sheen
2	Plants not bluish when dry and dead; lamina cells hexagonal,
2	20–24 μm wide
	Plants with metallic sheen; lamina cells linear, 8–12 µm
	wide
3	Costa 1/3 or more of leaf base4
-	Costa less than 1/3 of leaf base
4	Upper leaves linear, subulate <i>Leptobryum pyriforme</i> (w)
-	Upper leaves oblong-lanceolate, not linear
5	Leaves distinctly 3-ranked, patent to squarrose, carinate,
-	triangular-lanceolate

-	Leaves not as above6
6	Leaves strongly imbricate, plants julaceous
	Ptychostomum zieri (Plagiobryum zieri) (r)
-	Leaves not strongly imbricate, plants not julaceous7
7	Peristome teeth sigmoid; capsule asymmetrical
-	Peristome teeth straight; capsule symmetric, cylindrical to
	pyriform8
8	Leaves linear-lanceolate, more than 7 times as long as wide
	Orthodontium lineare (rr)
_	Leaves (except perichaetial leaves) less than 7 times as long
	as wide, lanceolate to ovate
9	Leaves narrowly ovate to lanceolate, unbordered or with a
	weak border of narrow cells, distinctly denticulate above;
	perichaetial leaves longer than vegetative leaves; costa
	rarely excurrent and then only shortly so, in section with
	large median guide cells; lamina cells linear to narrowly
	hexagonal, 5 or more times as long as wide; basal cells not
	differentiated Pohlia (p.198)
_	Leaves suborbicular to ovate, rarely lanceolate, usually with
	a distinct (sometimes thickened) border of narrow cells;
	margins entire or finely denticulate above, rarely distinctly
	so; perichaetial leaves hardly longer than vegetative leaves;
	costa often excurrent, sometimes as a long hair, in section
	with large ventral superficial cells; lamina cells rhomboidal
	to hexagonal, to 4 times as long as wide; basal cells quadrate
	or rectangular
Grou	up 23 Acrocarps with propagules on stems, leaves or in
rece	ptacles
1	Plants with axillary bulbils2
-	Plants without axillary bulbils
2	Leaves narrowly ovate to lanceolate, unbordered or with a
	weak border of narrow cells, distinctly denticulate above;
	perichaetial leaves longer than vegetative leaves; costa
	rarely excurrent and then only shortly so, in section with
	large median guide cells; lamina cells linear to narrowly
	hexagonal, 5 or more times as long as wide; basal cells not
	differentiated

-	Leaves suborbicular to ovate, rarely lanceolate, usually with a distinct (sometimes thickened) border of narrow cells; margins entire or finely denticulate above, rarely distinctly so; perichaetial leaves hardly longer than vegetative leaves; costa often excurrent, sometimes as a long hair, in section with large ventral superficial cells; lamina cells rhomboidal to hexagonal, to 4 times as long as wide; basal cells quadrate or rectangular
2	(incl. <i>Imbribryum</i> , <i>Ptychostomum</i>) (p.190)
3	Plants with foliose propagules in leaf axils
_	Plants with gemmae4
4	Gemmae crowded at ends of stems5
_	Gemmae in leaf axils or on leaves6
5	Gemmae globose, ovoidal or fusiform, in globose clusters on
	ends of leafless prolongations of stems
-	Gemmae lenticular or discoid, in the centre of a rosette of
_	apical leaves
6	Gemmae globose or ellipsoidal on axillary filaments
- 7	Gemmae globose or ellipsoidal, sessile, or filamentous9 Leaves with hair point
,	(dissimulata, lisae, muehlenbeckii, trichophylla) (p.175)
_	Leaves without hair point8
8	Gemmae clavate, few-celled, or ovoid, many-celled
•	
_	Gemmae spherical, sometimes irregularly so and with
	protruding cells Didymodon (cordatus, glaucus, rigidulus)
9	Gemmae globose, at tips of leaves, more than 80 μm
	Grimmia hartmanii (w)
-	Gemmae on costa, on leaf or in leaf axils, if at tips of leaves,
	then not globose or less than 70 μm $\!$
10	Gemmae irregularly globose, on ventral side of costa
	Syntrichia (latifolia, papillosa, virescens) (p.156)
-	Gemmae filamentous, fusiform, ovoid or ellipsoidal, on
11	lamina, at tips of leaves or in leaf axils
11	Gemmae filamentous, branched or not
-	or ovoid

12	Basal cells hyaline, forming a well-delimited group
-	Basal cells hyaline or not, not forming a well-delimited group
13	Leaves bordered, with 1–3 rows of elongate cells; lamina cells rectangular-hexagonal, smooth, more than 15 μm wide
-	Leaves unbordered; lamina cells rounded, papillose, to 14 µm wide
14 (11)	Calyptra cucullate, smooth, hairless; capsule much exserted on a thin seta; gemmae in leaf axils and in the tomentum of the stem
-	Calyptra mitrate, plicate, with or without hairs; capsule immersed to exserted on a strong seta; gemmae on the surface of leaves
15	Lamina cells papillose Zygodon rupestris (r)
-	Lamina cells smooth
16	Leaves concave, with erect incurved or involute margins; apex obtuse or rounded
-	Leaves ± keeled, rarely concave, with recurved, revolute or plane margins; apex obtuse, acute or acuminate, rarely rounded
17	Marginal cells at leaf base hyaline, rectangular, with thickened walls, ascending up margin, basal cells linear
-	Marginal basal cells elongate, similar to the rest of basal cells
Gro	up 24 Acrocarps with costa 1/3 or more width of leaf base
1	Costa excurrent in a hyaline hair point
_	Costa percurrent or excurrent in a coloured hair point2
2	Alar cells differentiated 3
-	Alar cells not differentiated 6
3	Costa consisting of green cells and hyaline cells (cross section)

-	Costa consisting of stereids and guide cells (cross section)
4	Lamina partly bistratose near costa <i>Dicranum fulvum</i> (r)
-	Lamina unistratose 5
5	Costa with dorsal stereids only or without stereids
	<i>Campylopus</i> (p.133)
-	Costa with dorsal and ventral stereids.
6 (2)	Costa with a central layer of green cells surrounded by hyaline cells (cross section); plants whitish or glaucous
-	Costa with a central layer of guide cells or not, not with
	outer layers of hyaline cells (cross section); plants green or brownish
7	Lamina cells papillose
-	Lamina cells smooth8
8	Leaves oblong-lanceolate; lamina cells longly hexagonal; costa without guide cells
-	Leaves linear to lanceolate; lamina cells rectangular to
0	linear; costa with guide cells
9	Leaves linear, flexuose, lamina cells linear, 45–55 μm long; rhizoidal tubers abundant
-	Lamina cells less than 45 μm long; rhizoidal tubers lacking or rare10
10	Costa wider than 180 µm at leaf base <i>Campylopus</i> (p.133)
-	Costa less than 180 µm wide at leaf base
11	Costa 80–110 µm wide at leaf baseFlexitrichum (p.133)
_	Costa to 30 µm wide at leaf base
	Dicranella (heteromalla, cerviculata) (p.135)
Gro	up 25 Acrocarps with lamina cells 18 μm wide or more
1	Leaves bordered by elongate cells, denticulate (sometimes
	very faintly so); plants often with creeping sterile shoots
-	Leaves not bordered; sterile shoots erect, not creeping2
2	Lamina cells quadrate, rounded or shortly polygonal3
-	Lamina cells longly polygonal or elliptical10
3	Costa ending below leaf apex4
_	Costa excurrent in apiculus or hair point

4	Leaf margin irregularly denticulate, plants bluish when dry (and dead)
_	Leaf margin entire; plants not bluish when dry
	Orthotrichum sprucei (rr)
5	Leaves apiculate, with reflexed, brownish yellow apiculus
	formed by a single elongate cell; margin irregularly
	denticulate above
	(Phascum leptophyllum, Leptophascum leptophyllum) (r)
_	Leaves not as above; margin entire
6	Capsule immersed Physcomitrium patens
	(Aphanorrhegma patens, Physcomitrella patens) (w)
-	Capsule exserted or emergent
7	Calyptra small, cucullate; spores less than 40 µm8
-	Calyptra large, inflated, cucullate, distinctly 4-angled; spores
	50–65 μm
8	Capsule dehiscent9
-	Capsule indehiscent
	(Pottia bryoides, Protobryum bryoides) (w)
9	Peristome single, teeth 16, irregularly divided; plants
	growing on soil
-	Peristome double, exostome teeth united in 8 pairs; plants
	growing in knot-holes
10(2)	
10 (2)	Plants bulbiform
- 11	Plants not bulbiform
11	Margin plane; costa excurrent in short apiculus
	Margin strongly revolute, with papillose cells (median
_	lamina cells smooth); costa excurrent in hyaline hair point
12	Plants rhizomatous; leaves crowded in terminal rosette
	Rhodobryum (p.190)
_	Plants not rhizomatous; leaves crowded in terminal rosette
	or evenly arranged along the stem
13	Leaf apex rounded 14
_	Leaf apex obtuse, acute or acuminate, but not rounded15
14	Costa percurrent; leaves 2–3 times as long as wide,
	decurrent, bordered by several rows of narrow, long cells
	Ptychostomum pseudotriquetrum fo. neodamense
	(Brvum neodamense) (n.s.)

_	long as wide, not decurrent, unbordered
	Splachnobryum obtusum (n.s.)
15	Capsule sulcate; peristome teeth fused at apices
	Funaria hygrometrica (w)
-	Capsule smooth
16	Capsule immersed, indehiscent (gymnostomous)
	Physicomitrium patens
	(Aphanorrhegma patens, Physcomitrella patens) (w)
- 17	Capsule exserted, dehiscent 17 Capsule asymmetrical and curved 18
_	Capsule symmetrical and straight
18	Capsule horizontal to pendulous; peristome teeth not
	sigmoid
_	Capsule inclined to horizontal; peristome teeth sigmoid
19	Capsule pendulous 20
-	Capsule erect or inclined21
20	Leaves narrowly ovate to lanceolate, unbordered or with a
	weak border of narrow cells, distinctly denticulate above;
	perichaetial leaves longer than vegetative leaves; costa
	rarely excurrent and then only shortly so, in section with large median guide cells; lamina cells linear to narrowly
	hexagonal, 5 or more times as long as wide; basal cells not
	differentiated
_	Leaves suborbicular to ovate, rarely lanceolate, usually with
	a distinct (sometimes thickened) border of narrow cells;
	margins entire or finely denticulate above, rarely distinctly
	so; perichaetial leaves hardly longer than vegetative leaves;
	costa often excurrent, sometimes as a long hair, in section
	with large ventral superficial cells; lamina cells rhomboidal
	to hexagonal, to 4 times as long as wide; basal cells quadrate
	or rectangular
21	Exothecial cells elongate
4 1	(hungaricus, muhlenbergii, pulchellus) (p.131)
_	Exothecial cells isodiametric 22
22	Calyptra mitriform; lid apiculate or rostellate; spores
	echinate

Calyptra cucullate or mitriform; lid convex or plane, without apiculus; spores not echinate..... Entosthodon fascicularis (r)

Group 26 Acrocarps with alar cells differentiated

1	Costa with stereids	Dicranum (p.139)
_	Costa without stereids	2
2	Leaves rigid, fragile, mostly broken	
	Di	cranum tauricum (w)
_	Leaves different; capsule symmetrical,	, smooth
		Blindia acuta (n.s.)

Group 27 Acrocarps with lamina cells isodiametric and leaf margins denticulate or dentate, at least near apex or base

1	teeth
_	Leaves not as above 2
2	Plants glaucous
_	Plants not glaucous
3	Leaves squarrose 4
_	Leaves not squarrose5
4	Basal cells hyaline, ascending up margin; leaf margin dentate above; frequent plants on open, dry calcareous soil
	Tortella squarrosa (Pleurochaete squarrosa) (w)
-	Basal cells not as above; leaf margin sharply serrate; very
	rare plants on peaty soils
5	Stem with dense brownish tomentum and usually with
	clusters of gemmae at the tip
-	Stem not as above6
6	Leaves linear-lanceolate, fragile, notched; margin dentate at
	the tip in young leaves
-	Leaves not as above
7	Leaves lanceolate to lingulate, with wide, acute or obtuse
	apex8
-	Leaves ovate or linear lanceolate, with gradually acuminate
	apex9
8	Lamina cells smooth
-	Lamina cells mamillose
9	Leaf margin plane or incurved 10

	_	Leaf margin recurved
	10	Marginal cells with slightly thicker walls than the rest of
		lamina cells, some longer, but not forming a distinct border; lid attached to columella after dehiscence; lamina cells not
		mamillose on ventral surface of leaf
		Hennediella heimii (Desmatodon heimii) (rr)
	_	Marginal cells similar to the rest of lamina cells; lid not
		attached to columella after dehiscence; lamina cells
		mamillose on ventral surface of leaf
	11	Leaves crisped when dry
	-	Leaves flexuose, slightly twisted or straight when dry16
	12	Leaves lanceolate, acuminate, with unistratose margins;
		lamina cells mamillose or finely papillose-striate13 Leaves linear-lanceolate, acute, often with bistratose
	_	margins; lamina cells smooth or mamillose14
	13	Plants pale green; median lamina cells mamillose; stem
		circular in cross section
	-	Plants dark green; median lamina cells finely papillose-
		striate; stem triangular in cross section
		Plagiopus oederianus (r)
	14	Lamina cells smooth
	- 15	Lamina cells mamillose
	15	Leaves tristicnous — Chestrum scristi (fi.s.) Leaves spirally arranged — Cynodontium (p.142)
16 (1	1)	Leaves with papillose-dentate margins, at least in upper half
)	
	-	Leaves with denticulate margins only at apex
	17	Leaf margin crenulate or papillose; apex denticulate, with
		few hyaline teeth; lamina cells papillose; capsule erect,
		smooth, not strumose
	_	Leaf margin not crenulate or papillose; apex slightly
	_	denticulate; lamina cells smooth; capsule inclined, striate,
		strumose
		p 28 Acrocarps with lamina cells isodiametric and
	excu	rrent costa
	1	Basal cells of leaf hyaline, ascending up margins in a V-shape
	1	
		· ur)

-	Basal cells of leaf hyaline or not, not ascending up margins
2	Basal cells of leaf hyaline, forming a well-delimited group on both sides of the costa, in arch-shaped area3
-	Basal cells of leaf hyaline or not, transition to cells above gradual4
3	Peristome teeth spirally twisted; calyptra partially covering
	the capsule
-	Peristome teeth not twisted or lacking; calyptra covering the whole capsule; basal hyaline lamina cells with pale orange cross walls distinctly thickened
4	Leaves partially bistratose in upper part
_	Leaves unistratose or with bistratose margins
5	Lamina cells smooth or mamillose
_	Lamina cells papillose
6	Costa excurrent in hyaline or yellowish hair point, rarely in apiculus; capsule indehiscent, or if dehiscent then leaf margin indistinctly bordered by longer cells
-	Costa ending below apex, percurrent or shortly excurrent; capsule dehiscent and leaf margin not bordered
7	Leaf margin revolute or recurved on one or both sides, at least partially
_	Leaf margin plane or incurved15
8	Leaf margins strongly revolute9
-	Leaf margins recurved on one or both sides10
9	Leaves lanceolate to ovate-lanceolate, triangular or lingulate ———————————————————————————————————
-	Leaves oblong, ovate, elliptical or obovate <i>Tortula</i> (p.166)
10	Capsule indehiscent11
-	Capsule dehiscent
11	Lid differentiated, persistent
	(Pottia bryoides, Protobryum bryoides) (w)
- 12	Lid not differentiated
12	Stereids numerous, in 3–5 (6) layers; capsule immersed or
	emergent
	(1. acaulon, Phascum cuspidatum) (p. 166)

_	Stereids few, in 1–2 layers or lacking; capsule emergent,
	rarely immersed
13 (10)	Peristome teeth erect, rudimentary or lacking <i>Tortula</i> pp.
	(p.166)
-	Peristome teeth filiform, spirally twisted
14	Cells of axillary hairs hyaline
-	Basal cells of axillary hairs brown
15 (7)	Capsule immersed to emergent
	<i>Weissia</i> (longifolia, rostellata) (p.173)
_	Capsule exserted 16
16	Leaves 3-4 (6) mm long; margins plane, sinuose, notched,
	papillose-crenulate; apex formed by a group of smooth,
	hyaline cells twice as long as papillose cells below
_	Leaves 0.4–3 (4.5) mm long; margin plane or incurved,
	entire17
17	Leaf margin plane or incurved, apex occasionally cucullate;
17	capsule longly cylindrical or ellipsoidal; peristome teeth
	straight or twisted, perforated
	•
-	Leaf margin incurved, at least in upper part, sometimes
	plane; capsule shortly cylindrical or ovoid; peristome teeth
	straight, entire or lacking
Cons	20 A come with its dismother leading calls loof on a
	up 29 Acrocarps with isodiametric lamina cells, leaf apex
obti	use or rounded, apiculate or not and costa not excurrent
1	Lamina cell walls sinuouse or incrassate with ± stellate
1	lumen
	Lamina cell walls neither sinuouse nor incrassate with
-	stellate lumen
2	
2	Lamina cell walls incrassate with ± stellate lumen
	Aulacomnium (p.211)
-	Lower lamina cell walls sinuoseGrimmiaceae (p.175)
3	Leaf margins plane or incurved
-	Leaf margins recurved 6
4	Lamina bistratose; capsule ovoid, asymmetric, immersed;
	perichaetial leaves ciliate
-	Lamina unistratose; capsules and perichaetial leaves not as
	above5

5	Basal lamina cells narrowly rectangular, (14) 16–40 (50)	
	(5) 6–7 μ m; costa 20–40 μ m wide at mid-leaf; leaves only 0.5	
	mm long, erect; capsule with annulus of large persisten	
	cells Gyroweisia tenuis (rr)	
-	Basal lamina cells short or long rectangular, 14–18 μm long	
	or if longer then costa 50–70 (90) µm wide at mid-leaf leaves various; plants sometimes taller; annulus of smal	
	cells, persistent or falling	
6 (3)	Costa homogeneous in cross section	
u(3)	(incl. <i>Lewinskya</i> , <i>Nyholmiella</i> , <i>Pulvigera</i>) (p.203)	
_	Costa heterogeneous in cross section	-
7	Leaves ovate-lanceolate; axillary hairs of 2–8 cells, 1–2 (3)	
,	basal cells brown	
_	Leaves obovate, lingulate or elliptical; axillary hairs of 4–5	
	cells, all hyaline	
	cens, an nyanne	
Gro	up 30 Acrocarps with isodiametric cells, apex acute	ρ
cun	acute or acuminate margins recurved at least on one side	
	acute or acuminate, margins recurved at least on one side	٠,
	acute or acuminate, margins recurved at least on one side ta not excurrent or lacking	.
	ta not excurrent or lacking Costa lacking)
1 -	ta not excurrent or lacking Costa lacking Hedwigia (p.188) Costa present 2)
cos	ta not excurrent or lacking Costa lacking Hedwigia (p.188) Costa present Upper lamina cells mamillose on both sides)
1 -	Costa lacking)
1 -	Costa lacking Hedwigia (p.188) Costa present 2 Upper lamina cells mamillose on both sides Cynodontium (p.142) Upper lamina cells smooth or papillose on one or both sides)
1 - 2 -	Costa lacking Hedwigia (p.188) Costa present Upper lamina cells mamillose on both sides Cynodontium (p.142) Upper lamina cells smooth or papillose on one or both sides)
1 -	Costa lacking Hedwigia (p.188) Costa present 2 Upper lamina cells mamillose on both sides Cynodontium (p.142) Upper lamina cells smooth or papillose on one or both sides Upper lamina cells smooth or papillose on one or both sides Upper lamina cells smooth 4)
1 - 2 - 3 -	Costa lacking)
1 - 2 -	Costa lacking) 2
1 - 2 - 3 -	Costa lacking) 2 .)
1 - 2 - 3 -	Costa lacking) 2 .) t
1 - 2 - 3 - 4 -	Costa lacking) 2 .)) t
1 - 2 - 3 -	Costa lacking) (2 ·) (3 ·) (4 ·) (7 ·)
1 - 2 - 3 - 4 -	Costa lacking) 2 ·) · · · ; ; ;) t) / .
1 - 2 - 3 - 4 -	Costa lacking Hedwigia (p.188) Costa present Zupper lamina cells mamillose on both sides Cynodontium (p.142) Upper lamina cells smooth or papillose on one or both sides Upper lamina cells smooth Upper lamina cells smooth Upper lamina cells papillose Supper lamina cells smooth Supper lamina cells)
1 - 2 - 3 - 4 - 5 -	Costa lacking) 2 ·) · · · · · ·) t // ·) j
1 - 2 - 3 - 4 -	Costa lacking)
1 - 2 - 3 - 4 - 5 -	Costa lacking) (2 ·) (3 ·) (4 ·) (5 ·) (7 ·)
1 - 2 - 3 - 4 - 5 -	Costa lacking)2 ·) · · · ; ; ;) t) / ;) 5 · · 7 3

-	Stomata phaneropore
8	Leaves ovate, elliptical or oblong; axillary hairs completely
	hyaline
_	Leaves linear-lanceolate to ovate-lanceolate; axillary hairs
	completely hyaline or with brown basal cells9
9	Axillary hairs with brown basal cells
_	Axillary hairs completely hyaline
10	Clavate or ovoid gemmae in leaf axils <i>Hydrogonium</i> (p. 171)
_	Axillary gemmae lacking
	Streblotrichum convolutum (Barbula convoluta) (p.169)
	Sir ebioti icham convolutum (barbaia convoluta) (p.107)
Cro	up 31 Acrocarps with isodiametric cells, apex acute,
	acute or acuminate, margins plane or recurved at base
only	y, costa not excurrent
1	Leaf margin entire2
_	Leaf margin crenulate or papillose-crenulate
2	Lamina cells smooth
_	Lamina cells papillose 4
3	Peristome double; lamina cells (7) 11–20 (23) µm wide
3	
_	Peristome simple; lamina cells 6–16 μm wide; plants to 5
_	mm tall
4	Plants rusty brown in lower part; lamina cells with c-shaped
4	papillae
- 5	Characters not as above
5	Lamina cells incrassate with ± stellate lumen; gemmae in
	groups at the end of pseudopodia
-	Lamina cells not incrassate with ± stellate lumen; without
_	gemmae in groups at the end of pseudopodia6
6	Lamina cells rounded or hexagonal, strongly papillose;
	gemmae fusiform, pluricellular
-	Lamina cells quadrate or rounded, papillose and finely
	longitudinally striate; gemmae lacking
	Amphidium mougeotii (r)
7 (1)	Leaves fragile, upper part often lost in older leaves; leaf
	margin irregularly erose, notched, ± undulate or sinuose,
	dentate-denticulate or distinctly papillose-crenulate8
-	Leaves not fragile; leaf margin neither sinuose, undulate nor
	notched, irregularly erose or dentate-denticulate9

8	Leaf apex formed by a group of smooth, hyaline cells, twice as long as papillose cells below
	(Oxystegus tenuirostris) (r)
_	Leaf apex not formed by smooth, hyaline cells; dentate in
_	young leaves or not
	(tophaceus subsp. erosus, D. sinuosus) (p.147)
9	Leaves carinate or keeled
9	
10	Leaves not carinate
10	Leaf margin bistratose above
-	Leaf margin unistratose
Cuor	un 22 Agragarma with lamina galla alangata lagyar
	up 32 Acrocarps with lamina cells elongate, leaves minate or subulate and apex consisting largely or entirely
of co	
or co	Sta
1	Lamina cells mamillose
_	Lamina cells smooth
2	Capsule indehiscent
_	Capsule dehiscent 4
3	Capsule exserted; neck distinct, about half the length of the
	urn
_	Capsule immersed; neck indistinct
4	Plants to 2.5 mm tall; saxicolous
_	Plants larger; terricolous or saxicolous
5	Seta arcuate when moist, straight when dry; plants growing
	on siliceous rocks or small calcareous stones
	Blindiadelphus (p.174)
_	Seta straight when moist and dry; plants growing on
	calcareous rocks
6	Plants without capsules, leaves with expanded sheathing
U	base, clasping the stem, and suddenly contracted into a
	narrow subula which is often squarrosely reflexed7
	Plants with capsules; leaves as above or otherwise8
7	Subula made up of prorate cells, therefore mamillose all
/	around, not just at margin
	(Ditrichum cylindricum) (w)
-	Subula cells not prorate, mamillae only at margin, not all
	around Dicranella schreberiana (w)

8	Peristome teeth divided to base with filiform segments				
	(Ditrichum cylindricum) (p.143)				
-	Peristome teeth divided to middle				
	up 33 Acrocarps with lamina cells elongate, leave apex use to acuminate, costa percurrent or excurrent, short or king				
1	Protonema persistent; plants minute, to 2.7 mm tall				
_	Protonema not persistent; plants small to robust				
2	Lamina cells mamillose; leaves dentate or serrate from base to apex (Bartramiaceae pp.)				
-	Lamina cells smooth or mamillose; leaves with entire margins or denticulate at apex only4				
3	Leaves narrowly or linear lanceolate, with sheathing base; margin with simple teeth				
-	Leaves lanceolate to ovate-lanceolate, without sheathing base; margin with simple or geminate teeth				
4					
4	Lamina cells quadrate to rectangular5 Lamina cells elongate hexagonal, rhomboidal or linear6				
- 5	Capsule dehiscent, exserted				
<i>-</i>	Capsule indehiscent, immersed				
6	Leaves appressed to erect when dry, imbricate				
_	Leaves erecto-patent to spreading, rarely imbricate				
7	Lamina cells hexagonal or rhomboidal-hexagonal; costa				
-	percurrent or excurrent in long or short point				
-	Lamina cells linear-hexagonal or linear-rhomboidal to vermicular; costa mostly not reaching leaf apex, rarely excurrent				
8	Leaves (except perichaetial leaves) less than 7 times as long as wide; capsule inclined or horizontal to pendulous				
-	Leaves more than 7 times as long as wide; capsule slightly inclined				

Group 34 Pleurocarps with long costa and lamina cells short, at least at margins

Leaf apex rour	ded		Leptodon	<i>smithii</i> (rr)	
Leaf apex obtu	se, acute or	acuminate		2	
Branch leaves	strongly de	ntate; plants :	± dendroio	d	
		<i>Tł</i>	namnobry	<i>rum</i> (p.229)	
Branch leaves irregularly bra				-	
Plants pinnate					
Plants irregula	,				
Stem 1-pinnat	2		Abietine	lla abietina	
•		(Thuidium	abietinum	(c) (p.219)	
Stem 2–3-pinn	ate		Thuid	i um (p.219)	
Leaf cells smoo					
Leaf cells papil					
Leaves with 2					
the costa; leaf margin entire or denticulate; plants slender,					
often with propaguliferous branchlets at the tips of stems					
and branches, growing on rock or tree bark; capsules rare					
		Pse			
Leaves not pl	icate; leaf	margin entir	e; plants	small, dark	
green, without propaguliferous branchlets, growing around					
vater-filled kn	ot-holes; ca	psules freque	ent, constr	ricted below	
nouth when e					
Note: For an acco	ant of this spec	cies in Hungary,	see Németh	and Erzberger	
(2015).	. 4/0.0	(O.). (
Costa extendin		-			
Costa nearly re					
Stem and bra					
reflexed, abruptly contracted from broad base; branch leaves					
slightly concave, ovate, obtuse or shortly acute					
Heterociadiei	la dimorph	a (Heteroclad	dium dimo	rphum) (rr)	
Note: For an acco Stem and bran					
stem leaves ei					
narrowed to a					
Note: For an acco					
(2019).	and or and spe	oloo iii iidiigai y	, see Baratii	and Braberger	
,					

9	Leaf without longitudinal plicae; alar cells not differentiated
	Pseudanomodon attenuatus) (p.230)
_	Leaf with 1–2 longitudinal plicae near base; alar cells
	differentiated
10	Leaf cells smooth or prorate
	Lescuraea saviana (Pseudoleskea saviana) (rr)
-	Leaf cells with a single central papilla each on the dorsal side
	Leskea polycarpa (cc)
	up 35 Pleurocarps with longitudinally plicate leaves, costa gand lamina cells elongate
-0	-
1	Plants dendroid
-	Plants irregularly or pinnately branched2
2	Branch leaves with reflexed teeth at apex
	Antitrichia curtipendula (r)
-	Branch leaves without reflexed teeth at apex3
3	Stem with paraphyllia4
-	Stem without paraphyllia6
4	Stems regularly pinnately branched; alar cells inflated or not
	5
-	Stems irregularly branched, alar cells not inflated, not
5	hyaline
3	Paraphyllia linear to linear-lanceolate; alar cells inflated, hyaline
_	Paraphyllia branched; alar cells not differentiated
_	
6	Stem leaves straight or only slightly falcate
-	Stem leaves straight of only slightly falcate
7	Plants very small and slender; leaves with two superficial
•	longitudinal plicae near base
_	Plants medium-sized to robust8
8	Stem with reddish or brownish tomentum of papillose
Ū	rhizoids
_	Stem without tomentum
9	Stem leaves triangular, gradually tapering into long fine
-	point
_	Stem leaves ovate to lanceolate, gradually or abruptly
	tapering into long or short point

10	Stem leaves ovate-lanceolate, gradually acuminate; costa of branch leaves ending in a dorsal projection or not; lid
	conical <i>Brachythecium</i> s.l. (incl. <i>Sciuro-hypnum</i>) (p.223)
_	Stem leaves ovate to lanceolate, sometimes cordate at base;
	costa usually ending in a projection at dorsal side of branch
	leaves; lid longly rostrate
11	Alar cells quadrate, incrassate, opaque, reaching costa
	Plasteurhynchium striatulum
	(Eurhynchium striatulum) (w)
_	Alar cells rectangular, thin-walled, not reaching costa
12 (6)	Stem with hyalodermis; alar cells inflated, hyaline
	Sanionia uncinata (r)
_	Stem without hyalodermis; alar cells not differentiated
	up 36 Pleurocarps with squarrose or falcate leaves, long a and elongated lamina cells
1	Leaves conspicuously transversely undulate
-	Leaves not transversely undulate2
2	Leaf apex acute or obtuse
-	Leaf apex acuminate3
3	Stem with paraphyllia4
-	Stem without paraphyllia5
4	Alar cells well developed, inflated, hyaline
_	Alar cells neither inflated nor hyaline
5	Leaf acumen channelled6
_	Leaf acumen not channelled 7
6	Plants growing in dry calcareous habitats; stem leaves to 1.7
Ü	mm long; alar cells slightly inflated, the widest cells 10.5–
	17.5 (21) µm wide
	(Campylium chrysophyllum) (W)
_	Plants growing in wetlands; stem leaves at least 1.3–1.6 mm
	long; alar cells strongly inflated, the widest cells 17–31.5 μm
	wide
	Drepanocladus polygamus (Campylium polygamum) (r)

7	Leaf margin distinctly denticulate, at least near apex
	Sarmentypnum exannulatus (Warnstorfia exannulata) (rr)
-	Leaf margin entire or obscurely denticulate8
8	Group of alar cells large, of 2–4 rows of cells, ascending up
	margin and sometimes reaching costa
-	Group of alar cells small, of 2–10 cells, not ascending up
	margin and not reaching costa
	up 37 Pleurocarps with long costa, elongated lamina cells rounded, obtuse, or obtuse and apiculate apex
anu	Tounded, obtuse, or obtuse and apiculate apex
1	Costa more than 40 µm wide
	<i>Hygroamblystegium fluviatile</i> (Amblystegium fluviatile) (r)
-	Costa less than 40 µm wide2
2	Group of alar cells well differentiated3
_	Group of alar cells not or hardly differentiated
3	Alar cells small, opaque
_	Alar cells large, inflated and hyaline
4	Leaves ovate-cordate; costa reaching apex or nearly so;
	group of alar cells triangular, nearly reaching costa, not
	ascending up margins; axillary hairs frequent, long, with 2–5
	apical hyaline cells
-	Leaves oblong or ovate; costa reaching 3/4 up leaf; group of
	alar cells ovate, extending up margin; axillary hairs scarce
	and short, with 2–3 apical hyaline cells; leaf apex often with
	rhizoids
	Straminergon stramineum (Calliergon stramineum) (rr)
5	Stems pinnately branched; lid conical
	Pseudoscleropodium purum (Scleropodium purum) (c)
-	Stems irregularly branched; lid longly rostrate
	Rynchostegium (n.220)

$\it Group~38$ Pleurocarps with long costa, elongated lamina cells and acute or acuminate apex

1	Branch leaves with reflexed teeth at apex
	Antitrichia curtipendula (r)
-	Branch leaves without reflexed teeth at apex2
2	Leaf margins dentate to ciliate or laciniate. <i>Fabronia</i> (p.214)
-	Leaf margins entire to denticulate3
3	Lamina cells 2–6 (7) times as long as wide4
-	Lamina cells more than 7 times as long as wide9
4	Alar cells hyaline, inflated, forming a distinct group
-	Alar cells forming a poorly differentiated group5
5	Margins at leaf base with ± recurved teeth; sometimes with
	rhizoids on dorsal side of costa; plants small
-	Margins at leaf base without recurved teeth; never with
	rhizoids on dorsal side of costa; plants small to medium-
	sized6
6	Costa ending in conspicuous projection at the dorsal side of
	branch leaves; plants small
	(Eurhynchium pumilum) (r)
_	Costa not ending in projection at the dorsal side of branch
_	leaves
7	Costa 25–75 (100) µm wide, reaching mid-leaf or above
	Hygroamblystegium (p.216)
-	Costa 15–35 (40) µm wide, reaching above mid-leaf or not
_	8
8	Costa reaching mid-leaf or shorter (in A. serpens often
	longer), sometimes double or lacking, capsules not
	constricted below mouth when empty; habitat various (incl.
	knot-holes)
_	Costa reaching to leaf apex; plants small, dark green,
	growing around water-filled knot-holes; capsules frequent,
	constricted below mouth when empty
	Note: For an account of this species in Hungary, see Németh and
	Erzberger (2015).
9 (3)	Alar cells hyaline, inflated
_	Alar cells neither hyaline nor inflated

Leaves acuminate	
Alar cells small, opaque	, incrassate; branches usually curved
Alar cells different; bran	nches ± straight12
Leaf acumen channelled	d
	elled or leaves acute13
Stem and branch leaves	s differentiated (heteromorphous) 14
Stem and brach leaves s	similar (homomorphous)15
Stem regularly pinnatel	y branched; seta rough
Kindbergia prae	longa (Eurhynchium praelongum) (r)
	ned; seta smooth
	Eurhynchiastrum pulchellum
	(Eurhynchium pulchellum) (w)
Costa ending in conspic	cuous projection at the dorsal side of
	16
Note: In <i>Oxyrrhynchium sp</i> developed.	eciosum this character is only occasionally
Costa not ending in pr	ojection at the dorsal side of branch
	onspicuous17
	e-lanceolate, shortly acuminate; lid
	vate-lanceolate, longly acuminate; lid
	Brachythecium s.l.
	heciastrum, Sciuro-hypnum) (p.223)
	racted or gradually narrowed into
large, filiform acumen;	axillary hairs with 3 short basal and 2
	ontracted into large acumen; axillary
	ls18
	near-lanceolate or narrowly oblong-
	or large; leaves lanceolate, ovate,
	19
Shoots subcomplanate;	plants aquatic
	Leptodictyum riparium (c)
Shoots not subcomplan	ate; plants aquatic or not20
Lid longly rostrate	
Brachytheci	<i>um</i> s.l. (incl. <i>Sciuro-hypnum</i>) (p.223)

Group 39 Pleurocarps with short or lacking costa, short lamina cells at least at margin

1	Leaves longitudinally plicate
-	Leaves with two short longitudinal plicae at base or not longitudinally plicate2
2	Lamina cells papillose; small to medium-sized plants3
_	Lamina cells smooth; slender plants
	Pseudoleskeella (p.219)
3	Plants ± dendroid; upper branches curved when dry
-	Plants not dendroid; branches not curved when dry4
4	Stem leaves very concave, imbricate; plants julaceous
-	Stem leaves plane or slightly concave, patent to squarrose,
	not imbricate; plants not julaceous5
5	Stem and branch leaves strongly dimorphic; stem leaves
	reflexed, abruptly contracted from broad base; branch
	leaves slightly concave, ovate, obtuse or shortly acute
	Heterocladiella dimorpha (Heterocladium dimorphum) (rr)
	Note: For an account of this species in Hungary, see Baráth <i>et al.</i> (2016).
-	Stem and branch leaves similar in shape, but differing in
	size; stem leaves erecto-patent to patent, not reflexed,
	gradually narrowed to acute apex
	Meterocladium heteropterum (rr) Note: For an account of this species in Hungary, see Baráth and Erzberger
	(2019).
Grou	up 40 Pleurocarps with short or lacking costa, elongated
lami	na cells and rounded, obtuse or apiculate apex
1	Lamina cells prorate on dorsal side; plants small
	Pterigynandrum filiforme (w)
-	Lamina cells smooth; plants ± robust
2	Stem and branch tips conspicuously cuspidate
	Calliergonella cuspidata (w)
-	Stem and branch tips not conspicuously cuspidate3
3	Plants pinnately branched4
_	Plants irregularly branched

4	Stems reddish; alar cells orange to brown
-	Stems greenish, yellowish or light brown; alar cells hyaline or greenish
5	Leaves longly decurrent; plants not aquatic
-	Leaves not or scarcely decurrent; aquatic plants6
6	Plants robust, turgid; alar cells hyaline, large, forming a well-delimited small group; plants growing on water-logged soil
-	Plants small to medium-sized, not turgid; alar cells not hyaline, not in well-delimited group; plants on rock or stones in streams
leav	up 41 Pleurocarps with distinctly falcate or squarrose ves, short or lacking costa, elongated lamina cells and acute cuminate apex
1	Leaves squarrose; acumen ± channelled2
_	Leaves not squarrose; acumen flat4
2 -	Stems reddish
3	Plants medium-sized to robust; margins entire to finely denticulate; alar cells inflated, well-differentiated
-	Plants small; margins dentate; alar cells poorly differentiated
	Campylophyllopsis calcarea (Campylium calcareum) (w)
4 (1)	Leaf apex acute; aquatic plants
_	Leaf apex acuminate; plants not aquatic
5	Plants robust, alar cells hyaline, large, forming a well-delimited small group; plants growing on water-logged soil
-	Plants small to medium-sized, not turgid; alar cells not hyaline, not in well-delimited group; plants on rock or stones in streams
6	Leaves wide and shortly acuminate; stems with hyalodermis
-	Leaves narrowly and longly acuminate; stems without hyalodermis

7	Leaves strongly longitudinally plicate; plants pinnately branched
-	Leaves not or weakly longitudinally plicate; plants pinnately
•	branched or not
8	Leaves cordate at base; branch leaves differing from stem
	leaves; plants pinnately branched
	Ctenidium molluscum (w)
-	Leaves not cordate at base, branch leaves similar to stem
	leaves; plants irregularly or pinnately branched
squ	up 42 Pleurocarps with straight, slightly falcate or arrose leaves, costa single, short, long and double, or sing, elongated lamina cells and acute or acuminate apex
1	Lamina cells distinctly papillose or prorate on dorsal side
-	Lamina cells smooth or only slightly prorate on dorsal side4
2	Plants slender, filiform
_	Plants robust, not filiform
3	Stems regularly bi-tripinnate, costa double, reaching 1/2 of
	leaf length
_	Stems irregularly branched; costa double, reaching 3/4 of
	leaf length
	(Rhytidiadelphus triquetrus) (w)
4 (1)	Plants aquatic; leaves carinate or not
_	Plants not aquatic; leaves not carinate
5	Lamina cells 2–5 times as long as wide; slender plants6
_	Lamina cells more than 5 times as long as wide; plants small
	to robust9
6	Leaves with 2 longitudinal plicae at base, on each side of the
	costa; lamina cells shorter at margin
	Pseudoleskeella (p.219)
_	Leaves not plicate; lamina cells ± homogeneous
7	Leaf margin entire
_	Leaf margin denticulate, dentate or ciliate8
8	Leaf margin denticulate <i>Platydictya jungermannioides</i> (rr)
_	Leaf margin distinctly dentate or ciliate <i>Fabronia</i> (p.214)
9 (5)	Leaves squarrose or spreading to reflexed

-	Leaves erect to patent11
10	Plants small; leaves to 1 mm long, spreading to reflexed
	Campylophyllopsis calcarea (Campylium calcareum) (w)
-	Plants robust, leaves 2.7–3.7 mm long, squarrose
	Rhytidiadelphus squarrosus (w)
11	Stem with branched paraphyllia
	Loeskeobryum brevirostre (Hylocomium brevirostre) (n.s.)
-	Stem without paraphyllia, but sometimes with linear
	pseudoparaphyllia12
12	Alar cells poorly or not differentiated13
-	Alar cells ± well developed, at least in small group of (2) 3-6
	cells
13	Plants robust, julaceous, glossy; leaves ovate, shortly
	pointed, concave; leaf margin distinctly denticulate,
	particularly near apex
-	Plants small to robust, not julaceous, glossy; leaves linear
	lanceolate, longly acuminate, not or slightly concave; leaf
	margin entire or slightly denticulate, sometimes near base
	only14
14	Basal cells usually porose, often brownish; axillary hairs
	with hyaline basal cell
-	Basal cells not or slightly porose, concolorous with the rest
	of cells; axillary hairs with brown basal cell
	Isopterygiopsis pulchella (r)
15	Alar cells inflated 16
-	Alar cells not inflated 18
16	Stem with hyalodermis
-	Stem without hyalodermis
7	Plants medium-sized to robust; capsule erect to slightly
	inclined, curved below mouth; lid conical to shortly rostrate,
	0.8–1 mm long; linear pseudoparaphyllia in leaf axils
	Plants small; capsule ± erect; lid including beak 0.5–0.6 mm
	long, very longly rostrate, beak 0.28-0.36 mm long, longer
	than conical basal part of lid; without pseudoparaphyllia
	Sematophyllum adnatum (rr)
8	Alar cells small, opaque; branches usually curved
-	Alar cells different; branches straight

19	Group of alar cells concave, of incrassate cells
	Hypnum s.l. (incl. Buckia vaucheri) (p.227)
-	Group of alar cells not concave, of thin-walled or incrassate
	cells
20	Leaf margin denticulate from base to apex
-	Leaf margin entire or denticulate only near apex21
21	Central strand distinct; external surface of exostome teeth
	smooth at base; corticole, rarely saxicole
	Pylaisia polyantha (cc)
_	Central strand indistinct; external surface of exostome teeth
	papillose or striate-papillose at base; corticole or saxicole
	22
22	Leaf margin narrowly recurved in lower 2/3; plants usually
	with propaguliferous branchlets at the tip of branches;
	capsule straight; corticole
_	Leaf margin plane; plants without propaguliferous
	branchlets; capsule curved; saxicole
	()

PART II: SPECIAL KEYS

5. KEYS TO SPECIES OF LIVERWORTS

Key to species of *Barbilophozia* group, incl. *Neoorthocaulis* floerkei

See also key to Lophoziaceae group below

	of the state of th
1	Lateral leaves mostly 3–4 (5)-lobed (occasionally 2-lobed leaves may occur)
_	Lateral leaves mostly 2-lobed, apex lobed only to 1/3;
	underleaves present, small, entire, often difficult to detect;
	lateral leaves rounded, sinus semi-lunate; gemmae reddish-
	brown, walls ± solid; leaves sometimes with erosed margins
	caused by the production of gemmae; on non-calcareous rocks and soil
	Note: This species can be recognised by clusters of red-brown ripe gemmae in combination with shallowly notched, concave leaves and small leaf cells $24-25 \times 18-20 \mu m$ with (4) 6–9 (15) oil bodies per cell. <i>Lophoziopsis excisa</i> , which also produces red gemmae, differs by larger leaf cells, $30-35$ (40) × (27) $28-30$ (32) μm and more numerous oil bodies, $11-24$ (28) per cell. <i>Lophoziopsis longidens</i> , another species with clusters of red gemmae at the tip of the leaf lobes, has also large cells, $28-35 \times 23-27$ (30) μm , but fewer oil bodies, $4-10$ (14) per cell.
2	Postical margins of lateral leaves with basal cilia; underleaves present
-	Postical margins of lateral leaves without cilia; underleaves absent; leaves (2) 4 (5)-lobed, lobes unequal; trigones small; shoots 3–8 cm long and up to 5 mm wide, procumbent
3	Cells of cilia subquadrate, 15–30 (38) µm long; lateral leaves
	lobed to 1/3 with (2) 3 (4) obtuse or acute lobes; shoots
	mostly ascending to erect; rarely with gemmae
-	Cells of cilia on underleaves and base of lateral leaves
	elongate, 20–80 µm long or more; gemmae usually present
	Barbilophozia hatcheri (rr)

Key to species of CephaloziaceaeBased mainly on Damsholt (2002)

1	Lateral leaves asymmetrical, with 2 long acuminate lobes (4-7 uniseriate cells); postical margin strongly inflexed and forming inflated water sac; on rotting wood
-	Lateral leaves not or only slightly asymmetrical; postical margin not inflexed, not forming sac
2	Leaves longer than wide, divided 0.5–0.7 of length, not decurrent, almost transversely inserted or not
-	Leaves as wide as long, divided 0.25–0.5 of length, often decurrent, obliquely to horizontally inserted; stem on mature leafy shoots with antical leaf-free zone 2 cells wide
3	Leaves obliquely to horizontally inserted, 4–7 cells wide; cells at base of lobes $40-50 \times 20-45 \mu m$; plants whitish, small, shoots $0.4-0.6 \ mm$ wide; perianth mouth laciniate; dioicous
_	Leaves almost transversely inserted, non-decurrent, 8–16 cells wide; cells at base of lobes $30-70 \times 25-50$ µm; plants often with secondary pigmentation, large to medium-sized, shoots $0.6-1.2$ mm wide; perianth mouth denticulate, with teeth formed by $1-3$ superimposed cells; autoicous; stem with antical end of leaf insertion extending to median cortical cells, sometimes almost to the antical mid-line (except in fertile or gemmiferous shoots); flagelliform shoots often present
4	Cells at base of lobes 18–36 \times 16–35 μm , thin- or thick-walled; stolons lacking
-	Cells at base of lobes (35) 40 –55 (70) × (28) 33–48 µm, thin-walled; plants often with stolons; leaves usually 12–25 cells wide; apical cell on leaf lobes with wall not thickened at apex; autoicous; perianth mouth crenulated
5	Leaves decurrent, lobes strongly connivent; cells at base of lobes 26–32 (40) × 20–28 (32) μ m; plants green, stem concolorous; leaves mostly (7) 9–14 (16) cells wide; apical cell on leaf lobes with wall thicker at apex than on margins; dioicous; female bracts entire; perianth mouth shortly lobed,

	the lobules crenulate-dentate with teeth 1–2 cells long
	Fuscocephaloziopsis lunulifolia
	(Cephalozia lunulifolia) (n.s.)
-	Leaves shortly decurrent, not or barely connivent; cells at
	base of lobes 20–35 × 15–30 μm; plants green to olive-green
	or yellowish brown, stem often also somewhat yellowish
	brown; leaves (12) 13–16 cells wide or wider6
6	Cells at base of lobes $20-25 \times 15-18 \mu m$, thick-walled; apical
	cell on leaf lobes with wall not thickened at apex; dioicous;
	perianth mouth laciniateFuscocephaloziopsis catenulata
	(Cephalozia catenulata) (n.s.)
_	Cells at base of lobes $28-35 \times 20-30 \mu m$, thin-walled; apical
	cell on leaf lobes with wall thicker at apex than on margins;
	dioicous; female bracts dentate; perianth mouth dentate, not
	laciniate
	(Cephalozia macrostachya) (n.s.)
Kev	to species of Cephaloziella
	ortant literature: Meinunger and Schröder (2007): Vol. 1: 176–200
1	Gemmae 2-celled, cubic, angulate or with warty elevations
	Cephaloziella integerrima (n.s.)
-	Gemmae 2-celled, elliptical, smooth or gemmae lacking2
2	Dioicous, often sterile; underleaves distinct; leaf cells small,
	8–12 μm Cephaloziella divaricata (w)
	Note: <i>C. divaricata</i> var. <i>scabra</i> (M. Howe) Haynes (rr) is doubtfully distinct from the typical variety and can be recognized by denticulate leaf
	margins, a verruculose cuticula and ± strong papilla-like outgrowths
	below the sinus on the abaxial surface of upper leaves.
_	Monoicous, mostly fertile3
3	Paroicous 4
_	Autoicous
4	Sterile shoots with distinct, often large and 2-lobed
	underleaves. <i>Cephaloziella stellulifera</i> (n.s.)5
_	Sterile shoots without underleaves, but occasionally with
	rudimentary underleaves consisting of 2 or 3 cell rows
	towards the shoot apex; cells 10–16 µm wide, in lower part
	of leaf lobes cells of regular rectangular shape; leaf lobes
	only 4–5 cells wide at base. <i>Cephaloziella rubella</i> agg. (w)
	pp6
	1.1

5	Cells large, 14–20 µm wide; leaves entire, patent to squarrose; female bracts partly in stellate arrangement
-	Cells smaller, narrower, 10–15 µm wide; leaf lobes 4–9 cells wide at base; underleaves small and indistinct except in gametangiophores; plants light to yellowish green, in insolated situations also brownish, only perianths and male bracts occasionally with a light reddish hue; similar to <i>C. varians</i> , but paroicous
6	Female bracts often connate forming a ring around perianth
	base, urn-like and extending far upward
	Cephaloziella rubella var. rubella
-	Female bracts more deeply divided nearly to base, not forming an urn around the base of the perianth (Boros 1968: on rotting wood, Bükk, Vértes)
7	Sterile shoots without underleaves
7	Sterile shoots without underleaves, at least towards
	shoot apex; cells medium-sized to small, 7–14 µm wide13
8	Cells large, 14–20 µm wide9
-	Cells medium-sized, 10–15 mm wide
9	Female bracts with rounded, entire, often somewhat undulate lobes; perianth pyriform, free
	Cephaloziella integerrima (n.s.)
-	Female bracts with acute, ± dentate lobes10
10	Plants light to pale green, in insolated places ± brownish; cell
	walls always thin to moderately thickened; leaf lobes 6-10
	cells wide at base
-	Plants at least partly red, especially the male bracts often
	deeply purplish to blackish red; cells thin- or thick-walled;
	female bracts often coarsely to squarrosely dentate; cells of
	perianth mouth conspicuously elongate; leaf lobes 4–6 cells wide at base
11	Plants in calcareous habitats; [perhaps to be expected in
11	Hungary, but not yet recorded]
	Cephaloziella baumgartneri
_	Calcifuge species 12
12	Plants pale green, in open places partly brownish; cells
	always thin-walled: leaf lobes 6–10 cells wide at base: in well

	developed plants cells of perianth mouths free for half their length, perianth mouth therefore denticulate
	Cephaloziella hampeana (rr)
-	Plants ± reddish, especially male bracts mostly strongly red brown; cell walls partly distinctly to strongly thickened; leaf
	lobes 4–6 cells wide at base; even in well developed plants
	cells of perianth mouth fused together for nearly all their
	length, perianth mouth therefore crenulated
	Cephaloziella rubella var. bifida
13 (7)	Leaf lobes mostly 2–5 cells wide at base, mostly long and
	narrow, occasionally with a few teeth; cells mostly distinctly,
	sometimes strongly thickened, often papillose; older and
	insolated parts of plants often conspicuously copper-
	coloured; calcifuge, in bogs, on peat and humid, lime-free
	sand (Boros 1968: on rotting wood, Zemplén, Bükk)
_	Leaf lobes 6–12 cells wide at base, shorter and wider than in
_	the preceding species, mostly entire; cells mostly thin-walled
	to moderately thickened; plants green, yellowish, brown or
	red, not conspicuously copper-coloured; in base-rich, ±
	calcareous habitats. <i>Cephaloziella varians</i> agg14
14	Stem at apex partly with a few hyaline teeth; leaves partly
	denticulate; cells at leaf base with conical mamilla on the
	dorsal side
-	Stem and leaves smooth, without such outgrowths15
15	Leaf lobes ovate, rounded at apex; male inflorescences
	julaceous, club-shaped
-	Leaf lobes pointed, often with two uniseriate cells at apex;
	male inflorescences mostly with homomallous leaves
	Cephaloziena varians var. varians (11)
Kev	to Lophoziaceae group (incl. Barbilophozia, Isopaches,
	ozia, Lophoziopsis, Mesoptychia, Neoorthocaulis, Obtusifolium,
	stochilopsis, Trilophozia and Tritomaria)
Impo	rtant references: Meinunger and Schröder (2007), Bakalin (2016)
1	Lateral leaves mostly 2.4 (E) labed (accessional 2 labed
1	Lateral leaves mostly 3–4 (5)-lobed (occasional 2-lobed leaves may occur in some species)
	icaves may occur in some species j2

-	Lateral leaves mostly 2-lobed, apex rarely emarginate or only shallowly 2-lobed (occasional 3–4 (5)-lobed leaves may
	occur in some species)
2	Lateral leaves obliquely inserted, mostly symmetrical or only weakly asymmetrical, (2) 3-4 (5)-lobed with sinus
	(1/8) $1/4-1/3$ leaf length; postical leaf margins with or
	without basal cilia; underleaves distinctly and deeply
	divided into 2 lobes or absent or minute except on fertile
	shoots; gemmae rare, mostly absent. <i>Barbilophozia</i> 3
_	Lateral leaves ± transversely inserted, distinctly
	asymmetrical, (2) 3-lobed with sinus to 1/4 leaf length;
	postical leaf margins without basal cilia; underleaves mostly
	absent or undivided; postical surface of stem sometimes
	brown or red (in <i>Trilophozia quinquedentata</i>)5
3	Postical margins of lateral leaves with basal cilia;
	underleaves present
_	Postical margins of lateral leaves without cilia; underleaves
	absent; leaves (2) 4 (5)-lobed, lobes unequal; trigones small;
	shoots 3–8 cm long and up to 5 mm wide, procumbent
	Barbilophozia barbata (w)
4	Cells of cilia subquadrate, 15–30 (38) µm long; lateral leaves
	lobed to 1/3 with (2) 3 (4) obtuse or acute lobes; shoots
	mostly ascending to erect; rarely with gemmae
	Neoorthocaulis floerkei (Barbilophozia floerkei)
	Note: excluded – the voucher was collected in Austria.
-	Cells of cilia on underleaves and base of lateral leaves
	elongate, 20–80 μm long or more; gemmae usually present
_	Barbilophozia hatcheri (rr)
5	Leaf lobes mostly narrowly triangular; leaf cells with equally
	thickened walls; reddish gemmae nearly always present,
	often abundant and conspicuous
-	Leaf lobes mostly broadly ovate-triangular; leaf cells with
	coarse trigones; lateral leaves as long as wide, 3-lobed in
	unequal lobes; lobes acute to acuminate or apiculate;
	gemmae rare, often sparse and inconspicuous
	Trilophozia quinquedentata (Tritomaria quinquedentata) (r)
6	Mid-leaf cells averaging 19–22 µm wide; gemmae obtusely
U	angled or pyriform, 14–22 µm wide, 16–28 µm long
	Tritomaria exsectiformis (n.s.)
	1 i ituliui iu easettijui iiiis (11.5.)

-	Mid-leaf cells averaging 10–14 μm wide; gemmae ellipsoid,
	smooth, 8–12 μm wide, 12–19 (22) μm long
	Tritomaria exsecta (r)
7 (1)	Perianths often present, plicate in the upper 1/3; gemmae
	often present, angulate in Hungarian species; underleaves
	mostly absent; plants generally on non-calcareous
	substrates (but Obtusifolium obtusum and Lophoziopsis
	excisa tolerating lime or base-rich substrates)8
_	Perianths absent, or without plicae; gemmae absent or if
	present then smooth (in Mesoptychia heterocolpos with
	attenuate shoots); underleaves present or if absent then
	lateral leaves with blunt to rounded lobes (Gymnocolea
	inflata: calcifuge; Mesoptychia badensis: calciphile)19
8	Cells of lateral leaves mostly equally thick-walled; capsule
	wall 2-layered; paroicous; lateral leaves closely imbricate, 2-
	lobed to 1/5-1/3; plants smelling of cedar oil when crushed;
	frequently with abundant gemmae, reddish yellow to
	reddish brown; on sandy or peaty acid soil, occasionally on
	rocks
	Note: This species often produces abundant perianths with a long-dentate
	mouth, cilia to 3–4 cells long, whereas the perianth mouth of <i>Lophoziopsis excisa</i> , with which it might be confused, is crenulate, at most with short
	finger-shaped marginal cells.
_	Cells of lateral leaves mostly thin-walled; trigones present or
	not; capsule-wall 3–4-layered; plants not aromatic9
9	Lobes of lateral leaves mostly obtuse, sinus gibbous; young
	stems with lanceolate or 2-lobed underleaves; gemmae rare,
	mostly 2-celled, polygonous; lime-tolerant
_	Plants lacking this combination of characters; leaf lobes
	acute, rarely obtuse (in <i>Lophozia wenzelii</i>)10
10	Lateral leaves 2–3 (5)-lobed to 1/3–1/2, lobes dentate to
	spinose dentate; upper lateral leaves somewhat plicate and
	crispate; underleaves absent on sterile shoots; gemmae
	usually abundant, 1–2-celled; bright to pale green plants on
	rotting wood and acid soil, peat, in rock crevices and among
	other bryophytes
-	Plants lacking this combination of characters11

11	lateral lateral lateral label and the 1/2 since and label
	lateral leaves rounded, lobed only to 1/3; sinus semi-lunate;
	gemmae reddish-brown, walls ± solid; leaves sometimes
	with erosed margins caused by the production of gemmae;
	on non-calcareous rocks and soil
	Note: This species can be recognised by clusters of red-brown ripe gemmae in combination with shallowly notched, concave leaves and small leaf cells $24-25 \times 18-20 \mu m$ with (4) $6-9$ (15) oil bodies per cell. Lophoziopsis excisa, which also produces red gemmae, differs by larger, thin-walled leaf cells, $30-35$ (40) × (27) $28-30$ (32) μm and more numerous oil bodies, $11-24$ (28) per cell. Lophoziopsis longidens, another species with clusters of red gemmae at the tip of the leaf lobes, has also large cells, $28-35 \times 23-27$ (30) μm , but fewer oil bodies, $4-10$ (14) per cell.
12	Underleaves absent on sterile stems; lateral leaves 2-lobed, occasionally 3-lobed; mid-leaf cells 20–30 (40) µm; oilbodies 4–8 (9) µm, smooth or granular, 8–24 per cell; lobes of female bracts mostly entire (except in <i>Lophoziopsis excisa</i>); stems 10–24 cells high in cross section; cells of medulla clearly differentiated, with antical band of wide hyaline cells and postical band of small cells infected with mycorrhiza; epidermal (cortical) cells often larger than cells of medulla
12	decumbent and ascending; apical leaves constantly with pale green, angulate gemmae 18–20 µm; lateral leaves ovate to oblong, with narrow, often horn-like lobes; trigones triangular, colourless; on rotting wood
-	Plants larger; mostly on soil, humus, peat, rocks, occasionally on wood
13	Apical leaves often imbricate; shoot apex resembling
13	miniature lettuce; lateral leaves asymmetrical, lobed to 1/4; leaf cells thin-walled, trigones small or absent; paroicous; perianths common, mouth dentate, but cilia absent; gemmae frequent, reddish, angulate, on young leaves
	above.
-	Plants lacking this combination of characters

14	Lower lateral leaves often somewhat reflexed, upper with narrow horn-like lobes bearing globose clusters of reddish-
	brown gemmae; trigones small
	Note: See note under <i>Barbilophozia sudetica</i> above.
-	Plants lacking this combination of characters
15	Cross section of stem in postical half with 5-20 layers of
	small cells reaching or extending beyond the middle16
-	Cross section of stem in postical half with 0-4 layers of small
	cells reaching at most to lower third
16	Group of small cells infected by mycorrhiza yellowish brown
	to brown, mostly only the postical cortex cells purplish-red;
	sometimes also the lower cell layers purplish-red, but
	always surrounded by a "halo" of brownish cells at the
	transition towards the larger hyaline medulla cells above;
	leaves rectangular to oval, widest below mid-leaf; gemmae
	mostly frequent. <i>Lophozia ventricosa</i> s.l. (r)
_	Group of small cells infected by mycorrhiza always dark
	purplish-red, without a "halo" of brownish cells; leaves often
	widest above mid-leaf, distinctly narrowed towards the
	base, sometimes nearly cuneate (wedge-shaped); gemmae
	sparse to lacking
	Note: excluded, specimens revised to <i>L. guttulata</i> .
17	Oil bodies homogeneous, uniformly granular
	Lophozia ventricosa s.str. (r)
	Oil bodies biconcentric, finely granular with ± central,
	glistening homogeneous globule, immediately fugacious, the
	globule fragmenting on drying
18	Cortex cells of stem mostly yellowish brown to brown,
	partly tinged red; medulla cells mostly ± infected by
	mycorrhiza and therefore appearing grey; leaves orbicular,
	oval to broadly oval, often ± concave, lobe apices inflexed,
	leaves thus appearing imbricate or shoots appearing
	succulent; gemmae mostly abundant; teeth of perianth
	mouth consisting of 1–2 cells; on soil, rocks and among
	bryophytes of siliceous substrates in open to shaded
	habitats

Key to species of Diplophyllum and the genus Scapania

- Postical lobe up to twice as long as wide, variously shaped, rarely lingulate or parallel-sided, not secund; antical lobe variously shaped, antical and postical lobes at similar angle to stem (except *S. umbrosa*); median leaf cells isodiametric, not differentiated; gemmae ovoid to elliptical; perianth ± compressed, mostly not plicate, truncate at mouth... *Scapania*

Key to species of *Scapania*

Based on Damsholt (2002), Meinunger and Schröder (2007)

1 Postical lobe of leaves not conspicuously decurrent or only decurrent to the level of the insertion of the keel, or in some cases with a decurrent strip only 1 cell wide, gradually Postical lobe of leaves distinctly decurrent, with the decurrent strip several cells wide, gradually tapering and visible for ca half of the length of the segment below the insertion of the keel: leaves often dentate 12 Antical lobe (0.6) 0.7–0.8 the length of the postical lobe; 2 gemmae 2-celled; usually both lobes standing away from each other; cuticle with hemispherical papillae; calcicole...... Scapania aeguiloba (rr) Antical lobe 0.3–0.65 the length of the postical lobe; gemmae 1–2-celled 3 Postical lobes narrow, 1.3-2 times as long as wide; plants 3 Postical lobes ovate to almost rotundate, 0.8-1.3 times as long as wide; plants 1.8-5 mm wide and 20-100 mm long; gemmae greenish *Scapania irriqua* (r) Leaves bordered by (1) 2-4 rows of equally thick-walled 4 cells lacking trigones 5 Marginal cells of lobes not differing from median leaf cells and thus not forming a border, except at the base of the postical lobe, where 1-2 rows of thick-walled cells may

5	Gemmae greenish to brownish; base of postical lobe never with vinaceous secondary pigmentation; cuticle with
	hemispherical papillae; calcicole
-	Gemmae greenish, 2-celled; base of postical lobe often with vinaceous secondary pigmentation; leaves keeled almost to
6	the base (sect. Curtae)6 Postical lobe rounded at apex; marginal cells of the leaves
O	(15) 16–22 μ m; perianth mouth with teeth 1–3 cells long wide entire sections between teeth lacking; keel 0.5–0.6 the
	length of the postical lobe; antical lobe not directed towards
	shoot apex
-	Postical lobe often acute; marginal cells of the leaves 12–18
	μm; perianth mouth entire; keel to 0.3–0.4 the length of the
	postical lobe; antical lobe directed towards shoot apex
	Scapania parvifolia (rr)
7(4)	Gemmae 1-celled, black or reddish-brown, formed at apex of
	erect shoots with reduced leaves; shoots to 1 mm wide and
	2–3 mm long; only on decaying logs
	Scapania apiculata (n.s.)
-	Gemmae (1) 2-celled, greenish, vinaceous or brown; shoot
_	mostly more than 1 mm wide and more than 3 mm long8
8	Cells with 2–5 (6) large, persistent oil-bodies, $10-14 \times 8-11$
	μm; gemmae green to brownish at maturity, 16–24 μm wide;
	perianth inflated and plicate towards the mouth; antical lobe
	large, 0.4–0.6 the size of the postical lobe, often distantly
	dentate; cuticle with hemispherical papillae; calcicole
	Scapania calcicola (r)
-	Cells with smaller, non-persistent oil-bodies, of which the
	largest are 9–10 × 5 μ m, gemmae colourless or greyish or
0	reddish to vinaceous, rarely yellow-brown (sect. Curtae)9
9	Plants pale green or reddish to purplish brown, when
	exposed; postical lobes mostly rounded at apex, antical lobes
	almost always acute; cells with 2–6 often large greyish oil-
	bodies, occluding lumen; gemmiparous leaves strongly
	dentate; perianth mouth entire (or with single, distant small
	teeth: var. argutedentata)
_	Plants with brownish secondary pigmentation, never with
	vinaceous or reddish postical lobe bases or gemmiparous
	leaves; leaves edentate or only dentate near gemmiparous areas or distantly dentate, but the dentition never reaches
	areas or distanció dentate, but the dentition never reaches

	the basal 0.2 of the leaf; perianth mouth laciniate to dentate
10	Postical lobe rounded, lingulate; marginal cells of lobes 19–25 μ m; leaves often with scattered teeth; cells of leaf middle with 6–12 oil-bodies; perianth mouth with teeth of 1–7 superimposed cells; gemmiparous lobes often dentate
_	Postical lobe apiculate, rarely rounded, marginal cells of
	lobes 13–20 µm; leaves edentate or only denticulate near gemmiparous areas; cells of leaf middle with 2–6 oil-bodies; perianth mouth with teeth of (1) 2–6 superimposed cells, terminating lobules 2–5 cells wide at base
11	Gemmae greenish, rarely reddish, 16–29 × 8–14 μm; antical
	lobe ca half as large as postical lobe; leaves entire; perianth
	mouth laciniate-dentate <i>Scapania mucronata</i> (r)
-	Gemmae yellowish-brown when ripe, $18-36 \times 10-19$ µm;
	antical lobe <i>ca</i> 0.65 the size of the postical lobe; leaves occasionally with minute dentition; perianth mouth with many teeth, each with several superimposed cells
	Scapania praetervisa (r)
12 (1)	Antical lobe longly or shortly decurrent
-	Antical lobe not decurrent, transversely inserted14
13	Antical lobe longly decurrent; cuticle verruculose, especially at marginal teeth; gemmae 1-celled, brown
	Scapania nemorea (w)
-	Antical lobe shortly decurrent; cuticle coarsely verruculose,
	with hemispherical papillae; gemmae 1-2-celled, whitish
	green Scapania aspera (r)
14	Shoot 5–12 mm long; gemmae reddish brown; leaf lobes
	elliptical-lanceolate, dentate; keel winged; on decaying logs
	or humus
-	Shoot 15–100 mm long; gemmae yellow-green or pinkish, sometimes totally lacking, on wet or submerged stones in
	streams
	Su eamsscupunia unautata (1)

Supplementary 'key' according to habitats

Based on Nebel and Philippi (2005)

1 wet habitats (swamps, springs, streams):

- *S. irrigua*: leaf lobes not decurrent, plants green
- S. undulata: postical lobe long decurrent, antical lobe not decurrent, plants sometimes pigmented
- 2 limestone rocks:
- *S. aequiloba*: antical lobe crossing stem; lobes entire
- *S. aspera*: antical lobe crossing stem; lobes mostly dentate
- S. calcicola: antical lobe not crossing stem; lobes edentate to serrate-dentate

3 soil:

- **S. nemorea**: robust species with dentate lobes and mostly conspicuous brown gemmae
- *S. curta, S. mucronata, S. scandica, S. lingulata*: small species, difficult to distinguish, see key 6 and 9–11
- 4 decaying wood:
- S. apiculata: small plants with erect attenuate-leaved gemmiparous shoots
- *S. umbrosa*: small plants with dentate leaf lobes
- (*S. nemorea*): large plants, usually on soil, see above

Key to species of Calypogeia

- Leaves bidentate with divergent and usually very acute teeth; cuticle striolate-verruculose..... Calypogeia arguta (rr) Note: missing in Erzberger and Papp (2020).
 Leaves rounded at apex or bidentate, but teeth not

- Underleaves entire, emarginate or 2-lobed to 1/3, with 4–14 rows of cells between sinus and rhizoid area.....
- 3 Mature shoots rarely more than 2 mm wide; on decaying coniferous wood and raw conifer humus, rarely on peat; lateral leaves imbricate; underleaves 2–3.5 times as wide as stem, 2-lobed, outer margins often with small obtuse tooth....

4	Mature shoots often more than 2 mm wide; on loamy or sandy-loamy soil, non-calcareous rocks, humus, peat, and in swampy or boggy habitats; if on wood then underleaves to 2.5 times as wide as stem, lacking lateral teeth
-	Oil-bodies azure blue (when fruiting, seta therefore blue-hyaline). Lateral leaves obtuse to rounded, mostly widest near base. Underleaves divided into 2 mostly obtuse, triangular lobes, outer margins without knobs or teeth
5	Calypogeia azurea (r) Lateral leaves mostly as long as wide, ± translucent; underleaves with (2) 4–6 rows of cells between sinus and rhizoid area, these cells 30–80 μm long; oil-bodies present in all underleaf cells; shoots 1.5–3.5 (4) mm wide. Calypogeia muelleriana (r)
-	Lateral leaves 1–1.3 times as long as wide, rather opaque, often with translucent margins; underleaves with 7–14 rows of cells between sinus and rhizoid area, these cells 20–50 μm long; oil-bodies soon lost from all or most underleaf cells; shoots not > 3 mm wide
6	At least some lateral leaves with tangentially elongated marginal cells forming a continuous border; underleaves decurrent at base; rhizoid area shallowly linear; oil-bodies present only in 1–3 marginal cell rows of lateral leaves, stem cells lacking oil-bodies; shoots 0.7–1.8 (2.5) mm wide
_	Marginal cells of lateral leaves not tangentially elongated or rectangular and forming a discontinuous, ill-defined border; underleaves not or slightly decurrent; rhizoid area deeply oval or suborbicular; oil-bodies present in nearly all cells of lateral leaves and in stem cells; shoots 1.5–3.0 mm wide

Key to species of Marsupella

- 1 Plants robust, (0.5) 1–5 cm long; lateral leaves boat-shaped, when flattened rounded-quadrate to ± orbicular; lobed to 1/20-1/5 (1/4), basal part of leaf not sheathing stem; orientation of leaf sinus parallel to stem when viewed from side; leaf lobes obtuse or rounded and apiculate; leaf margin usually narrowly recurved or revolute on one or both sides Plants small, up to 1 (1.5) cm long, lateral leaves bilobed to (1/5) 1/4-1.2; lobes of apical leaves acute; leaf margins plane 2
- Paroicous, plants mostly with sporophytes; lateral leaves 2 increasing in size towards apex of stem; fertile plants clavate; plants 1-5 (7) mm long; lateral leaves erectspreading, without sheathing base, rounded-quadrate when flattened; sinus acute to rectangular.....

Marsupella sprucei (rr)

Dioicous, plants rarely fertile; lateral leaves of \pm equal size; plants larger, shoots 5-15 mm long; trigones scarcely

Key to species of Nardia

- Plants green, light-green to brown-green, lateral leaves rounded, orbicular, not lobed or incised, not erectappressed; oil-bodies hyaline, glistening, homogeneous, persisting several years after drying, mostly 2 per cell, often segmented; perigynium not bulbous or sac-like; trigones small to medium-sized and bulging; shoots 0.5-3 (6) cm
- Plants green, brown to red-brown, lateral leaves nearly orbicular, retuse to emarginate or entire; oil-bodies granular, opaque, not persisting; perigynium bulbous or saclike, forming marsupium (but sometimes not); underleaves only rudimentary, except at apex of fertile shoots; paroicous

Key to Jungermanniaceae group (including *Endogemma, Jungermannia, Liochlaena, Nardia, Solenostoma, Syzygiella*)

1	Lateral leaves rounded-rectangular, flat (not concave) spreading in lower part of stem, appressed towards apex
	perianth mouth longly ciliate; male bracts with small
	dentate antical lobe; plants in green to dark green, ofter
	reddish or copper-coloured turfs, also on rotting wood
	Syzygiella autumnalis (Jamesoniella autumnalis) (r)
-	Leave arrangement different; perianth mouth at most
	crenulate; male bracts entire2
2	Underleaves lanceolate, distinct, usually restricted to
	younger parts of stem; oil-bodies large, homogeneous
	weakly segmented, or granular, 1 to several per cell
	perianth very short, immersed in bracts
-	Underleaves absent (stems of Solenostoma hyalinum and
	Endogemma caespiticia occasionally with a few
	underleaves); oil-bodies granular, not homogeneous, few or
	many per cell; perianth exserted from female bracts4
3	Plants green, light-green to brown-green, lateral leaves
	rounded, orbicular, not lobed or incised, not erect
	appressed; oil-bodies hyaline, glistening, homogeneous
	bean- or sausage-shaped, persisting several years after
	drying, mostly 2 per cell, often segmented; perigynium no
	bulbous or sac-like; trigones small to medium-sized and
	bulging; shoots 0.5–3 (6) cm long; dioicous
	Nardia scalaris (rr)
-	Plants green, brown to red-brown, lateral leaves nearly
	orbicular, retuse to emarginate or entire; oil-bodies
	granular, opaque, not persisting; perigynium bulbous or sac-
	like, forming marsupium (but sometimes not); underleaves
	only rudimentary, except at apex of fertile shoots; paroicous
4	Female bracts and perianth arising from well-developed
	perigynium 0.2-0.6 length of perianth bearing 1 pair of
	bracts; perianth conical, plicate above, emergent to about
	1/2 from bracts, cells in upper part 2-3 times as long as
	wide; at least some rhizoids purple or violet, sometimes
	brownish or colourless; underleaves absent, occasionally

	single at stem apex, lanceolate; leaves reniform to semicircular, rounded to weakly retuse; plants procumbent;
	dioicous; leaf cells 25–40 μ m (30–40 × 25–35 μ m)
_	Perigynium absent, or, if present, short; perianth cylindrical
	to ovate, with or without plicae above, exceeding bracts,
	cells in upper part not elongate, similar to leaf cells; rhizoids
	colourless to brownish, not violet; underleaves absent on
	sterile shoots
5	Lateral leaves ± lingulate, distinctly longer than wide (1.2–
_	1.7 times); perianth emergent, cylindrical, smooth, abruptly
	contracted to beaked mouth6
_	Lateral leaves orbicular, ovate to reniform, cordate, as long
	as wide or slightly longer (1.3 times); perigynium absent or
	only short; perianth emergent, plicate above, not beaked
	(Jungermannia) or contracted to beaked mouth
	(Solenostoma, Endogemma)7
6	Plants 4–6 mm wide, nearly always with cylindrical
	perianths, gemma-bearing shoots very rare or completely
	lacking; paroicous; on shaded, humid, limefree, but base-rich
	to moderately acidic substrates
	Liochlaena lanceolata (Jungermannia leiantha) (r)
	Note: According to Meinunger and Schröder (2007), the following
	characters are not reliable for the distinction between <i>L. lanceolata</i> and <i>L.</i>
	subulata: <i>L.</i> lanceolata – only rarely with gemma-bearing shoots; leaf cells up to 50 (65) μm wide; leaves in situ resembling those of <i>Plagiochila</i> , vs. <i>L.</i>
	subulata – gemma-bearing shoots more frequent; leaf cells up to 40 (50)
	μm wide.
-	Plants 2–3 mm wide, usually sterile or with young pyriform
	perianths not emerging from bracts; gemma-bearing shoots
	very frequent; dioicous
	Liochlaena subulata (Jungermannia subulata) (rr)
7	Plants with many small-leaved innovations (often only these
	are found), well-developed lateral leaves bordered with one
	row of somewhat larger, thick-walled cells, orbicular,
	concave (leaves of small plants and of small distant-leaved
	innovations are often unbordered); dioicous; trigones small
	or absent; plants up to 1 mm wide
	Solenostoma gracillimum (Jungermannia gracillima) (r)

Plants without small-leaved innovations; margins of lateral leaves unbordered or only indistinctly bordered, marginal cells not larger than adjoining leaf cells; dioicous or paroicous; trigones mostly present......8 Lateral leaves elliptical, ovate, cordate, mostly slightly 8 longer than wide (up to 1.3 times); perianth weakly plicate. not beaked, mouth eventually divided into 4 short, toothed Lateral leaves as wide as long or slightly wider than long, reniform to orbicular; perianth with 4-5 plicae above, ± abruptly contracted to beaked mouth, mouth eventually Dioicous; plants to 0.5-4 (5) cm long; lateral leaves ovate, 9 0.6–1.8 mm long and 0.6–1.5 mm wide..... Jungermannia atrovirens (rr) Paroicous; plants 0.3-1 (2) cm long; lateral leaves 0.3-1.2 mm long and 0.3-1 mm wide; perianth clavate to cylindrical, plicate above, ± acute: lateral leaves on fertile stems broadly elliptical to broadly ovate; mid-leaf cells 16-28 (32) µm 10 Dioicous; gemmae frequent, 1-celled, forming dense mass at shoot apices; trigones absent; oil-bodies only 1 (2) per cell, fugacious; small plants, shoots < 1 cm long, in yellowish green patches; usually ephemeral, with many perianths.....Endogemma caespiticia (Jungermannia caespiticia) (n.s.) Paroicous; gemmae absent; trigones present, usually distinct; several oil-bodies per cell; without perigynium; plants dark green, in green to greenish-black tufts, without small-leaved innovations........Solenostoma sphaerocarpum (Jungermannia sphaerocarpa) (rr)

Key to species of Mesoptychia

-	Plants lacking both gemmae and attenuate shoots with appressed leaves; underleaves absent or with 1 to several	
2	marginal cilia	
	Mesoptychia badensis (Leiocolea badensis) (r) Note: Trigones may be inconspicuous in lateral leaves, but usually are more pronounced in bracts and perianths.	
-	Underleaves present, but hidden among the rhizoids, small and often difficult to detect; plants usually strongly aromatic; lateral leaves mostly 0.5–1.3 mm long, 2-lobed to 1/5 (1/3); mid-leaf cells 20–32 μ m wide; on calcareous substrates, damp soil, rocks, loess	
Key to species of Lophocoleaceae		
1	At least lowest lateral leaves 2-lobed or, if all lateral leaves rounded to retuse, antical margin not or hardly decurrent; perianth well developed, apical on main stems or on ± full-sized branches; calyptra shorter than perianth; paroicous or dioicous; underleaf lobes mostly divergent; underleaves free or connate with a lateral leaf; strongly aromatic. <i>Lophocolea</i>	
-	Mature lateral leaves rounded to retuse, rectangular, rarely lateral leaves of side branches 2-lobed, antical margin decurrent; perianth on short side branches; calyptra protruding beyond perianth; autoicous; underleaf lobes generally nearly parallel; underleaves free; not strongly aromatic. <i>Chiloscyphus</i>	
2	Apical leaves distinctly 2-lobed or apex 2-dentate; mid-leaf cells 25–50 µm wide; caducous flagelliform branches absent; leaf insertion not or scarcely arcuate postically; plants 1–6	

conditions, it is perhaps appropriate to deal with the collective taxon.

taxon must be called L. bidentata (L.) Dumort.

4 Paroicous, perianths usually abundant; lowermost lateral leaves and leaves of branches 2-lobed to 1/3, apical leaves retuse or emarginate; gemmae rare, mostly 1-celled......

.....Lophocolea heterophylla (c)

- 5(1) Plants dark to greyish green; median leaf cells 30--35 (40) × (21) 24--30 µm; each cell with (1) 2--4 (5) oil bodies; each oil body $7\text{--}12 \times 4\text{--}7$ µm; lateral leaves broadly rounded to truncate or weakly retuse, those of branches rarely 2-lobed; lobes of perianth mouth entire or obscurely dentate; calyptra long emergent from perianth apex; plants closely appressed to mostly siliceous rocks by running water......

Note: According to Meinunger and Schröder (2007), the best differentiating character between the two species is cell size. According to Damsholt (2002), the differences in the dentition of the perianth mouth

are not significant. Cell size is variable in both species, small leaves have small cells, larger leaves have larger cells.

Key to species of Plagiochilaceae

- 1 Lateral leaves ± plane, margin entire or retuse or with 2-4 apical teeth; underleaves small, filiform, limited to young shoots; shoots usually with rhizoids ± to apex; monoicous; leaves not inserted to the antical stem mid-line, but with a leaf-free zone 2-4 cells wide: leaves often variable in size or shape even on a single stem Pedinophyllum interruptum (w) Lateral leaves usually convex, usually dentate; underleaves absent or only present at apex as cell filaments; rhizoids rare or absent; dioicous; leaves inserted to antical mid-line 2 2 Plants up to 10 cm long and 5-9 mm wide; lateral leaves 3- $4.5 \times 2.5-4.5$ mm, mostly dentate with numerous teeth, sometimes ± entire; mid-leaf cells 30-33 (35) µm wide, trigones very small or absent; flagella absent..... Plagiochila asplenioides (w) Plants 2–5 cm long and 1.5–5.5 mm wide; lateral leaves 3 \times 2.5 mm, dentate to nearly entire; mid-leaf cells 25-30 (33) μm wide; trigones small to medium-sized; flagella present.....
- Key to species of *Frullania*

- 2 Antical lobes ± acute to apiculate (at least on branch leaves),

concave when moist, with generally long line of ocelli, additional ocelli often present, scattered or forming small discrete group; underleaves on main stems decurrent, margin recurved, base auriculate to crispate; leaves not caducous; lobe cells (10) 12–23 µm wide; plants up to 10 cm long, reddish brown to copper coloured..... Frullania tamarisci (r) Antical lobes (even of branch leaves) always rounded, plane when moist; ocelli of antical lobes scattered, at most short line present at base of lobe, ocelli often pale brown; underleaves on main stems not decurrent, divided to 1/3-2/5, often with lateral tooth on both sides, margins plane, entire: lateral leaves sometimes caducous: lobe cells 16-22 (25) µm wide; delicate species growing in neatly delimited patches; with the smell of roses when rubbed with a moist finger *Frullania fragilifolia* (r) Underleaves on main stem reniform, much wider than long, 2.5–3.5 times as wide as stem, entire or retuse; mid-leaf cells with strongly bulging trigones, colourless..... Frullania jackii (n.s.) Underleaves on main stem longer than wide, at most twice as wide as stem, narrowly incised with an acute sinus; lobes acute......4 Dioicous, but perianths common, trigonous, tuberculate; antical base of antical lobes on main stem auriculate; midleaf cells of antical lobes mostly with strongly bulging trigones and intermediary thickenings; postical lobe helmetshaped or evolute, as long as wide or wider than long; stylus (triangular flap between lobule and stem) mostly large, lanceolate, 3-10 cells wide at base; underleaves with acute lobes and 1–2 blunt teeth on lateral margins..... Frullania dilatata (cc) Autoicous, perianth with 5 plicae, smooth externally; antical base of antical lobes on main stem not auriculate; mid-leaf

3

4

cells of antical lobes with small trigones and generally without intermediary thickenings; postical lobes evolute, lanceolate, < 1/2 width of antical lobe; antical lobes (on main stems) plane, mostly broadly ovate to ovate; leaf cell walls thin, colourless; plants mostly green......

Key to species of Lejeuneaceae

1	Leaf cells smooth; oil-bodies simple, numerous, persistent
- 2 -	At least cells of antical lobe distinctly conically mamillose2 Postical lobe with smooth cells <i>Cololejeunea calcarea</i> (r) Postical lobe with cells conically mamillose
Key	to species of <i>Porella</i>
1	Leaves <i>in situ</i> with antical margin neatly appressed, margins of postical lobes and underleaves ± spinose-dentate, at least on branches; antical lobes ovate, apex acute; plants with strong metallic sheen, glossy; tasting acrid ————————————————————————————————————
-	Leaves <i>in situ</i> with antical margin of at least some leaves elevated or recurved, postical lobes of lateral leaves and underleaves entire or only weakly dentate and distantly toothed (sometimes with toothed decurrent wings); antical lobes mostly not acute; plants dull; not tasting acrid
2	Postical lobe of lateral leaves tapering to acute apex, small, not more than 0.2 times as wide as antical lobe, narrower than stem; underleaves dentate at base, longly decurrent; mid-leaf cells of antical lobe 28–40 µm wide
	Postigal laboratoridad et en eu mastly (0.2) 0.2 0 (times es
_	Postical lobe rounded at apex, mostly (0.2) 0.3–0.6 times as wide as antical lobe, as wide or hardly wider than stem, to about 1/2 as wide as underleaves; shoots irregularly 2–3-pinnately branched; plants dull, not glossy
3	Postical lobes and underleaves longly decurrent; cells in the middle of the antical lobe (36) 38–40 µm in diameter; perianth mouth long-dentate to densely short-ciliate; female bracts dentate
_	Postical lobes and underleaves not longly decurrent; cells in the middle of the antical lobe 32–35 µm in diameter; perianth mouth somewhat dentate; female bracts almost edentate

Key to species of Radula

- Paroicous, perianths common; male bracts saccate at base in
 (1) 2-4 pairs below female bracts... Radula complanata (cc)
- Dioicous, perianths very rare; male inflorescences spicate, closely imbricate, with 10–20 leaf pairs (but male plants rare)
 Radula lindenbergiana (n.s.)

Key to species of Aneuraceae

- 2 Thallus light-green, strap-like, irregularly pinnately branched, 1–2 cm long and 0.5 mm wide; dorsal side of thallus flat to concave, typically channelled, semi-orbicular in section; gemmae often present; dioicous......

- 3 Thallus palmate; thallus branches dark green, 0.2–0.3 mm wide, densely arranged; dioicous..........*Riccardia palmata* (r)
- 4 Upper part of thallus ± regularly 2–3-pinnate, branches strap-like, 0.3–1 mm wide, biconvex in section; thallus margins unistratose, translucent, 2–3 cells wide; epidermal and marginal cells without oil-bodies (in fresh material)..........

Upper part of thallus irregularly pinnate or palmate, dorsal surface flat to weakly convex in section; thallus branches 1-2 mm wide: unistratose margins mostly absent or only 1-2 cells wide; epidermal and marginal cells with or without oil-Thallus irregularly 1–2 (3)-pinnate; branch apices rounded; 5 Thallus irregularly branched, branches narrowed at base; branch apices with heart-shaped incision; oil-bodies absent. Key to species of *Metzgeria* 1 Thallus densely covered with hairs on both sides, without gemmae or adventive thalli *Metzgeria pubescens* (Apometzgeria pubescens) (r) Thallus without hairs or hairs confined to margins and ventral surface of midrib; with or without gemmae and adventive thalli 2 Thallus 2-3 cm long and 1-2 mm wide; margins strongly down-curved; adventive gemmae not present on thallus margins; hairs of ventral surface of midrib bristly; hairs of thallus margins ± straight, often in pairs; thallus cells 45–65 (*Metzgeria conjugata* subsp. *conjugata*) (w) Thallus up to 1 (2.5) cm long and 0.4-1.0 (1.5) mm wide; adventive thalli and / or gemmae present or not; ventral surface of thallus not or hardly hairy; hairs of thallus Thallus branches linear, of equal width throughout; margins 3 ± plane when moist; thallus margins sometimes with adventive thalli; thallus never blue........ Metzgeria furcata (c) Thallus branches attenuate towards apices, bearing gemmae on both faces of thallus as well as on margins; margins recurved when moist; midrib mostly 4-5 cells wide on each surface; thallus becoming blue upon drying.....

.....Metzgeria violacea (Metzgeria fruticulosa auct.) (excluded)

Key to species of Fossombronia

1	Spores with 5–7 alveolae across the convex face; alveolae 8–12 µm wide, regular
-	Spores not alveolate, ornamented with lamellae free or anastomosing in the centre, ± parallel at least on the sides even if reticulate towards the centre2
2	Spores in side view with 5–7 lamellae 6–10 µm apart; spore margin with 15–19 spines in profile; wing often translucent. <i>Fossombronia pusilla</i> (w)
_	Spores in side view with 10–13 lamellae 6–10 µm apart and
	with 25–35 (40) spines in profile
	Fossombronia wondraczekii (r)
Key t	to species of Pelliaceae
1	Slime papillae restricted to ventral side of costa near apex, on stalk 1–5 (6) cells long; thalli with miniature apical branches proliferating in autumn; dioicous; involucres
	subhorizontal to erect, tubular, with dentate-ciliate lobes; male tubercles <i>ca</i> 200 μm wide, cells surrounding aperture not papilliform; calcicole
	Apopellia endiviifolia (Pellia endiviifolia) (w)
-	Slime papillae on stalk 1 cell long, on both ventral and dorsal
	side of costa near apex; caducous branches absent; dioicous or monoicous; calcifuge
2	Monoicous and normally paroicous; involucres nearly
	horizontal, flap-like, subentire or irregularly lobed; male
	tubercles to 400 µm wide, aperture surrounded by

 Key to Marchantiales pp. (incl. *Asterella, Clevea, Conocephalum, Lunularia, Mannia, Marchantia, Reboulia*)

1	Dorsal side of thallus nearly always with gemmae cups, distinctly reticulate; female receptacles stellate, on stalks (archegoniophores)
-	Thalli without gemmae cups, reticulate or not4
2	Gemmae cups semi-lunate; air-pores volcano-shaped, simple; female receptacles cruciate, 4-rayed
	Lunularia cruciata (r)
-	Gemmae cups goblet-shaped, circular; air-pores barrel-
	shaped, each composed of 4 (6) rings of superimposed cells;
	female receptacles at apex of thallus, stellate, deeply divided
	with usually (5) 8-9 (11) terete lobes; thallus dark green,
	often with ± distinct blackish median band on dorsal
	surface; ventral scales colourless, not conspicuous, of 3
	different forms; spores 9–13 μm in diameter3
3	Thallus prostrate, 6-9 mm wide, densely branched; with a
	discontinuous, dark median band on dorsal surface of
	thallus; appendages of median scales sharply toothed;
	gametangiophores frequent; dorsal epidermis usually at
	least partly verruculose; gemma cups with 7-13 gemmae,
	each with 50-60 marginal oil cells
-	Thallus usually growing erect, > 9 mm wide, distantly
	branched; with a long, conspicuous, dark median band
	(lacking air-chambers) on dorsal surface of thallus; margins
	of appendages of median scales entire or crenulate;
	gametangiophores infrequent; dorsal epidermis smooth;
	gemma cups with 22-25 gemmae, each with 70-75 marginal
	oil cellsMarchantia polymorpha subsp. polymorpha (r)
4 (1)	Air-pores barrel-shaped; archegonia and antheridia born on
	stalked receptacles; air-chambers with branched
	chlorenchyma (chlorophyllose filaments); female receptacle
	a disc with 4 ridges (if with 8-9 lobes, see Marchantia
	polymorpha), with stalk 5–10 cm long, male receptacles with
	stalk 1–2 cm long; thallus ± carinate; ventral scales purplish,
	soon vanishing; taste pepperish (after initial sweetness)

Air-pores elevated and volcano-shaped, or simple, or absent, not barrel-shaped; antheridia on sessile receptacles or immersed in the thallus: female receptacles stalked, rarely nearly sessile 5 chlorenchyma 5 Air-chambers with (chlorophyllose filaments); thallus very large, up to 1–2 cm wide and 15 cm regularly dichotomously branched, reticulate, green to dark green, with a strong smell of turpentine; air-pores elevated, volcano-shaped, simple, with 5-6 concentric rings, each ring with 7-8 cells; female receptacles conical, stalk to 6 (10) cm long, arising at the chlorenchyma (chlorophyllose Air-chambers without filaments), but often with secondary walls, or air-chambers absent; if air-chambers with chlorenchyma, then thallus Dorsal surface of moist thallus distinctly glossy, to the naked 6 eve the pores are more prominent than the lines: hvaline margin of thallus 3-4 cells wide; air-chambers between costa and thallus margin usually in 6-7 rows; boundaries of air-chambers on surface of thallus level with the surface (not recessed); epidermis of air-chambers of archegoniophore Dorsal surface of moist thallus dull, to the naked eye the lines are more prominent than the pores; hyaline margin of thallus 1-2 cells wide; air-chambers between costa and thallus margin usually in 4-5 rows; boundaries of airchambers on surface of thallus recessed (forming furrows); epidermis of air-chambers of archegoniophore heads unistratose Conocephalum salebrosum (w) Radial walls of air-pores thin (partially thickened in 7 Reboulia hemisphaerica); stalk of female receptacle with 1 rhizoid furrow; each receptacle with (2) 3-8 sporophytes...8 Radial walls of air-pores thickened, air-pores therefore appearing stellate; stalk of female receptacle without rhizoid furrow; each receptacle with (1) 2-6 sporophytes; archegoniophores arising from middle part of dorsal surface of thallus; thallus not coriaceous, mostly < 5 mm wide, not glaucous; ventral side of thallus pale green; ventral scales

- Air-pores with (1) 2–4 rings of 4–8 (9) guard cells; cells thin-walled; female receptacles spherical to conical......9
- Female receptacles ± conical; each sporophyte surrounded by involucre and laciniate pseudoperianth; colourless ventral scales projecting beyond thallus margin at thallus apex; spores brown, not alveolate; thallus 2–3 mm wide; pseudoperianth divided into 6–8 lanceolate lobes, connected at tips; crushed fresh thallus with a faint smell of rotten fish; cells of thallus epidermis thin-walled, polygonate......

Thallus delicate; dorsal surface reticulate, becoming slightly lacunose with age; air-chambers broad; walls of chlorenchyma without lateral chlorophyllose filaments; ventral scales small, distant, light-green or hyaline; stalk of

Key to species of *Riccia* **group** (incl. *Oxymitra, Riccia,* and *Ricciocarpos*)

Important references: Müller (1951–1958), Szweykowski and Mendelak (1964), Jovet-Ast (1986), Gradstein and van Melick (1996), Schumacker and Váňa (2000), Damsholt (2002), Meinunger and Schröder (2007), Hugonnot (2010)

- Thalli floating on water; ventral scales large, in several ill-defined rows, strap-shaped, with serrulate margins, pendent in water; plants sometimes also terricolous with small ventral scales; epidermis and ventral scales with scattered dark oil cells, chlorenchyma with large air-chambers; air-pores present; sporophytes very rare......

- 3 Dorsal tissue of thallus similar to an epidermis, consisting of fully connected cells with chlorophyll, and with air-pores above air-chambers (aquatic forms partly without either), sometimes dorsal tissue spongiose, with air-chambers becoming exposed by decay of epidermal cells; chlorenchyma mostly chambered. Subgenus Ricciella............4

-	Dorsal chlorenchyma consisting of cell columns with an upper epithelium of one or two layers of hyaline cells without sharply delimited air-pores (air-spaces opening
	through gaps between hyaline cells); dorsal tissue of lobes compact. Subgenus Riccia9
4	Thallus not spongiose, with air-chambers; dorsal surface of older parts of thallus not lacunose; epidermis with inconspicuous pores; capsule protuberant and opening on ventral side of thallus. Sect. Ricciella
-	Thallus spongiose, with ± large cavities on dorsal surface, at least in older parts; capsule immersed, or ± protuberant on ventral or dorsal surface and opening on dorsal side. Sect. Spongodes
5	Dorsal surface of thallus conspicuously reticulate, at least at apex of lobes; thallus lobes 0.5–1.5 (2) mm wide, dorsal surface flat, in cross section 3–8 times as wide as high, apices somewhat enlarged; thallus without or with only inconspicuous ventral scales and rhizoids
-	Dorsal surface of thallus inconspicuously reticulate; thallus lobes up to 0.5 mm wide, in cross section 1.5–2.5 times as wide as high, with a dorsal furrow (best seen in dry material), apices somewhat narrowed; ventral scales towards thallus apices distinctly 2-rowed; rhizoids abundant; on muddy soil
6	Dorsal surface of thallus often finely reticulate, at least towards apex; areolae up to 0.3 mm in diameter; thallus lobes in cross section 3–6 times as wide as high, 0.5–1.5 mm wide, forming an angle of 40° , somewhat enlarged at apex; marginal cells of thallus 30 – 45×12 – 20 µm; almost always sterile; submerged in standing water, more rarely as a terricolous form on periodically inundated muddy soil
-	Dorsal surface of thallus more coarsely reticulate; thallus lobes in cross section 5–8 times as wide as high, up to 2 mm wide, forming angle of 90°, rounded at apex; marginal cells of thallus 40–50 × 25 µm; capsules unknown; in similar sites to <i>R. fluitans</i>
7(4)	Branches of young and adult thalli elongate, up to 1 mm wide, lobes up to 3 times as wide as high; thalli not in rosettes or rosettes indistinct; adult thalli vivid green, often

	ventral surface of thallus and spores exiting through ventral epidermis; distal face of spores alveolate
-	Lobes up to 2 mm wide, 3–6 times as wide as high; thalli in rosettes, disc-shaped or fan-shaped, pale green to bluish green, sometimes tinged with pink; capsules protuberant on dorsal surface of thallus and spores exiting through dorsal epidermis; distal face of spores with ± parallel to divergent
8	ridges or irregularly alveolate
-	Riccia frostii (w) (Figure 1, p.4) Monoicous; thallus often forming circular disc, green, rearly tinged pink; divided to half its length, with lobes 1–2 mm wide; cavities large; spores reddish brown, 60–120 μm in diameter, distal face with irregular ridges and alveolae Riccia cavernosa (w)
9 (3)	Thallus lobes winged, 4–8 mm long, 2.5–3 (4) mm wide, mostly nearly as wide as high; perennial, mostly dioicous taxa; thallus lobes short, transversely truncated; terminal
-	cell of columns elongate, bottle-shaped
10	Thallus lobes bluish green, distal parts thickened and tuberous; spores 110–180 μm in diameter, distal face with 5–7 ± indistinct alveolae, restricted to polar region; cells of ventral scales 33–45 x 70–90 μm
-	Thallus lobes green to yellowish green; thallus apices with stalked bulbils (not always present); spores 130–215 µm in diameter; distal face of spore with 8–20 alveolae; cells of ventral scales 50–65 x 100–120 µm <i>Riccia gougetiana</i> (r) Note: Meinunger and Schröder (2007) doubt that <i>R. gougetiana</i> is a good species. According to them, difference in thallus colour between <i>R. ciliifera</i> ("female thallus bluish-green, deeply spotted with violet on all faces and

on scales") and *R. gougetiana* ("female thallus yellow-green with brownish-yellow wings rarely spotted with violet on the faces and on scales" (Schumacker and Váňa 2000: 141) are considered unreliable since they probably are related to environmental conditions and different age of plants. In the sites, where both "species" occur in Germany, no significant ecological or sociological differences can be found (Marstaller in Meinunger and Schröder 2007). Spore size (*R. ciliifera*: 110–180 μm; *R. gougetiana*: 130–220 μm; Schumacker and Váňa 2000: 141) seems to vary continuously, the few fruiting German plants had spores 120–160 μm. The only character that separates these entities seems to be cell size of ventral scales (*R. ciliifera*: 33–45 x 70–90 μm; *R. gougetiana*: 50–65 x 100–120 μm; Müller 1951–1958: 435, 439), a difference that remained constant in cultivation experiments (Szweykowski and Mendelak 1964).

Sporophytes are rarely produced in Hungary, but spore morphology seems to be sufficiently different, compare the illustrations in Jovet-Ast (1986).

-	Thallus normally glabrous, but sparse short cilia may be rarely present in <i>R. glauca</i> var. <i>ciliaris, R. warnstorfii</i> var.
	ciliaris, R. subbifurca and R. bifurca var. subinermis
14	Thallus glistening when fresh (not visible in herbarium specimens); margin of lobes ± densely ciliate with cilia up to 1000 µm long; spores very finely papillose (difficult to see in
	light microscope), dark brown to blackish; distal face with
	7-12 alveole across the diameter, each (6) 8-12 (14) μm
	wide, wing (at the margin of the spores) narrow or lacking
	Note: R. ciliata var. epilosa: Thallus lobes without cilia.
-	Thallus dull when fresh, margins mainly on the apex of the
	lobes with cilia up to $550 \mu m$ long; distal face of spores with
	(6) 8–10 (12) alveolae across the diameter, each 12–16 μm
	wide, wing 3.5–7.5 μm wide
	Note: According to Meinunger and Schröder (2007), additional characters for the identification of <i>R. crozalsii</i> are: (i) plants small, thallus lobes 3–6 ×
	0.4–1 mm, growing in mostly irregular rosettes; (ii) thallus margin
	distinctly purple; (iii) lobe apex very thick, turgid, almost cushion-like;
	(iv) epidermal cells pear-shaped; (v) spores often remaining light
	yellowish or light brown for a long period, but mature spores always very dark to nearly black, often all shades mixed.
15	Robust plants, thallus 1.8–3 mm wide; often with cilia;
15	spores 95–130 µm, brown, wing mostly papillose-crenulate;
	thallus not decaying after ripening of spores
	Riccia beyrichiana
	Note: doubtfully recorded from Hungary.
-	Plants medium-sized to small; spores mostly less than 100
	μm, wing smooth or notched; internal thallus tissue mostly
	soon decaying after spore ripening
16	Thallus (seen in transverse section) 3-5 times as wide as
	high; margins thin, sharp, curved downward; thallus lobes
	green or bluish green without a trace of purple; ventral
	scales colourless, quickly vanishing; cilia mostly lacking;
	spores 80–100 μm, deep reddish-brown, small alveolate on
	proximal face, with distinct wing; medium-sized plants
-	Thallus in transverse section 1–3 (4) times as wide as high;
	margins rounded or bulging, not decurved17

- - (ii) From *R. glauca*, which also has a rather wide thallus with a \pm flat groove, in section 3–5 times as wide as high, *R. bifurca* can be distinguished by being only 3 (4) times as wide as high in section and having raised, convex margins forming small tumid ridges (sharp, \pm deflexed margins in *R. glauca*). Spores of *R. glauca* are larger (80–100 μ m in diameter vs. 60–80 μ m in *R. bifurca*) and also differ in having usually well delimited alveolae on the proximal faces, and 6–8 (12) alveolae 6–12 μ m wide across the distal face. The spores of *R. bifurca* are light brown and translucent even when old, whereas the spores of *R. glauca* are darker red brown to brown, blackish when old, opaque to hardly translucent (but light brown translucent when just ripe).
 - (iii) R. bifurca var. subinermis: Thallus apex with a few cilia.
- - Notes: (i) According to Meinunger and Schröder (2007) the plants in the drawings under this name in Gradstein and van Melick (1996), Paton (1999), Damsholt (2002) and Siebel and During (2006) apparently represent *Riccia glauca* var. *subinermis* (= var. *ciliaris*).
 - (ii) Cilia, papillose in their upper part, 70–120 (200) μ m, are not infrequently found near the thallus margin (in *R. bifurca*, smooth cilia, 120–340 μ m, may be sometimes found).
- Spores 60-80 μm, wing mostly distinct to well-developed_19
 Distal face of spores with 10-12 alveolae across, proximal face with fine reticulate pattern; mature spores reddish brown; cilia few, but regularly present; thallus 1-3 times as
 - wide as high, margins mostly with reddish tinge, otherwise resembling *R. glauca.......Riccia glauca* var. *ciliaris* (rr)

- See the note under R. subbifurca.

6. KEYS TO SPECIES OF MOSSES

Kev	to	sections	and s	pecies	of S	phac	num

Branch leaves means leaves of spreading branches unless stated otherwise. References: Dierssen (1996), Hill in Smith (2004), Szurdoki (2000, 2003), Szurdoki and Nagy (2002), Szurdoki *et al.* (1999–2000, 2014), Laine *et al.* (2011, 2018), Hassel *et al.* (2018)

1	Hyalodermis of stems (mostly) and branches (always) with spiral fibrils, best seen on pendent branches; branch leaves large, concave, 1.5–2 times as long as wide (Subgens Sphagnum)
-	Hyalodermis of stems and branches without spiral fibrils; branch leaves smaller, less concave, usually more than twice as long as wide
2	Branch leaves 2–3 mm, stem leaves smaller, < 0.7 mm long; apex of branch leaves broadly truncate; upper branches often pointing upwards; chlorocysts completely enclosed between hyalocysts (like in <i>S. divinum</i>); hyalodermis of branches of uniform cells with 1 apical pore each (Subgen Rigida)
_	Stem leaves usually > 0.7 mm long, often longer than branch leaves (if stem leaves small, 0.7 mm or less, then branch leaves with acute or cuspidate apex); apex of branch leaves narrowly truncate; chlorocysts not completely enclosed between hyalocysts; hyalodermis of branches consisting of 2 types of cells, short poreless cells and long "retort cells" with apical pore
3	Chlorocysts of branch leaves exposed on ventral side in transverse section (or exposed on both sides, but more so on ventral side); branch leaves small, lanceolate; plants often with red colours (Subgen. Acutifolia Sect. Acutifolia)12
-	Chlorocysts of branch leaves with a wider exposure on the dorsal side or ± equally exposed on both sides in transverse section; branch leaves small or large; plants never with red colours, green, yellow-green, yellow or brown
4	Chlorocysts of branch leaves ± equally exposed on both sides in transverse section; hyalocysts of branch leaves mostly with numerous small ringed pores on dorsal side arranged in rows along commissures; spreading and

	pendent branches weakly differentiated, branches in capitulum often homomallous, horn-like; plants often with
	yellow and brown colours (Subgen. Subsecunda)8
	•
-	Chlorocysts of branch leaves more exposed on dorsal than
	on ventral side; pores on dorsal side of hyalocysts of branch
	leaves few or numerous, but not arranged in rows;
	spreading and pendent branches strongly differentiated,
_	branches in capitulum not homomallous, not horn-like5
5	Stem leaves with border of several rows of elongate cells;
	branch leaves undulate when dry, 3–7 times as long as wide;
	stem hyalodermis sometimes not clearly distinguishable
	from sclerodermis (Subgen. Cuspidata)
-	Stem leaves with few rows of elongate cells at leaf base only;
	branch leaves shorter, not undulate when dry, often with
	apices reflexed; stem hyalodermis sharply differentiated
	from sclerodermis (Subgen. Acutifolia Sect. Squarrosa)24
6 (1)	Plants usually purplish red or tinged with red; chlorocysts
	elliptical in transverse section, completely enclosed by
	hyalocysts, centrally placed
	(Sphagnum magellanicum auct. pp.) (rr)
-	Plants green, brown, not red; chlorocysts exposed on ventral
	side or if enclosed between hyalocysts then placement not
	central 7
7	Chlorocysts in transverse section narrowly triangular,
	exposed on ventral side
_	Chlorocysts in transverse section centered but very
	narrowly exposed on both sides, lenticular or spindle-
	shaped to narrowly triangular with broader side close to
	ventral surface, cell walls thickened
8 (3)	Hyalodermis of stem (1) 2–3-layered in transverse section
	9
_	Hyalodermis of stem unistratose, stem often ± dark
9	Hyalodermis of stem 1-2-layered; stem leaves to 2.5 mm
	long, broadly lingulate
_	Hyalodermis of stem 2–4-layered, stem leaves < 1mm long,
	triangular-lingulate
10	Stem leaves 0.4–0.6 (0.9) mm long; plants slender, yellowish
	to brownish green
_	Stem leaves at least 0.9 mm long

11	Stem leaves 0.9–1.5 mm long, triangular-lingulate, widest at
	leaf base, fibrillose in upper third; spreading branches not or only weakly homomallous
_	Stem leaves to 2.5 mm long, lingulate to spathulate, widest
_	above leaf base, sometimes fibrillose to base; branches often
	homomallous, horn-like
	Sphagnum auriculatum (Sphagnum denticulatum) (rr)
12 (4)	Stem leaves spathulate, broadly truncate, fringed at apex;
12 (1)	plants with distinct terminal bud; plants always green, never
	red
_	Stem leaves triangular to lingulate, not fringed at apex;
	plants red or green
13	Stem leaves expanded at apex, margins fringed from apex to
	lower half of leaf
-	Stem leaves not expanded, fringed only at apex
	Sphagnum girgensohnii (rr)
14	Stem leaves lingulate, apex broadly rounded, the sides ±
	parallel, flat, not incurved
-	Stem leaves triangular, with acute or obtuse apex, the sides
	tapering slightly at base, more strongly towards apex, flat or
	incurved towards apex17
15	Outer cells of stem hyalodermis with scattered pores, not
	more than 1 pore per cell; branch leaf hyalocysts with
	numerous pores on both sides; stem leaves broadly
	lingulate, slightly denticulate-fringed at rounded apex
	Sphagnum russowii (rr)
-	Outer cells of stem hyalodermis without pores; branch leaf
16	hyalocysts with numerous pores on dorsal side only16 Branch leaves in 5 distinct ranks, their hyalocysts towards
10	apex with many small ($ca 5 \mu m$) ringed pores on dorsal side;
	branch leaf apices pointing outward when dry; plants
	reddish violet
_	Branch leaves not in 5 ranks, their hyalocysts with larger,
	elliptical, less distinctly ringed pores on dorsal side; branch
	leaf apices not pointing outward when dry; plants reddish
	Sphagnum capillifolium (r)
17	Branch leaves distinctly 5-ranked in and below capitulum;
	fascicles with 4–5 branches in total, of which 2–3 are
	spreading; stem leaves triangular-acuminate, with wide
	zone of elongate cells at base: stem hyalodermis with

	scattered pores or pseudopores; sclerodermis green with
	red spots
-	Branch leaves not distinctly 5-ranked outside capitulum;
	fascicles with 3-4 branches in total, of which rarely more
	than 2 are spreading; stem hyalodermis without pores, but
	sometimes with pseudopores; sclerodermis reddish or
	brownish even in otherwise green plants18
18	Stem leaves 1.5–1.7 mm long, contracted into tubulose apex,
	at base with narrow to moderately widened zone of elongate
	cells, hyalocysts usually without fibrils, several times
	septate; plants lustrous when dry, violet reddish
-	Stem leaves 0.9-1.4 mm long, not contracted into tubulous
	apex, triangular to triangular-lingulate, often rounded at
	apex, at base with widened zone of elongate cells, hyalocysts
	usually with fibrils in apical part of stem leaf, ± septate, but
	rarely more than once; plants not lustrous when dry,
	reddish, often in compact tufts Sphagnum capillifolium (r)
19 (5)	Stem leaves to 1.6 mm long, at apex with a cleft of 1/3 of leaf
	length; branch leaves to 3 mm long; very robust, green
	plants
-	Stem leaves without apical cleft
20	Stem leaves triangular-lingulate to lingulate, rounded at
	apex 21
-	Stem leaves triangular, narrowed above, rounded or acute at
	apex 23
21	Branch leaf hyalocysts with numerous dorsal pores
	arranged in 1 or 2 rows, pores often indistinct and visible
	only after extensive staining; stem leaves > 1 mm long;
	robust plants
-	Branch leaf hyalocysts with few pores in the cell corners
	only22
22	Stem leaves 1 mm long or little longer, longer than wide;
	axes of branches and stems pale; robust plants
	Sphagnum flexuosum (rr)
-	Stem leaves mostly < 1 mm long, as long as wide; axes of
	branches and stems often pink; small to medium sized
	plants
23	Stem leaves 1–2 mm long, triangular, longer than wide;
	branch leaves > 4 times as long as wide: chlorocysts

_	
-	sides
	Stem leaves < 1 mm long, triangular, as long as wide, apex
	cuspidate; chlorocysts in transverse section widely exposed
	on dorsal side, narrowly exposed on ventral side
	Sphagnum fallax (r)
	Note: A useful way to distinguish <i>S. fallax</i> from <i>S. cuspidatum</i> in the field is to tear off a capitulum and inspect the pendent branches at its underside:
	these are in pairs, whitish and downwards pointing in <i>S. fallax</i> , whereas in
	<i>S. cuspidatum</i> they are greener and stick out sideways (Godfrey and Hill 2012).
24 (5)	Plants mostly bright green to greenish yellow; branch leaves
	with apices distinctly reflexed, $1.7-3.1 \times 1.0-1.8$ mm;
	hyalocysts elongate throughout the leaf, distinctly wider in
	basal part
-	Plants yellowish brown, rarely pure green; branch leaves
	usually imbricate, apices rearly reflexed, $1.0-2.3 \times 0.5-1.2$
	mm, hyalocysts shorter towards apex, of equal width
	throughout leaf
Kow	to the varieties of Andreaea rupestris
	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long;
	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long; plants to 2 cm high; papillae on dorsal side of leaves less than twice as high as wide
	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long; plants to 2 cm high; papillae on dorsal side of leaves less than twice as high as wide
	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long; plants to 2 cm high; papillae on dorsal side of leaves less than twice as high as wide
Refer	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long; plants to 2 cm high; papillae on dorsal side of leaves less than twice as high as wide
Refer Key For a	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long; plants to 2 cm high; papillae on dorsal side of leaves less than twice as high as wide
Refer Key For a	Leaves shortly and bluntly apiculate, 0.7–1.5 mm long; plants to 2 cm high; papillae on dorsal side of leaves less than twice as high as wide

-	Costa wide, filling upper part of leaf, leaves therefore opaque, not changed when dry, whithout border of narrow long cells, never transversely undulate; usually leaf base sheathing; capsule with campanulate, usually hairy calyptra
2	Leaves about 1 mm wide; lamina cells 10–16 (20) μm in diameter; leaves ± taut, not much transversely undulate; lamellae 5–6 (7) in number, 6–9 cells high; spores 12–14 μm
-	Leaves 1.5–2 mm wide; lamina cells 20–30 (50) μm in diameter; leaves distinctly transversely undulate; lamellae 4–5 (6) in number, to 5 cells high; spores 16–20 μm3
3	Autoicous (antheridia terminal on first year's growth, archegonia terminal on second year's growth); sporophytes mostly single, seta red, capsule red brown, inclined
- 1 (1)	Paroicous (antheridia terminal, archegonia outside perigonial leaves, stem continuing growth for 2 or more years, so that persistent setae appear lateral); often several sporophytes in a single perichaetium, seta yellowish (straw-coloured to reddish-yellow), capsule dark purple to reddish, narrow, almost erect
r (1)	5
5	Capsule with apophysis and stomata, 4–6-angled or terete7 Plants bluish-green, > 2 cm tall, often branched or with subterranean stolons; protonema not persistent; uppermost cells of lamellae papillose, wider than tall in cross section
- 6	Note: for the difference between this species and <i>Polytrichastrum alpinum</i> , see the note under that species. Plants dark green, < 2 cm tall, unbranched; protonema persistent; uppermost cells of costal lamellae smooth

-	Leaf margins serrate only at apex; capsule (without lid) to 2.3 mm long, ovoid to almost globose, 1–1.5 times as long as wide, exothecial cells finely papillose.
	Pogonatum nanum (r)
7	Capsule cylindrical, rounded; uppermost cells of costal
	lamellae papillose, taller than wide in cross section
	Note: Pogonatum urnigerum also has papillose end cells of lamellae, but
	these are wider than tall. Other differences between these two species which share also round, not angled capsules, are: (i) colour: dull green
	in <i>P. alpinum</i> , glaucous green in <i>Pogonatum urnigerum</i> ; (ii) length of
	sheathing part of leaf: 24–30% of total leaf length in <i>P. alpinum</i> , to 22%
	in <i>Pogonatum urnigerum</i> ; (iii) stomata lacking in capsules of <i>P. urnigerum</i> ,
	stomata present at base of urn in <i>P. alpinum</i> .
-	Capsule angled; uppermost cells of costal lamellae smooth.
	Polytrichum 8
8	Leaf margins sharply serrate in upper part of leaf, plane
	when moist 9
-	Leaf margins almost entire (except at tip), involute when
	moist 12
9	Uppermost cells of costal lamellae furrowed or flat in
	transverse section, at least in part
-	Uppermost cells of costal lamellae all rounded11
10	Uppermost cells of costal lamellae all grooved
-	Uppermost cells of costal lamellae variable in shape, some
	asymmetric, some grooved, some flat-topped (especially in
	median part of costa), some rounded (especially in marginal
	part of costa)
	Polytrichum perigoniale (P. commune var. perigoniale) (r)
11	Lamina in upper part of leaf consisting of 3–5 rows of cells;
	teeth 1-celled; lamina cells 10–15 µm wide; capsule with 4–6
	sharp edges; spores 12–16 μm <i>Polytrichum formosum</i> (c)
	Notes: (i) Fully developed leaves of P. formosum have a V-shaped ridge at
	their back formed by the prominent costa, whereas the dorsal side of the
	leaf is rounded in <i>P. commune</i> . (ii) The basal sheathing part of the leaf is dull in <i>P. formosum</i> , but shiny in <i>P. commune</i> , (iii) Young leaves of <i>P.</i>
	formosum may be indistiunguishable from <i>P. longisetum</i> .
_	Lamina in upper part of leaf consisting of 6–15 rows of cells;
	teeth formed of 1–2 cells; lamina cells 15–20 µm wide,
	capsule with 5–6 blunt edges; spores 18–26 µm
	Polytrichum longisetum (r)

12 (8)	Excurrent part of costa hyaline, 1/4–1/3 as long as leaf; lamellae crenulate with crenulae directed towards leaf apex
-	Excurrent part of costa brownish; crenulae of lamellae ± erect
13	Plants to 7 cm tall, leaves patent when moist, stems not or slightly tomentose; capsule <i>ca</i> 5 mm long; on dry soil
-	Plants to 20 cm tall, leaves erect-spreading when moist, stems densely tomentose with whitish rhizoids; capsule <i>ca</i> 3 mm long; in bogs
	liary key for sterile plants of <i>Polytrichum, Polytrichastrum</i> Pogonatum
1	Leaf margin inflexed over blade, partly covering lamellae, appearing entire (except at leaf tip)
	Polytrichum piliferum, P. juniperinum, P. strictum
-	Leaf margin sharply serrate, not inflexed 2
2	End cell of lamellae papillose
-	End cell of lamellae not papillose4
3	Sheathing part of leaf relatively short, to 22% of total leaf length, plants glaucous; end cell of lamellae wider than high
-	Sheathing part of leaf longer, 24–30% of total leaf length, plants dull green; end cell of lamellae higher than wide
4	End cell of lamellae grooved, at least in part of lamellae
_	End cell of lamellae rounded5
5	Plants small, to 2 cm high, with persistent protonema; costa
	not excurrent
_	Plants mostly much higher than 2 cm, robust; without
	persistent protonema; costa excurrent as apiculus
	Polytrichum formosum. P. longisetum

Key to species of Buxbaumia

References: Deme et al. (2020), Erzberger et al. (2018)

 Sporophyte reddish brown, glossy, capsule ovoid, strongly flattened, with a sharp edge; capsule epidermis not peeling off on dorsal side, or only to a mostly small extent at capsule mouth; peristome teeth uniseriate; stomata (at neck of capsule) deeply immersed (cryptopore); on soil.......

......Buxbaumia aphylla (w)

Key to species of *Timmia*

Reference: Mastracci (1993)

- Sheathing part of leaf orange or red brown; dorsal surface of costa in upper part of leaf serrate to obtusely dentate; costa of sheathing part of lamina without stereids; endostome cilia without appendages; dioicous, capsules rare......

Key to species of *Encalypta*

References: Nyholm (1998), Meinunger and Schröder (2007)

- 1 Plants with clusters of brownish filamentous gemmae in leaf axils; plants dioicous, rarely with sporophytes; urn spirally striate, spirally furrowed when dry; uppermost leaves obtuse or subacute; costa not excurrent......
 - Encalypta streptocarpa (w)

2	Peristome absent; calyptra smooth or erose at base; spores coarsely papillose; leaves bluntly pointed, costa ceasing below leaf apex (rarely shortly excurrent)
	Note: In <i>Encalypta vulgaris</i> (w) Note: In <i>Encalypta vulgaris</i> var. <i>apiculata</i> (rr) the costa is shortly excurrent or ending in a more or less long hairpoint.
-	Peristome present, single; costa excurrent as short apiculus or longer hair point
3	Leaf apex with short apiculus; calyptra lobed ("ciliate") at base, shiny, smooth; seta yellow; capsule smooth; spores with numerous radial plicae on the proximal surface and with 5–7 plicae on the distal surface, not papillose
	Encalypta ciliata (r)
-	Leaf apex mostly with long hair point; calyptra smooth or erose at base, papillose throughout; seta red; capsule with prominent longitudinal red-brown striae, when dry deeply furrowed; spores on distal surface with large hemispherical papillae
	nces: Erzberger (2002), Guerra <i>et al.</i> (2010)
1	Seta only 1–2 mm long, capsule immersed in leaves; capsule indehiscent, without lid, globose with short apiculus; plants < 5 mm high
	(Aphanorrhegma patens, Physcomitrella patens) (w)
-	Seta longer, capsule exserted, dehiscent
2	Calyptra persistent, inflated, contracted at base, with four angles; capsule emergent to exserted, not reached by upper leaves, ovoid to subglobose, as long as seta; spores 45–70
	μm
_	exserted; subglobose or not; spores less than 45 µm
3	Capsule furrowed when dry and empty; seta cygneous; peristome sigmoid, teeth fused apically in a disc; calyptra cucullate
-	Capsule smooth, not furrowed; seta not cygneous; peristome teeth not fused apically in a disc or peristome lacking or rudimentary; calyptra cucullate or mitrate3

3	Peristome present; capsule obovoid or pyriform, inclined; exothecial cells rectangular; calyptra cucullate4
_	Peristome lacking or rudimentary; capsule subglobose or
	pyriform, erect; exothecial cells rectangular or isodiametric;
_	calyptra cucullate or mitrate5
4	Leaf margins sharply serrate in upper half of leaf by
	projecting cells; marginal cells usually longer and narrower
	than adjacent cells; seta 7–11 mm long; capsule curved, <i>ca</i> 3 mm long, with <i>ca</i> 70 stomata; spores coarsely papillose
	Entosthodon muhlenbergii (Funaria muhlenbergii) (r)
_	Leaf margins entire to bluntly denticulate by slightly
	projecting cell ends; marginal cells similar to adjacent cells;
	seta 5–8 mm long; capsule curved, <i>ca</i> 2 mm long, with <i>ca</i> 30
	stomata; spores finely papillose
5	Calyptra of ripe capsule cucullate; lid without apiculus;
	exothecial cells isodiametric, incrassate
	Entosthodon fascicularis (r)
-	Calyptra of ripe, fully developed capsules mitrate, usually
	trilobed at base; lid with a short apiculus to rostrate;
	exothecial cells ± isodiametric, thin-walled or rectangular
_	and strongly incrassate
6	Capsule pear-shaped to club-shaped, with a long neck;
	exothecial cells long and narrow, nearly linear, incrassate;
	peristome present, but rudimentary, slightly protruding from the mouth of the urn and soon disappearing after
	dehiscence; leaves entire; spores coarsely papillose
_	Capsule semiglobose to pear-shaped, usually with a short,
	indistinct neck; exothecial cells lax, predominantly rounded
	quadrate to hexagonal; peristome absent; leaf margin ±
	entire to crenulate or distinctly denticulate by protruding
	cells longer and often narrower than adjacent lamina cells;
	spores densely echinate. <i>Physcomitrium</i> pp
7	Capsule pear-shaped, gradually narrowed into seta; plants
	(including seta) 5–10 mm tall; spores 24–35 μm; leaf margin
	distinctly denticulate by protruding narrow and long cells
	Physcomitrium pyriforme (w)
	Note: Young capsules, green and still covered by the calyptra, of this species have a rostrate lid (from a conical base gradually narrowed to a
	species have a rostrate nu (from a confical base gradually harrowed to a

- beak), whereas young capsules in the same state of *E. hungaricus* have a low conical, but not rostrate lid, through which the peristome is visible; in the process of ripening and drying, the lid of *E. hungaricus* becomes flat and shortly rostrate.
- 8 Leaf margin distinctly denticulate above; costa percurrent or excurrent; seta 2–5 mm long; spores 30–40 μm; plant (including sporophyte) 5–7 mm tall......

Key to species of *Flexitrichum*

Key to species of *Campylopus*

References: Erzberger and Németh (2014), Csiky *et al.* (2014, 2015) Note: transverse sections of the costa should be made at about 1/4 of leaf length above the base

1 Leaves piliferous due to hyaline excurrent costa (which may be reduced in shaded habitats), hair points reflexed when dry; dorsal surface of costa ribbed; ventral cells wide, empty, occupying *ca* 25–50% of total costa thickness, about as

	(cross section); seta cygneous before capsule maturation,
-	calyptra fringed at base
2	Costa without stereids in transverse section; vegetative propagation by caducous shoot tips with long, erecto-patent leaves; mostly small plants
	(stunted forms of) Campylopus pyriformis
3	Costa with stereids
	more often brown auricles; vegetative propagation by caducous branches, small-leaved at base, but with a long-
	leaved apex, at stem tips
-	Ventral cells of at least equal size as and not more in number than median cells in costa cross sections; alar cells variable, vegetative propagation mostly by caducous leaves4
4	Leaves widest at or almost at base; ventral cells usually smaller than guide cells (but larger in slender plants: Smith (2004: Fig. 65,7)), occupying <i>ca</i> 32–54% of total costa thickness; alar cells usually undifferentiated, hyaline or pale red, rarely forming distinct auricles; leaf apex canaliculate.
	denticulate; vegetative propagation by broadly oval brood leaves at stem tips
-	Leaves widest 1/8 to 1/4 distance from base, tapering downwards; ventral cells larger than median cells and about equal in number, occupying between 1/3 and 2/3 of total costa thickness, dorsal groups of stereids distinct; basal cells hyaline, not forming auricles, conspicuously shining whitish in dry and moist plants; vegetative propagation by bundles of caducous minute, linear leaves in axils of normal leaves at shoot apex
	onoccupes

Key to species of *Leucobryum*

Reference: Zündorf (1988)

- Most cells in the middle of the leaf base > 30 μm wide; hyaline marginal cells (=lamina cells) mostly in 5–8 rows, at extreme margin narrow and elongate cells with an abrupt transition; capsule distinctly strumose; pores of hyalocysts (transverse section!) 15–20 μm wide, many large; basal expanded part of leaf usually longer than narrowly triangular tubular apical part, transition usually ± gradual; cushions higher, more swelling than in the following species

 Leucobryum glaucum (w)

Key to species of Dicranella

- 3 Plants almost always sterile, always with irregularly shaped, brownish rhizoidal tubers; leaves erect-spreading, shortly acuminate from broadly lanceolate base, margin plane, denticulate at apex; lamina cells 9–14 μm wide; plants to 5

	mm tall
-	Plants often with sporophytes, different combination of
_	characters 4
4	Seta yellow; costa filling 1/3 of leaf width at base or more5 Note: compare also <i>D. howei</i> with costa indistinctly delimited at leaf base (9).
-	Seta red; costa filling up to 1/5 of leaf base6
5	Capsule strumose, brown; leaves suberect to secund, entire except at apex; basal cells 70–115 µm long
	Dicranella cerviculata (rr)
-	Capsule not strumose, reddish brown; leaves falcate, dentate; basal cells 30–50 µm long
_	Dicranella heteromalla (c)
6	Perichaetial leaves differentiated from normal leaves, suddenly contracted into acumen from sheathing base; leaves erect or secund; leaf margins almost entire; capsule striate, lid longly rostrate; rhizoidal tubers often present, dark brown; plants 5–20 mm tall <i>Dicranella subulata</i> (n.s.)
-	Perichaetial leaves not differentiated from lower leaves, not
	sheathing, gradually narrowed into acumen7
7	Capsule erect, symmetric; leaves mostly homomallous, reddish; stems and setae reddish; leaf margin plane, very weakly denticulate in upper part of leaf
	Dicranella rufescens (r)
	Note: When sterile, this species can be recognized by its mostly reddish stems and ± homomallous leaves, which appear cross-striated, since the chloroplasts tend to crowd in the cell corners and the cells are arranged in transverse rows. It often grows together with <i>Pohlia lescuriana</i> , from which it can be distinguished by the marginal cells which are wider than median leaf cells (narrower in <i>P. lescuriana</i>).
-	Capsule curved, asymmetric; leaves erect or weakly homomallous; margins recurved or plane, entire or weakly denticulate only near the tip, plants not reddish8
8	Leaf margins recurved; leaves erect when dry, stiff;
	longitudinal walls of exothecial cells more heavily thickened than transverse walls; plants in tufts to 1 cm tall
	Dicranella varia (w)
_	Leaf margins ± plane9
9	Lamina cells irregularly bistratose; costa strong, filling 1/3
	leaf base, indistinctly differentiated from lamina; longitudinal walls of exothecial cells hardly wider than

Lamina cells twice as long as wide; plants 1–3 mm long with 2 3-5 pairs of leaves; leaves narrowly lanceolate, tapering to apex from below middle; terricolous..... *Fissidens curvatus* (Fissidens algarvicus) (n.s.) Lamina cells as long as wide; leaves tapering only at apex....3 3 Limbidium mostly confined to sheathing part of perichaetial leaves, sometimes variously reduced on cauline leaves; apex rounded to obtuse, cells below apex often in concentric rows; margin slightly crenulate by protruding cell ends; plants growing on limestone or other base-rich rock in or Limbidium well developed on all laminae and all leaves except sometimes the lowermost.......4 4 Leaf cells conspicuously small, usually not longer than 8 µm and not wider than 6 µm; leaves oblong to ovate-lanceolate, > 2.5 times as long as wide, median lamina cells distinctly protuberant on both sides, higher than wide (transverse Limbidium confluent with excurrent or percurrent costa; 5 antheridia often in bud-like perigonia or naked in leaf axils.... Limbidium not confluent with costa; antheridia not axillary.....8 6 Antheridia terminal in dwarf male plants: capsule Antheridia axillary, not in dwarf male plants; capsule erect

137

	or inclined
7	Antheridia nearly always naked in leaf axils; plants mostly
	growing on periodically inundated, silt-covered tree trunks
	or roots (rarely also rocks or stones; in other European
	countries often on silt-covered stones of artificial riverbank
	enforcements along great rivers)
	Fissidens gymnandrus (w)
-	Antheridia in axillary bud-like perigonia, rarely naked
	plants growing on moist loamy soil usually away from water
	Fissidens bryoides (w)
8	Limbidium bi- to pluristratose on vaginant laminae
	sheathing part often with intramarginal limbidium; costa
	ceasing below leaf apex; median lamina cells (10)12–18(20)
	μm long; plants 5–30 mm long; mostly on calcareous or
	base-rich rocks near or in water
	(incl. Fissidens crassipes subsp. warnstorfii) (w)
-	Limbidium 1–2-stratose on vaginant laminae; plants often <
	5 mm long, terrestrial or aquatic9
9	Terricolous plants10
-	Plants growing on moist siliceous or calcareous rock or in
	streams 11
10	Capsules symmetric, erect
	Fissidens viridulus (incl. F. bambergeri) (w)
-	Capsules asymmetric, inclined
11	Perichaetial leaves 4–6 times as long as wide, lanceolate
	rather broad, suddenly narrowed to a short and wide
	somewhat obtuse or ± acute apex; plants hydrophilous
	growing on siliceous rock often close to flowing water or
	inundated Fissidens pusillus (w)
-	Perichaetial leaves to 6-7 (9) times as long as wide
	narrower, more gradually narrowed to slightly cuspidate
	point; plants growing on moist to wet calcareous rock
49(1)	Fissidens gracilifolius (w)
12 (1)	Mature plants small, 1–4 (6) mm long
- 12	Mature plants > 1 cm long
13	Leaves in 2–4 pairs, plants procumbent; margin at aper
	finely crenulate, margin on sheathing part of perichaetia
	leaves often coarsely dentate; dorsal lamina not extending to
	leaf base; terricolous, nearly always with capsules
	Fissidens exilis (w)

-	Leaves in 4–5 (10) pairs, increasing in length towards stem apex, plants erect; margin at apex entire or indistinctly crenulate by protruding cells, sheathing part of leaves entire; dorsal lamina reaching leaf base; plants growing on limestone or other base-rich rock and stones in or near water
14	Leaf margin dentate with large and smaller teeth
	alternating, particularly near apex; several rows of cells near margin forming pale band15
_	Leaf margin entire to crenulate, sometimes with 2–3 teeth
	near apex; leaf margin without pale band or just one marginal cell row paler; costa excurrent as a short apiculus; upper lamina cells $610~\mu m$ long, distinctly bulging to
	mamillose; perichaetia at the base of shoots
15	Fissidens taxifolius (c)
15	Lamina cells (10) 12–20 μ m wide, lamina unistratose, \pm translucent; plants 2–10 cm tall <i>Fissidens adianthoides</i> (r)
-	Lamina cells (5) 6–12 μm wide; lamina irregularly bistratose, opaque; plants 1–3 cm long. <i>Fissidens dubius</i> . 16
16	Costa ending shortly below leaf apex in uppermost leaves Fissidens dubius var. dubius (w)
_	Costa shortly excurrent as mucro in uppermost leaves
	Fissidens dubius var. mucronatus (r)
	to species of Dicranaceae ences: Erzberger (1999), Hedenäs and Bisang (2004)
1	Costa broad, 1/3–4/5 leaf width at base or more2
-	Costa narrower, < 1/3 leaf width4
2	Costa in transverse section with large hyaline cells
	(hyalocysts) present on both the dorsal and ventral sides;
	costa filling 1/2–2/3 leaf width, ridged on dorsal surface; leaf apex denticulate; lamina at leaf base consisting of <i>ca</i> 13
	cell rows; leaves hamate, plants in tufts to 4 cm tall
-	Costa in transverse section with a ventral and a dorsal layer
3	of stereids, separated by the guide cells
3	straight and erect when moist, upper leaf portions very narrow and tips frequently broken (touch cushion with

	moistened finger and inspect finger for broken tips); capsules straight and erect or nearly so4
-	Leaves in upper part not at the same time fragile, straight
4	and erect, upper leaf portions not markedly narrow5 Costa in basal portion lacking stereid bands, with up to 1–2
4	costa in basal portion lacking stered bands, with up to 1–2 cell layers above and below guide cells (transverse section);
	basal lamina cells narrowly rectangular to linear (35–120
	μm, most cells > 40 μm long), eporose; alar cells unistratose;
	dark green glossy dense tufts
-	with up to 2–3 (4) layers of cells above and below guide
	cells; basal lamina cells quadrate to short-rectangular (16–
	$50 \mu m$, most cells < $30 \mu m$ long), not or hardly porose, alar
	cells mostly bistratose, at least partially, rarely entirely
	unistratose; lamina above partially bistratose; plants light green, to 4 cm tall, leaves slightly curled when dry
	Dicranum viride (r)
5	Cells in upper leaf mainly prosenchymatous, elongate and
	porose; usually robust plants
-	Cells in upper leaf mainly parenchymatous, quadrate or rectangular, rarely elongate-rectangular, with or without
	pores8
6	Costa above with 2 dorsal lamellae or ridges; leaves ±
	transversely undulate
-	Costa above with 4 dorsal lamellae; leaves transversely undulate or not; variable species <i>Dicranum scoparium</i> (c)
7	Leaves at stem apex erect, further down spreading; leaf
	lamina strongly transversely undulate; costal lamellae tall;
	margin in upper part spinosely denticulate or dentate;
	margins recurved at leaf base; plants to > 10 cm high,
_	densely whitish tomentose
	clearly different; leaf lamina ± transversely undulate (at
	least in some leaves); costal lamellae low or weakly
	developed; margin obtusely to sharply denticulate; margins
	flat at leaf base; plants to 10 cm high
8	Upper leaf lamina in at least some leaves rugose, or ±
-	transversely undulate; scattered cells of lamina and costa on
	upper dorsal surface of leaf with conical mamillae; plants in

	loose tufts, to 6 cm high; leaves curled when dry, appressed
	to the stem in basal portion and suddenly narrowed to short
	acumen
_	Leaves smooth, neither rugose nor undulate9
9	Small plants (rarely above 3 cm high, leaves up to 4 mm long
	and up to 0.5 mm wide); dry leaves strongly crisped; upper
	lamina cells mamillose, normally unistratose; costa
	occupying 1/5 leaf base; leaf margins denticulate from apex
	down to mid-leaf
	Notes: (i) <i>D. montanum</i> can be distinguished from <i>Dicranoweisia cirrata</i>
	occurring in the same habitat (tree bark, siliceous rock) in the field by the
	size distribution of leaves: whereas in <i>D. cirrata</i> all leaves are of
	approximately the same size, in <i>D. montanum</i> leaves occur in a wider size range within one cushion, larger and in particular many smaller leaves
	mixed. (ii) When growing on rock, <i>D. montanum</i> could also be confused
	with species of <i>Cynodontium</i> , which are somewhat similar in habit (leaves
	crisped when dry), but distinguished by bistratose leaf margins (except
	the rare <i>C. tenellum</i>).
- 10	Different combination of characters
10	Small plants that almost invariably have numerous easily
	detached erect small-leaved flagellae from upper leaf axils;
	alar cells unistratose; costa occupying 1/3 leaf base; leaf
	margins denticulate only at apices <i>Dicranum flagellare</i> (r) Note: For the difference to <i>Campylopus flexuosus</i> , see the note under that
	species.
_	Plant size variable; if flagellae present, then never several on
	each plant; alar cells at least partially bistratose11
11	Upper leaf lamina bistratose in large portions; costa very
	broad, 1/3 of basal leaf width; leaves crispate when dry; leaf
	margin denticulate in upper part of leaf; plants dark green,
	to 5 cm tall
_	Upper leaf lamina unistratose; costa narrower, filling < 1/4
	leaf width; upper leaf tubular (cross section with circular
	outline); leaves strongly curled when dry; leaf margin near
	apex irregularly and coarsely denticulate to dentate; robust,
	usually tomentose plants in tufts to 6 cm tall, leaves erect-
	spreading, 5–7 mm long
	opicading, o / min long rel unum muchienbeckii (11.5.)

Key to species of Rhabdoweisiaceae

1	Lamina cells smooth 2
-	Lamina cells mamillose; upper leaf margin ± bistratose,
	mamillae of marginal cells forming double teeth
2	Capsules striate when dry and empty, less than 3 times as
	long as wide; leaf margin denticulate in upper part of leaf,
	sometimes only very slightly so
_	Capsules smooth, more than 3 times as long as wide; leaf
	margin entire; often with elliptical to cylindrical gemmae on
	basal part of lamina
	Note: For the difference between this species and Dicranum montanum
	see the note under the latter species.
3	Leaves not with parallel margins, tapering from lower third;
	peristome teeth divided to base; annulus formed of small
	cells, persistent, not separating. <i>Cynodontium tenellum</i> (rr)
-	Leaves with ± parallel margins, tapering only in upper 1/3
	to 1/5; peristome teeth undivided; annulus not
	differentiated 4
4	Peristome teeth from broad base suddenly narrowed to a
	filiform, fragile tip; leaves almost entire or weakly
	denticulate; lamina mostly 3–4 cell rows wide on either side
	of the costa at 220 μm below apex; upper lamina cells 7–12
	µm wide
-	Peristome teeth gradually narrowed, linear-lanceolate, not
	fugacious; leaves mostly distinctly denticulate; lamina
	mostly 5–6 cell rows wide on either side of the costa at 220
	μm below apex; upper lamina cells 9–12 (14) μm wide
E(1)	Peristome teeth undivided; perigonia on short stalks below
5 (1)	perichaetium; capsule short, < 3 times as long as wide
_	Peristome teeth divided halfway or below; perigonia sessile;
_	capsules > 3 times as long as wide
6	Capsules erect, symmetric, not strumose
J	Cynodontium polycarpon (w)
_	Capsules curved, asymmetric, strumose
	Cynodontium strumiferum (rr)
	Sylloudillian (11)

Key to species of Ditrichaceae s.l. incl. Saelania and Flexitrichum

1	Plants bluish green with waxy surface, 2–3 cm tall; leaves lanceolate, the upper longer; costa percurrent; lamina cells quadrate or shortly rectangular <i>Saelania glaucescens</i> (rr)
_	Plants not glaucous, without waxy surface
2	Upper leaves squarrose, from sheathing base ± abruptly tapering to long, flexuose subula consisting of costa, cells prorate, therefore subula denticulate all around, not just at
	margins. <i>Trichodon cylindricus</i> (Ditrichum cylindricum) (w)
-	Upper leaves not squarrose, not abruptly narrowed to subula
3	Capsules indehiscent, enclosed by the leaves; seta hardly a few millimeters long4
-	Capsules with dehiscent lid, exserted above the leaves, seta longer than capsule, or capsules lacking6
4	Perichaetial leaves slightly differentiated from stem leaves, lanceolate; costa ending below leaf apex; plants without comal tuft, 2–4 mm tall; sometimes with yellowish-orange
-	rhizoidal gemmae
5	Antheridia 1–3, enclosed in buds in leaf axils; lamina unistratose at shoulder; costa in basal part clearly delimited from lamina
-	Antheridia single, naked in axils of comal leaves; lamina irregularly bistratose at shoulder; costa weakly delimited from lamina in basal part of leaf
	Pleuridium acuminatum (w)
6 (3)	Upper lamina cells quadrate; capsule ribbed, inclined7
-	Upper lamina cells elongate; capsule smooth, erect. Ditrichum s.l. 8
7	Peristome teeth with pale translucent border; costa percurrent, rarely excurrent; capsule strumose, inclined; leaf shape very variable, leaf apex usually with some blunt teeth.
	Ceratodon purpureus (cc)

-	Peristome teeth unbordered; costa longly excurrent; capsule ovoid, scarcely inclined, not or indistinctly strumose
8	Plants 1-8 cm tall, densely tomentose; leaves with long
	subula; upper lamina cells unistratose, predominantly oval
	to rounded, cells at shoulder irregularly quadrate, triangular or short rectangular; on calcareous soil. <i>Flexitrichum</i> 9
_	Plants to 1.5 cm tall, not tomentose; upper lamina cells
	quadrate to rectangular, more regular; leaves not subulate
	or if subulate, then upper lamina partially 2-stratose; on
9	acidic soil. <i>Ditrichum</i>
9	transition to lamina continuous, cross section forming a
	smooth ring; basal paracostal cells narrow, elongate,
	distinctly porose to nodulose; plants 4-11 cm tall, leaves 4-
	11 mm long Flexitrichum gracile (Ditrichum
_	<i>crispatissimum</i> , <i>D. flexicaule</i> var. <i>sterilis</i> , <i>D. gracile</i>) (rr) Costa in transverse section projecting dorsally, transition to
	lamina discontinuous; often pseudopapillose with projecting
	cross walls ('Pfeilerpapillen') seen in cross section; basal
	paracostal cells wider, rectangular, not or only indistinctly
	porose; plants 1–5 cm tall, leaves 1–3.5 mm long
10	Leaves erect to ± appressed to stem, in 3 ranks, 2–4 times as
10	long as wide, margins entire, recurved on one or both sides;
	capsule and seta yellowish brown, peristome yellowish
	Ditrichum lineare (rr)
- 11	Leaves > 4 times as long as wide
	(transverse section!); leaves spreading, with somewhat
	channelled acumen, weakly denticulate at margins; capsule
	and seta reddish, peristome reddish; sterile plants with
	yellowish brown rhizoidal gemmae 100–150 μm in diameter Ditrichum pusillum (r)
_	Leaf margin plane; leaves from ovate base narrowed to long,
	canaliculate subula 12
12	Seta red at least in lower part; leaves homomallous; leaf
	apex entire or with few teeth; leaf margin partially
	bistratose in subulate part of leaf (cross section!); rhizoids brown; rhizoidal gemmae dark red-brown, consisting of 5–7

Key to species of Acaulon

Key to species of Aloina

- 1 Basal marginal leaf cells elongate, thin-walled, forming distinct hyaline border of 2–3 (4) cell rows, which differ conspicuously from adjacent leaf cells; annulus of inflated cells, separating; peristome teeth twisted 1–3 times.....2

- Leaves 4–6 times as long as wide; costa in cross section with
 3–6 (8) layers of dorsal stereids; spores 11–15 (22) μm in

	diameter; lid half as long as the capsule; dioicous	
3	Leaf apex apiculate or mucronate, acute or obtuse, cucullate or not; spores 14–25 µm in diameter; basal membrane of peristome 1–2 cells high, not projecting above mouth of capsule; peristome teeth united in pairs at base, erect or slightly twisted; capsule slightly curved	
-	Leaf apex neither apiculate nor mucronate, rounded, cucullate; spores (12) 17.5–24 (27.5) µm in diameter; basal membrane of the peristome to 5 cells high, projecting above mouth of capsule; peristome teeth not united in pairs at base, usually distinctly twisted	
Key to species of Cinclidotus including Dialytrichia		
1	Leaf cells strongly papillose, leaves opaque; dorsal surface of costa not covered by epidermis, stereid cells therefore exposed; leaf margin recurved below; peristome segments free	
-	Leaf cells smooth or weakly papillose, translucent; dorsal surface of costa covered by quadrate to short rectangular cells 20 (100) x 10–15 μ m in upper 2/3 of leaf; leaf margin plane; peristome segments linked and often with crossbars, therefore lattice-like at least towards the base2	
2	Leaves widest at middle, elliptical, to 3 times as long as wide; costa $100-140~\mu m$ wide at leaf base, $ca~1/10~of$ maximal leaf width; leaf margins moderately thickened, border internally uniform; lamina cells $8-10~\mu m$; leaves appressed when dry, spreading when wet; setae $3-6~mm$ long, terminal	
-	Leaves widest shortly above insertion, gradually narrowed towards apex or with nearly parallel margins in lower third, mostly > 4 times as long as wide; costa $100-200~\mu m$ wide at leaf base, $ca~1/10$ of maximal leaf width or wider, $1/6-1/5$; leaf margin moderately to strongly thickened, sometimes with internal cells differentiated, lamina cells $8-15~\mu m$; leaves slightly or strongly curled when dry; setae very short, sporophytes lateral, immersed or sporophytes very rare	

(found only once)......3

3 Leaf border 2-5 layers thick, with differentiated internal stereids (long, thick-walled narrow cells with oblique apical walls); leaf cells 7.5-10 (12.5) μm; costa 100-140 μm wide at leaf base, ca 1/10 of maximal leaf width, rarely excurrent as a mucro; leaves strongly flexuose when dry; setae very short, sporophytes lateral, immersed Cinclidatus fontinaloides (w) Leaf border usually 2 layers thick, without differentiated internal cells (in cross section all cells of equal thickness): leaf cells 12-15 (19) um; costa 150-200 um wide at leaf base, often excurrent as mucro; leaves slightly flexuose when dry; sporophytes usually absent (found only once)...... Cinclidotus danubicus (rr) **Key to species of** *Crossidium* Leaf margins plane to incurved, hyaline; upper lamina cells 1 and end cells of filaments heavily incrassate; leaves with Leaf margins recurved, not hyaline; upper lamina cells and end cells of filaments thin-walled; hair points of variable length 2 Terminal cells of filaments smooth or almost so; peristome 2 350 μm long; cells at capsule mouth 5–10 μm wide..... **Crossidium laxefilamentosum** (rr) Terminal cells of filaments papillose; peristome teeth long, > 400 μm to 800 μm; cells at capsule mouth 9–15 μm wide...... Crossidium crassinervium (rr) **Key to species of** *Didymodon* This key includes 2 taxa of doubtful status in Hungary: *D. austriacus* and *D. validus*. For a key to infraspecific taxa of D. tophaceus, see below. References: Kučera (2000), Jiménez (2006), Müller (2017), Kučera et al. (2018) 1 Leaf apex not fragile......2 Leaf margins in upper part distinctly erose, irregularly 2 notched; plants with rhizoidal tubers..... **Didymodon tophaceus** subsp. **erosus** (rr)

-	Leaf margins in upper part entire or papillose-crenulate, not erose or notched; plants with or without rhizoidal tubers3
3	Ventral surface of costa with elongate cells above mid-leaf,
-	narrower than adjacent lamina cells (near apex sometimes
	isodiametric)4
_	Ventral surface of costa with quadrate, rarely short
	rectangular cells (cells of lamina extending over costa)7
	Note: See also <i>Didymodon tophaceus</i> subsp. <i>sicculus</i> with short rectangular
4	ventral costa surface cells.
4	Leaves strongly reflexed when moist, often distinctly 3-
	ranked, to 2 mm long; stem in transverse section with weak
	central strand and thick-walled parenchyma; plants in lax brown-green tufts, to 2.5 cm tall (or taller in wet habitats)
	Didymodon ferrugineus (r)
_	Leaves erect-spreading to recurved when moist, not
_	arranged in distinct rows; stem in transverse section with
	mostly thick central strand and thin-walled parenchyma5
5	Leaves lingulate to ovate with obtuse to subacute apex;
3	costa usually ending below leaf apex; plants to 1 cm high, in
	wet places to 6 cm, olive- or brown-green
	Note: For a key to infraspecific taxa of <i>D. tophaceus</i> see below.
_	Leaves acute, lanceolate, recurved when moist; costa ending
	in leaf apex or shortly excurrent6
6	Peristome spirally twisted, mostly more than 800 µm high;
	leaf margins mostly recurved to broadly recurved at 2/3-
	4/5 of leaf length (less in dry habitats); archegonia mostly
	$400500~\mu\text{m}$ long; small to robust plants, leaves 0.8 to more
	than 3.5 mm long
-	Peristome straight or nearly so, mostly < 500 μm high; leaf
	margins mostly not or only weakly recurved in upper half of
	leaf; archegonia mostly 700–800 µm long; robust plants,
5 (2)	leaves mostly > 1.7 mm long
7 (3)	Costa in transverse section (in lower third of leaf in well
	developed plants) without ventral stereids, dorsal stereids
	mostly in 2 or more layers; guide cells mostly in 2 layers8 Costa in transverse section with ventral and dorsal stereids,
-	guide cells usually in a single layer (if in 2 layers then
	ventral stereids well developed)12
	venu ai stereius wen uevelopeu J12

0	or more, narrower than adjacent lamina cells9
_	Dorsal surface cells of costa isodiametric at least in upper
	half of leaf, equally wide as lamina cells10
9	Basal lamina cells hyaline, sharply differentiated from upper
	lamina cells; lamina cells mostly strongly papillose and
	mamillose, papillae large, covering \pm all of cells; leaves (1.3)
	1.7–2.6 (3.8) mm long; costa (45) 60–90 (120) μm wide; leaf
	margins nearly always recurved; axillary gemmae mostly present
	Basal lamina cells coloured, but mostly lighter than upper
_	lamina cells, indistinctly differentiated; upper lamina cells
	densely papillose, papillae small, several per cell, thus
	obscuring cell areolation; axillary gemmae lacking
	Didymodon insulanus (w)
10	Lamina cells (or at least isodiametric cells on ventral side of
	costa) densely papillose, papillae small, several per cell, thus
	obscuring cell areolation, lamina opaque; leaf apex often
	with one or three pellucid cells11
-	Lamina cells smooth, if papillose than only 1 papilla per cell;
	lamina translucent and areolation clearly visible12
11	Leaves mostly longly linear-lanceolate (margins nearly
	parallel in upper half of leaf); margins recurved only to ±
	1/2 of leaf length; lamina cells nearly always thin-walled,
	densely papillose, opaque
-	Leaves mostly from ovate base triangular-lanceolate
	(margins towards apex forming an angle); margins recurved
	at 2/3 or 4/5 of leaf length; lamina cells sometimes incrassate and weakly papillose (papillae often only visible
	in section)
12 (7, 10)	Leaf margin unistratose
_	Leaf margin bistratose; leaves lanceolate with thick,
	subacute apex formed from costa and thickened leaf
	margins; upper lamina cells smooth to weakly papillose;
	plants to 2 cm tall (usually shorter), usually with nearly
	spherical but somewhat irregularly-shaped axillary gemmae
	Didymodon rigidulus (w)
13	Plants with gemmae on modified rhizoids in leaf axils14
-	Plants without axillary gemmae

14	Leaf margins spirally revolute in well developed plants; costa very broad and strong, 60–120 μm wide; axillary gemmae regularly spherical; plants in brown-green cushions to 5 cm high
-	Leaf margins recurved, not revolute; costa narrower; axillary gemmae of irregular shape, with protuberant cells
15	Lamina cells at mid-leaf and above small, 6–8 µm wide; gemmae regularly present
-	Lamina cells wider, <i>ca</i> 12 µm wide; leaves shorter, margin less strongly recurved; gemmae sometimes missing
16	Ventral stereids of costa in one layer; leaves (0.8) 1.0–1.4 (2.1) mm long; costa (30) 45–65 (80) µm wide; gemmae very rarely present
-	Ventral stereids of costa mostly in two layers; leaves (1.3) 1.8–2.7 (4) mm long; costa (40) 60–85 (120) µm wide
17 (13)	Leaves ovate-triangular (1.6–2.8 times as long as wide); costa mostly ending below or in leaf apex; peristome very short, mostly < 150 (290) μm
-	Leaves longer triangular (2–5 times as long as wide); costa excurrent, with excurrent part $1/6$ – $1/8$ leaf length; perichaetial leaves with acute apex and excurrent costa; peristome > 300 μ m
18	K+ red; plants 0.2–2 cm tall; lamina cells smooth; perichaetial leaves with rounded apex and costa ending below apex; leaf margins recurved nearly to apex, leaves therefore not curled when dry but straight. **Didymodon luridus** Didymodon luridus**
-	K+ greenish-yellowish; plants 0.2–0.4 (0.7) cm tall; lamina cells strongly papillose
19	Ventral stereids of costa in one layer; leaves (0.8) 1.0–1.4 (2.1) mm long; costa (30) 45–65 (80) µm wide
-	Ventral stereids of costa mostly in two layers; leaves (1.3) 1.8–2.7 (4) mm long; costa (40) 60–85 (120) µm wide

Key to infraspecific taxa of Didymodon tophaceus

From Kučera et al. (2018)

The key works for well-developed plants growing in non-extreme habitats in most, certainly not all cases. In particular, large hygrophytic plants of subsp. *erosus* might key out to subsp. *tophaceus*, hygrophytic morphs of subsp. *sicculus* might key out to subsp. *erosus*, while weak morphs of subsp. *tophaceus* might key out to subsp. *sicculus*. Sporophytic characters have only been observed in single cases for subsp. *sicculus* and subsp. *erosus*, respectively. Hence, barcoding using one of the molecular markers described in Kučera *et al.* (2018) is advisable in ambiguous cases.

-	Ventral superficial costa cells on all leaves elongate (> 3:1), costa on leaves of well-developed plants often > 100 μ m wide, leaf base markedly and widely decurrent; rhizoidal gemmae not knownsubsp. <i>tophaceus</i> 2 Ventral superficial costa cells on most leaves short-rectangular to irregularly quadrate (< 1.5:1), costa on leaves of well-developed plants mostly < 70 μ m wide, leaf base shortly, not conspicuously decurrent; rhizoidal gemmae often present (sufficient amount of substrate to be checked!)
2	Costa ending below apexvar. <i>tophaceus</i> (w)
-	Costa excurrentvar. anatinus
3	Leaves ovate, ovate-lanceolate to ovate-lingulate, to 1.45 mm long, costa < 85 µm wide; peristome present
-	Leaves lanceolate to lingulate, to 1.9 mm long, costa < 100 μm wide; peristome absent (observation on plants from Ecuador)subsp. <i>erosus</i> (rr)

Key to species of Microbryum

1	Capsule immersed; plants only 1 mm tall, brownish; costa
_	excurrent as a brownish cusp; leaf margins crenulate at
	apex; spores 27–30 μm
	Microbryum floerkeanum (Phascum floerkeanum) (r)
-	Capsule emergent or exserted 2
2	Capsule exserted to side of leaves, seta arcuate; leaves to 1
	mm long; plants conspicuously reddish brown
-	Capsule erect, exserted, seta straight; leaves longer

3 Peristome teeth present, well developed; lid bluntly conical or mamillate; spores raspberry-like, with coarse warts but otherwise smooth walls; calyptra distinctly papillose in upper part; upper lamina cells densely and strongly papillose.....

Peristome teeth absent or rudimentary; lid bluntly conical. 4
 Capsule short, empty capsule wide-mouthed, widest at mouth, not (or only slightly) longer than wide; peristome seemingly absent (reduced to a basal membrane not protruding above the capsule mouth); lid bluntly conical, with a short (erect) mamilla; spores echinate; leaf margins distinctly recurved; upper lamina cells distinctly papillose......

Note: M. davallianum shares with M. muticum the bluntly conical capsule lid, but differs in capsules not contracted at mouth (widest at mouth), strongly widened at mouth when dry and empty, and the tip of the lid appearing more like a beak when capsules are dry, whereas in M. muticum the capsules are widest some distance below the mouth, in empty capsules the mouth remains narrow, and the lid of dry capsules takes up a more hemispherical shape upon drying, without differentiated mamilla or beak. The spores of both species can be echinate, but the spines are shorter in M. muticum, and spore ornamentation is sometimes a mixture of spines and warts. Spores of M. muticum are normally warty with hemispherical papillae 2.5 μ m high and placed rather distantly (4 μ m apart), whereas spores of M. davallianum have spines up to 5 μ m long, also on average 4 μ m distant from each other. Spores of M. davallianum are often larger, 22–42 μ m, often about 35 μ m, spores of M. muticum are 16–32 μ m, often about 25 μ m (Erzberger, unpublished).

(Pottia mutica, P. starckeana var. brachyoda, Microbryum starckeanum var. brachyodus) (w)

Note: For the difference between this species and *M. davallianum*, see the note under that species. Capsules of *M. muticum* have longitudinal stripes caused by large spherical cells between the capsule wall and the spore sac that are arranged in longitudinal rows, best seen in fresh capsules just

before dehiscence, but detectable also in dry specimens and older capsules. Capsules of *M. davallianum* seem to lack these stripes (Limpricht 1890: 529).

Key to Pottia s.l. (including Hennediella heimii, Microbryum davallianum, M. muticum, M. starckeanum, Tortula caucasica, T. lindbergii, T. protobryoides, T. truncata, but without species formerly in Phascum: Microbryum curvicollum, M. floerkeanum)

1	Capsule indehiscent; peristome teeth differentiated but united with the indehiscent lid; costa excurrent; capsule elongate-ovoid, reddish brown like the seta; plants to 10 mm tall, brownish green
_	Capsule with dehiscent lid
2	Peristome teeth absent or rudimentary
_	Peristome teeth present, well developed
3	Leaf margins denticulate in upper part of leaf; costa ending
J	in leaf apex; lid remaining attached to ripe capsule by
	columella; brownish green plants to 1 cm tall and with seta
	5–10 mm long; on soil in saline habitats
	Hennediella heimii (Pottia heimii, Desmatodon heimii) (rr)
_	Leaf margins entire, costa mostly excurrent4
4	Capsule short, empty capsule wide-mouthed, widest at
	mouth, not (or only slightly) longer than wide; peristome
	seemingly absent (reduced to a basal membrane not
	protruding above the capsule mouth)5
-	Capsule longer, ovoid, ellipsoid or shortly cylindrical, empty
	capsule not wide-mouthed, widest at middle, distinctly
	longer than wide; peristome rudimentary, protruding above
	the capsule mouth6
5	Lid obliquely rostrate; plants (with sporophyte) mostly
	more than 5 mm tall; calyptra smooth; spores papillose; leaf
	margins mostly flat, occasionally slightly recurved; upper
	lamina cells smooth or slightly papillose; rhizoidal gemmae
	unknown
	Note: For the difference to <i>Tortula caucasica</i> see note under the latter.
-	Lid bluntly conical, with a short (erect) mamilla; plants
	(with sporophyte) often less than 5 mm tall; spores
	echinate; leaf margins distinctly recurved; upper lamina

cells distinctly papillose

Note: M. davallianum shares with M. muticum the bluntly conical capsule lid, but differs in capsules not contracted at mouth (widest at mouth), strongly widened at mouth when dry and empty, and the tip of the lid appearing more like a beak when capsules are dry, whereas in M. muticum the capsules are widest some distance below the mouth, in empty capsules the mouth remains narrow, and the lid of dry capsules takes up a more hemispherical shape upon drying, without differentiated mamilla or beak. The spores of both species can be echinate, but the spines are shorter in M. muticum, and spore ornamentation is sometimes a mixture of spines and warts. Spores of M. muticum are normally warty with hemispherical papillae 2.5 μ m high and placed rather distantly (4 μ m apart), whereas spores of M. davallianum have spines up to 5 μ m long, also on average 4 μ m distant from each other. Spores of M. davallianum are often larger, 22–42 μ m, often about 35 μ m, spores of M. muticum are 16–32 μ m, often about 25 μ m (Erzberger, unpublished).

Note: The presence (in *T. caucasica*) or absence (in *T. truncata*) of an annulus (recommended for separation of these species by Meinunger and Schröder (2007) and others, e.g. Smith (2004) and Chamberlain in Smith (1978)) appears not to be a reliable character (Ros and Werner in Guerra *et al.* 2006); however, the degree of development of the peristome (rudimentary in *T. caucasica* with usually several papillose segments prodruding above the capsule mouth; missing or reduced to a basal membrane not exceeding 20 μ m and not protruding beyond the capsule mouth in *T. truncata*) appears to be more reliable; see illustrations in Guerra *et al.* (2006).

Note: For the difference between this species and *M. davallianum*, see the note under that species. Capsules of *M. muticum* have longitudinal stripes caused by large spherical cells between the capsule wall and the spore sac

- that are arranged in longitudinal rows, best seen in fresh capsules just before dehiscence, but detectable also in dry specimens and older capsules. Capsules of *M. davallianum* seem to lack these stripes (Limpricht 1890: 529).
- 7(2) Plants small, (with sporophyte) often less than 5 mm tall; lid bluntly conical or mamillate; spores raspberry-like, with coarse warts but otherwise smooth walls; calyptra distinctly papillose in upper part; upper lamina cells densely and strongly papillose.....

Key to species of Pseudocrossidium

- Leaves narrowly lanceolate to lingulate, obtuse, apiculate; stalked multicellular gemmae present in leaf axils; plants brownish green, in dense tufts......

Key to species of *Pterygoneurum*

1 Capsule globose, without peristome, on short seta, immersed in the leaves; calyptra mitrate; costa excurrent in long spinose hyaline hair point almost twice as long as leaf; lamina cells papillose; plants low, to 2 mm tall......

-	Capsule exserted above leaves; calyptra cucullate; hyaline hair point smooth or only faintly denticulate, as long as leaf; lamina cells smooth
2	Plants to 1 cm high, seta to 10 mm long; peristome present (but fugacious); cells of lid in spirally twisted rows; lamellae towards leaf apex often dissolved into filaments with papillose end cells; spores 16–20 µm
_	Pterygoneurum lamellatum (r) Plants low, < 3 mm high, seta short, hardly exceeding 5 mm; peristome absent; cells of lid in straight rows; spores 25–30 µm
(see a	to species of <i>Syntrichia</i> llso combined key to <i>Syntrichia</i> and <i>Tortula</i> below) ences: Gallego (2005), Gallego <i>et al.</i> (2018), Homm (2017), Hedenäs <i>et al.</i>
1 - 2	Leaves without hair point, with or without mucro
3	Costa excurrent as a short mucro
-	Costa without hydroids; leaves not contracted in the middle; margin recurved to near apex; mucro consisting of stereids and papillose outer cells similar to lamina cells
4 (1)	Hair point spinulose to spinose, often strongly so
5	Lamina irregularly bistratose in upper third; leaves not contracted at mid-leaf; dorsal surface of costa strongly papillose, with simple or bifurcate papillae 2.5–5 µm long in the lower half, and pedicellate and branched papillae 12.5–

	37.5 µm long near the apex; leaf margin recurved from base
	to apex; costa in section generally with substereids; hair
	point strongly spinose, 0.3-2.6 mm long
	Syntrichia caninervis var. gypsophila
	(Tortula caninervis subsp. spuria) (rr)
-	Lamina unistratose in upper third 6
6	Costa in section without hydroids; leaves contracted at the
	middle or not
-	Costa in section with hydroids; leaves contracted at the
	middle 12
7	Hair point brownish red or orange, sometimes hyaline at
	apex; dorsal superficial costa cells in upper third similar to
	lamina cells <i>Syntrichia norvegica</i> (<i>Tortula norvegica</i>) (rr)
	Note: Many species of <i>Syntrichia</i> have hair points that are hyaline
	throughout except at the base, where they are often brownish. This is not to be confused with the hair point of <i>S. norvegica</i> , which is coloured
	throughout except sometimes at extreme apex. The leaf apex in S.
	norvegica is acuminate, acute to obtuse, and sometimes ascending the
	base of the hair point, a character also known in S. ruraliformis and S.
	subpapillosissima.
-	Hair point hyaline; dorsal superficial costa cells (stereids) in
^	upper third different from lamina cells
8	Costa in section with 1–2 (3) layers of dorsal stereids; leaves
	contracted at mid-leaf; margins plane or weakly recurved
	from base to mid-leaf; mid-leaf cells with 4–6 (8) papillae,
	papillae bifurcate, not pedicellate, 2.5 μm long; sometimes
	with spherical gemmae on ventral side of costa
	Syntrichia virescens (Tortula virescens) (c)
-	Costa in section with (2) 3–6 layers of dorsal stereids; leaves
	not contracted at mid-leaf; margins recurved from base to
9	upper third or apex
9	Margins recurved from base to upper third, rarely only to
	mid-leaf; mid-leaf cells 12.5–17.5 μm wide; leaves erectopatent, not recurved when moist
	•
	(0.2-1.7 mm), (ii) the short extent of the basal hyaline area (ca 25%), (iii)
	areolation of lamina cells hardly obscured by papillae, due to the
	predominantly central position of papillae and mamillosely-bulging cell
	walls on ventral and dorsal leaf surface. S. subpapillosissima is similar in
	that respect (mamillose cells with ± centrally placed papillae), however, that species has pedicellate and not sessile papillae.
	mas peareonate and not become pupmae.

S. calcicola with few stereid layers could be confused with *S. virescens*. However, apart from the characters mentioned in the key, *S. calcicola* differs (i) in relative length of the hyaline area formed by basal paracostal cells (19–25 (33)% in *S. calcicola*, 20–45% in *S. virescens*), (ii) in habitat (*S. calcicola* is saxi-terricolous, *S. virescens* mainly corticolous). Additional characters which may be used to avoid confusion with *S. montana* and *S. ruralis* are: (i) the size of mid-leaf cells (up to 17.5 μm wide in *S. calcicola*, 5–10 μm wide in *S. montana*, (5) 7.5–10 (15) μm wide in *S. ruralis*), (ii) the extent of the basal hyaline area (19–25 (33)% in *S. calcicola*, 27–45% in *S. ruralis*), (iii) the form of moist leaves in situ (patent in *S. calcicola*, squarrose in *S. ruralis*), (iv) the armation and length of the hair point (spinulose, 0.2–1.7 mm in *S. calcicola*, strongly spinose, 0.4–2.8 mm in *S. ruralis*, spinose, (0.3) 0.7–2.4 mm in *S. montana*).

- Margins recurved from base to apex, sometimes to upper third; mid-leaf cells (5) 7.5–12.5 μm wide; leaves mostly squarrosely recurved when moist.......10
- Papillae of mid-leaf cells not pedicellate, 2.5 μm long, (4) 6 (10) per cell......11

(Tortula papillosissima var. submamillosa) (w)

Note: *S. subpapillosissima* resembles *S. ruraliformis* in the acuminate apex, sometimes hyaline or dentate, ascending the base of the hair point. (Compare also *Hilpertia velenovskyi*.) It differs, however, in the structure and size of the constantly pedicellate papillae of lamina cells.

- Leaf apex not hyaline, rounded in general, not ascending the base of the hair point.....
 - Syntrichia ruralis (Tortula ruralis) (cc) Note: Differs from *S. montana* in the following characters: (i) leaves not contracted at middle, (ii) margin recurved to near apex (iii) lack of hydroids.
- Leaf apex in general hyaline, acuminate, ascending the base of the hair point.....
 - Note: According to some authors *S. ruraliformis* (Tortula ruraliformis) (r) Note: According to some authors *S. ruraliformis* has smaller mid-leaf cells than *S. ruralis* (Nyholm (1990): *ruraliformis* (6) 8–12 μ m wide versus *ruralis* 11–14 (16) μ m wide, Nebel and Philippi (2000): *ruraliformis* 8–14 μ m, *ruralis* 11–18 (20) μ m).

12 (6)	Hair point smooth or only weakly spinulose, (0.2) 0.4–0.9 (1.6) mm long; dorsal surface of costa smooth; stem with large central strand of cells becoming incrassate and brownish with age; plants usually corticolous
	Note: <i>S. laevipila</i> shares with <i>S. virescens</i> (i) the predominantly epiphytic habitat, (ii) the leaves contracted at the middle, (iii) degree of margin recurvature, (iv) lamina cell size; however, they can be separated by the following characters: (i) hair point smooth to weakly and indistinctly spinulose in <i>S. laevipila</i> , vs. distinctly spinulose in <i>S. virescens</i> , (ii) costa cross section with hydroids and 3–5 (7) layers of dorsal stereids in <i>S. laevipila</i> , lack of hydroids and only 1–2 (3) layers of stereids in <i>S. virescens</i> , (iii) leaf margin in <i>S. laevipila</i> may appear as a border of 2–5 rows of incrassate and less papillose cells, whereas this is not observed in <i>S. virescens</i> , (iv) if present, the different structure of gemmae (leaf-like in
-	S. laevipila, spherical in S. virescens). Hair point spinulose, (0.3) 0.7–2.4 mm long; dorsal surface of costa papillose, rarely smooth; stem without central strand or central strand weakly developed; plants usually
	saxicolous
	(Tortula crinita var. crinita) (w) Note: S. montana has the smallest lamina cells of all species with spinulose hair point, only $5-10$ (12.5) μ m wide.
13 (4)	Plants with gemmae 14
-	Plants without gemmae; leaves contracted at the middle; hair point smooth or slightly spinulose, (0.2) 0.4–0.9 (1.6) mm long; costa in cross section with hydroids; mostly corticolous; dioicous or autoicous
14	Gemmae in the form of leaves, papillose, near the stem apex or at the base of the upper leaves; lamina cells with 4–6 (8) bifurcate papillae, on both the ventral and dorsal surface; leaf margins plane or weakly recurved to mid-leaf
	Syntrichia laevipila (Tortula laevipila) (rr)
-	Gemmae globular, smooth, at the ventral face of leaves; lamina cells with 1–2 simple (sometimes bifurcate) papillae, on the dorsal surface only; leaf margins plane or incurved
	- J

Key to species of *Syntrichia* **and** *Tortula* **pp.** (excl. *T. acaulon, T. caucasica, T. lindbergii, T. protobryoides, T. truncata, incl. Tortula cernua, Hilpertia velenovskyi*)

1	Leaves without hair point, costa percurrent or excurrent in a mucro or apiculus
_	Leaves with hyaline or sometimes coloured hair point12
2	Costa ending in or below leaf apex, not excurrent; leaves contracted at the middle; plants often with spherical gemmae on the ventral surface of the leaf lamina
-	Costa percurrent or excurrent, if ending in leaf apex then leaves not contracted at the middle; plants without gemmae
3	Costa widened towards leaf apex, with large, papillose ventral cells; leaf margin revolute from base to apex4
-	Costa not widened towards apex; ventral costa cells not conspicuously large5
4	Leaves > 3 times as long as wide; mid-leaf cells 8–11 µm wide; dorsal superficial costa cells differentiated from stereids; marginal lamina cells not different from median lamina cells
-	Leaves 2.5–3 times as long as wide; mid-leaf cells 9–15 (17.5) μm wide; dorsal superficial costa cells not conspicuously different from stereids; 2–6 rows of marginal lamina cells less papillose and more incrassate than median lamina cells, but not forming distinct border
	Tortula atrovirens (r)
5	Upper and median lamina cells smooth or inconspicuously papillose; marginal cells in upper half of leaf quadrate similar to other lamina cells (but in general with more incrassate transverse walls)
-	Upper and median lamina cells papillose6
6	Costa percurrent or excurrent in short mucro, 10–200 µm long7
-	Costa excurrent in longer apiculus9

7	Central strand distinct; marginal cells differentiated from median lamina cells by being less papillose and having
	thicker walls, often oblate; costa ending in or below apex or
	excurrent in a short mucro, 10–40 µm long; autoicous;
	peristome with conspicuous tubular basal membrane
	making up about half of peristome length; peristome teeth
	strongly spirally twisted; plants of warm, dry, open habitats
	on base-rich soil or limestone
	Central strand indistinct; marginal leaf cells ± from base to
_	apex similar to median lamina cells with respect to shape,
	papillosity and wall thickness, not forming border; costa
	excurrent in short mucro 20–200 µm long8
8	Costa with hydroids in transverse section; leaves ±
0	contracted in the middle; margin plane in upper third; mucro
	20–60 μm long, consisting of stereids only
	montana var. calva (Tortula crinita var. calva) (rr)
	Costa without hydroids; leaves not contracted in the middle;
_	margin recurved to near apex; mucro 70–200 µm long,
	consisting of stereids and papillose outer cells similar to
	lamina cells
9	Leaf border conspicuous, formed by elongate to linear cells,
9	at least in lower half of leaf10
_	Marginal cells differentiated from median lamina cells by
	being larger and less papillose, but not forming distinct
	border; costa excurrent in short, yellowish awn or rarely
	ending in leaf apex; dioicous; peristome with low basal
	membrane, sometimes not visible above capsule mouth;
	peristome teeth not or weakly spirally twisted; plants of
	shady moist habitats
10	Capsule ovoid-cylindric, asymmetric, horizontal; peristome
	without conspicuous tubular basal membrane; leaves
	bordered in lower 2/3 by elongate, yellowish bistratose
	cells; central strand absent
_	Capsule cylindric, 4–6 mm long, erect; peristome with
	conspicuous tubular basal membrane making up about half
	of peristome length; leaves bordered in lower 2/3 by
	rectangular to linear cells in one layer or if bistratose then
	reaching near apex; central strand distinct11

11	Leaves narrowly lanceolate, acuminate, irregularly
	denticulate near apex; border of linear cells strongly
	developed, extending almost to apex, in at least one cell row
	bistratose
-	Leaves narrowly lingulate to ovate-lanceolate, obtuse to
	acuminate; border extending 1/2-3/4 way up leaf,
	unistratose
12 (1)	Hair point spinulose to spinose, often strongly so13
-	Hair point smooth or if very weakly spinulose then costa
	smooth on dorsal surface
13	Lamina irregularly bistratose in upper third; leaves not
	contracted at mid-leaf; dorsal surface of costa strongly
	papillose, with simple or bifurcate papillae 2.5–5 μm long in
	the lower half, and pedicellate and branched papillae 12.5-
	37.5 µm long near the apex; leaf margin recurved from base
	to apex; costa in section generally with substereids; hair
	point strongly spinose, 0.3–2.6 mm long
	Syntrichia caninervis var. gypsophila
	(Tortula caninervis subsp. spuria) (rr)
- 14	Lamina unistratose in upper third
14	middle or not
-	Costa in section with hydroids; leaves contracted at the
	middle
15	Hair point brownish red or orange, sometimes hyaline at
	apex; dorsal superficial costa cells in upper third similar to
	lamina cells Syntrichia norvegica (Tortula norvegica) (rr)
	Note: Many species of <i>Syntrichia</i> have hair points that are hyaline throughout except at the base, where they are often brownish. This is not
	to be confused with the hair point of <i>S. norvegica</i> , which is coloured
	throughout except sometimes at extreme apex. The leaf apex in S.
	norvegica is acuminate, acute to obtuse, and sometimes ascending the base
	of the hair point, a character also known in <i>S. ruraliformis</i> and <i>S. subpapillosissima</i> .
_	Hair point hyaline; dorsal superficial costa cells (stereids) in
	upper third different from lamina cells
16	Costa in section with 1–2 (3) layers of dorsal stereids; leaves
10	contracted at mid-leaf; margins plane or weakly recurved
	from base to mid-leaf; mid-leaf cells with 4–6 (8) papillae,
	papillae bifurcate, not pedicellate, 2.5 µm long; sometimes
	with spherical gemmae on the ventral face of the costa

	Syntrichia virescens (Tortula virescens) (W)
_	Costa in section with (2) 3–6 layers of dorsal stereids; leaves
	not contracted at mid-leaf; margins recurved from base to
	upper third or apex
17	Margins recurved from base to upper third, rarely only to
	mid-leaf; mid-leaf cells 12.5–17.5 µm wide; leaves erecto-
	patent, not recurved when moist
	Note: Additional characters of <i>S. calcicola</i> are: (i) a rather short hair point
	(0.2–1.7 mm), (ii) the short extent of the basal hyaline area (ca 25%), (iii)
	areolation of lamina cells hardly obscured by papillae, due to the
	predominantly central position of papillae and mamillosely-bulging cell
	walls on ventral and dorsal leaf surface. <i>S. subpapillosissima</i> is similar in that respect (mamillose cells with ± centrally placed papillae), however,
	that species has pedicellate and not sessile papillae.
	S. calcicola with few stereid layers could be confused with S. virescens.
	However, apart from the characters mentioned in the key, S. calcicola
	differs (i) in relative length of the hyaline area formed by basal paracostal
	cells (19–25 (33)% in <i>S. calcicola</i> , 20–45% in <i>S. virescens</i>), (ii) in habitat (<i>S. calcicola</i> , is apply a special to the calcicolate of the cal
	calcicola is saxi-terricolous, <i>S. virescens</i> mainly corticolous). Additional characters which may be used to avoid confusion with <i>S. montana</i> and <i>S.</i>
	ruralis are: (i) the size of mid-leaf cells (up to 17.5 µm wide in <i>S. calcicola</i> ,
	5–10 μm wide in <i>S. montana</i> , (5) 7.5–10 (15) μm wide in <i>S. ruralis</i>), (ii) the
	extent of the basal hyaline area (19–25 (33)% in <i>S. calcicola</i> , 27–45% in <i>S.</i>
	ruralis), (iii) the form of moist leaves in situ (patent in S. calcicola,
	squarrose in <i>S. ruralis</i>), (iv) the armation and length of the hair point (spinulose, 0.2–1.7 mm in <i>S. calcicola</i> , strongly spinose, 0.4–2.8 mm in <i>S.</i>
	ruralis, spinose, (0.3) 0.7–2.4 mm in S. montana).
_	Margins recurved from base to apex, sometimes to upper
	third; mid-leaf cells (5) 7.5–12.5 µm wide; leaves mostly
	squarrosely recurved when moist
18	Papillae of mid-leaf cells not pedicellate, 2.5 μm long, (4) 6
	(10) per cell
_	Papillae of mid-leaf cells pedicellate, (5) 7.5–10 μm long; (2)
	3-4 (6) papillae per cell, bifurcate, rarely branched and
	stellate at apex; papillae on dorsal surface of costa simple,
	2.5–5 μm long
	(Tortula papillosissima var. submamillosa) (w)
	Note: S. subpapillosissima resembles S. ruraliformis in the acuminate apex,
	sometimes hyaline or dentate, ascending the base of the hair point.
	(Compare also <i>Hilpertia velenovskyi</i> .) It differs, however, in the structure
	and size of the constantly pedicellate papillae of lamina cells.

19	Leaf apex not hyaline, rounded in general, not ascending the
	base of the hair point
	Syntrichia ruralis (Tortula ruralis) (cc)
	Note: Differs from <i>S. montana</i> in the following characters: (i) leaves not
	contracted at middle, (ii) margin recurved to near apex (iii) lack of hydroids.
-	Leaf apex in general hyaline, acuminate, ascending the base
	of the hair point.
	than <i>S. ruralis</i> (Nyholm 1990: <i>S. ruraliformis</i> (6) 8–12 µm wide versus <i>S.</i>
	ruralis 11–14 (16) μm wide, Nebel and Philippi 2000: S. ruraliformis 8–14
	μm, S. ruralis 11–18 (20) μm).
20 (14)	Hair point smooth or only weakly spinulose, (0.2) 0.4-0.9
	(1.6) mm long; dorsal surface of costa smooth; stem with
	large central strand of cells becoming incrassate and
	brownish with age; plants usually corticolous
	Syntrichia laevipila (Tortula laevipila) (rr)
	Note: S. laevipila shares with S. virescens (i) the predominantly epiphytic
	habitat, (ii) the leaves contracted at the middle, (iii) degree of margin
	recurvature, (iv) lamina cell size; however, they can be separated by the
	following characters: (i) costa cross section with hydroids and 3–5 (7)
	layers of dorsal stereids in <i>S. laevipila</i> , lack of hydroids and only 1–2 (3) layers of stereids in <i>S. virescens</i> , (ii) leaf margin in <i>S. laevipila</i> may appear
	as a border of 2–5 rows of incrassate and less papillose cells, whereas this
	is not observed in <i>S. virescens</i> , (iii) if present, the different structure of
	gemmae (leaf-like in S. laevipila, spherical in S. virescens).
-	Hair point spinulose, (0.3) 0.7-2.4 mm long; dorsal surface of
	costa papillose, rarely smooth; stem without central strand
	or central strand weakly developed; plants usually
	saxicolous
	(Tortula crinita var. crinita) (w)
	Note: S. montana has the smallest lamina cells of all species with spinulose
	hair point, only 5–10 (12.5) μm wide.
21 (12)	Plants with gemmae 22
-	Plants without gemmae 23
22	Gemmae in the form of leaves, papillose, near the stem apex
	or at the base of the upper leaves; lamina cells with 4-6 (8)
	bifurcate papillae, on both the ventral and dorsal surface;
	leaf margins plane or weakly recurved to mid-leaf
_	Gemmae globular, smooth, at the ventral face of leaves;
	lamina cells with 1–2 simple (sometimes bifurcate) papillae.

	on the dorsal surface only; leaf margins plane or incurved
23	Costa widened in upper third of leaf; cells on ventral surface
	of costa $20-25 \times (17.5) \ 20-25 \ \mu m$, subglobose, arranged in 1–2 layers; costa in transverse section with 2–3 layers of
	stereids; leaves strongly concave; hair point flexuose when
	dry, less so when moist
_	Costa not thickened in upper third of leaf; ventral superficial
	cells of costa inconspicuous, smooth or if papillose then
	similar to lamina cells; costa with 3-6 (7) layers of dorsal
	stereids; leaves slightly to strongly concave; hair point not
	flexuose24
24	Leaf margins strongly revolute (to 2 times) from \pm base to
	apex, entire or broadly toothed at or near the base of the
	awn, margins ascending the awn and thereby forming a ±
	hyaline triangle; cells of leaf apex rhomboidal to fusiform, epapillose; plants bud-like, leaves crowded, ovate to circular,
	1.3–2 mm long (including the awn); costa in transverse
	section with a single layer of epapillose ± wide cells at the
	ventral surface
_	Leaf margins plane or weakly recurved to mid-leaf or
	recurved to revolute from base to apex; leaf apex not hyaline,
	rounded to obtuse or emarginate, not ascending the base of
	the awn, made up of isodiametric papillose cells; plants not
	bud-like, leaves lingulate to spathulate, or linear-lingulate to
	ovate-lingulate, to 3.8 mm long; costa in transverse section
	with guide cells in 1–2 layers, and in addition with a ventral
25	layer of ± wide papillose cells similar to lamina cells
23	contracted at the middle; hair point smooth or slightly
	spinulose, (0.2) 0.4–0.9 (1.6) mm long; costa in cross section
	with hydroids; plants sometimes with leaf-like gemmae;
	mostly corticolous; dioicous or autoicous
-	Leaf margins recurved to revolute from base to near apex;
	leaves not contracted at the middle; hair point smooth, 0.3-2
	mm long; costa in cross section without hydroids; gemmae
	unknown; plants mostly saxicolous; monoicous

Key to species of *Tortula* References: Erzberger (1998), Košnar and Kolář (2009)

1	Costa percurrent or excurrent in mucro or in greenish or yellowish hair point
-	Costa excurrent in hyaline hair point
2	Costa widened towards leaf apex, with large, papillose ventral cells; leaf margin revolute from base to apex; costa percurrent to very shortly excurrent
-	Costa not widened towards apex; ventral costa cells not
•	conspicuously large; costa percurrent or excurrent
3	Leaves > 3 times as long as wide; mid-leaf cells 8–11 μm wide; dorsal superficial costa cells differentiated from stereids; marginal lamina cells not different from median
	lamina cells
	<i>Tortula muralis</i> subsp. <i>obtusifolia</i> (<i>Tortula obtusifolia</i>) (r)
-	Leaves 2.5–3 times as long as wide; mid-leaf cells 9–15 (17.5)
	μm wide; dorsal superficial costa cells not conspicuously
	different from stereids; 2–6 rows of marginal lamina cells
	less papillose and more incrassate than median lamina cells,
_	but not forming distinct border
4	Costa percurrent, in transverse section with hydroids, dorsal
	and ventral cells differentiated
_	Costa excurrent in short or long point
5	Lamina cells smooth or very faintly papillose6
-	Lamina cells conspicuously papillose 12
6	Capsule immersed or emergent, indehiscent
_	Capsule exserted, dehiscent9
7	Capsule emergent, ellipsoid to ovoid <i>Tortula protobryoides</i>
	(Pottia bryoides, Protobryum bryoides) (w)
-	Capsule immersed, ovoid to shortly ovoid8
8	Lamina cells completely smooth, occasionally ventral costa
	cells very weakly papillose; hair point 300–600 μm or longer;
	spores small, <25 (28) μm
	(Phascum cuspidatum var. piliferum) (w)
	Note: Moderately small spores (25 μ m) and smooth lamina cells are sometimes also observed in var. <i>acaulon</i> , therefore the length of the hair point is diagnostic for var. <i>pilifera</i> .

Lamina cells weakly to moderately papillose; hair point inconspicuous, shorter; spores 25–35 µm..... Tortula acaulon var. acaulon (*Phascum cuspidatum* var. *cuspidatum*) (w) Note: In the field, small plants of this species might be confused with Weissia longifolia. That species, however, has perichaetial leaves that from a sheathing, ovate base (hiding the capsule), are contracted to a long, ± parallel-sided lamina with an acute to acuminate apex, whereas *T. acaulon* has ovate-oblong perichaetial leaves ending in a conspicuous mucro. When dry, confusion is precluded because the leaves of Weissia are strongly crisped. Peristome well developed 10 9 10 Peristome with inconspicuous basal membrane to 70 µm high; upper lamina cells weakly papillose to smooth, 13-17 μm, thin-walled; costa in upper third covered by large, papillose ventral cells..... Peristome with conspicuous basal membrane $400-800~\mu m$ Note: The lack of papillae in this species is best seen in transverse sections of leaves; this should avoid confusion with weakly papillose forms of other species, e.g. T. subulata var. graeffii. Capsule short, empty capsule wide-mouthed, widest at 11 mouth, not (or only slightly) longer than wide; peristome seemingly absent (reduced to a basal membrane not protruding above the capsule mouth)..... Capsule longer, ovoid, ellipsoid or shortly cylindrical, empty capsule not wide-mouthed, widest at middle, distinctly longer than wide; peristome rudimentary, protruding above the capsule mouth; annulus present; upper lamina cells 12-25 μm wide, smooth or weakly papillose; costa in upper third with or without wide, smooth ventral cells; leaf margins meeting at acute angle at apex; rhizoidal gemmae sometimes Note: The presence (in T. caucasica) or absence (in T. truncata) of an annulus (recommended for separation of these species by Meinunger and Schröder (2007) and others, e.g. Smith (2004) and Chamberlain in Smith (1978) appears not to be a reliable character (Ros and Werner in Guerra et al. (2006)); however, the degree of development of the peristome (rudimentary in *T. caucasica* with usually several papillose segments prodruding above the capsule mouth; missing or reduced to a basal

	membrane not exceeding 20 µm and not protruding beyond the capsule mouth in <i>T. truncata</i>) appears to be more reliable; see illustrations in Ros and Werner in Guerra <i>et al.</i> (2006).
12 (5)	Capsule immersed
	(Phascum cuspidatum var. papillosum, P. cuspidatum var.
	mitraeforme) (r)
_	Capsule exserted 13
13	Capsule ovoid, cernuous to horizontal; leaves bordered
	below
_	Capsule cylindrical, erect; leaves bordered or not14
14	Peristome with conspicuous basal membrane 400–1600 μm
	high, visible above the capsule mouth as a long tube15
_	Peristome without conspicuous basal membrane
15	Leaves narrowly lanceolate, acuminate, irregularly
	denticulate near apex; border of linear cells strongly
	developed, extending almost to apex, in at least one cell row
	bistratose
-	Leaves narrowly lingulate to ovate-lanceolate, obtuse to
	acuminate; border extending 1/2-3/4 way up leaf,
	unistratose
16 (1)	Costa widened in upper third of leaf; cells on ventral surface
	of costa 20–25 \times (17.5) 20–25 μ m, subglobose, papillose,
	arranged in 1–2 layers; costa in transverse section with 2–3
	layers of stereids; leaves strongly concave; leaf margin
	recurved at leaf base and revolute at apex; hair point
	flexuose when dry, less so when moist
	Tortula brevissima (r)
-	Costa not thickened in upper third of leaf; ventral superficial
	cells of costa $1020 \times 8.817.5 \ \mu\text{m}$, papillose, arranged in a
	single layer; costa with 3-6 (7) layers of dorsal stereids;
	leaves slightly to strongly concave; hair point not flexuose

Key to varieties of Streblotrichum convolutum

Concerning the characters differentiating the two varieties of *S. convolutum, S. convolutum* var. *convolutum* and *S. convolutum* var. *commutatum,* there are contradictory statements in the literature. Although Frahm and Ahmed (2004), Frey *et al.* (2006) and Smith (2004) state that a central strand is lacking in the typical variety, other sources claim its presence in both varieties (Limpricht 1890, Guerra *et al.* 2006, and own observations). According to Kučera *et al.* (2013), rhizoidal tubers are present in both varieties, contrary to statements in Smith (2004) and Frey *et al.* (2006). Therefore, the following key is based only on the confirmed morphological differences.

-	Leaves small, < 1 mm long; basal leaf cells quadrate, incrassate; leaf margins flat, not undulate
	Streblotrichum convolutum var. convolutum
	(Barbula convoluta var. convoluta) (c)
-	Leaves > 1 mm long; basal leaf cells rectangular, thin-walled; leaf margins undulate
	Streblotrichum convolutum var. commutatum (Barbula convoluta var. commutata, B. convoluta var. sardoa) (r)

Key to species of Ephemerum

Reference: Ellis and Price (2015)

1	Leaves costate; leaf margin denticulate, sometimes in upper
	part of leaf only2
_	Leaves ecostate; leaf margins serrate
2	Upper leaves lanceolate to oblong-lanceolate, with 1-2
	asymmetric shoulders; branches of chloronema in fascicles;
	leaf margin denticulate in upper part
	Ephemerum cohaerens (r)
	Note: According to Infante et al. in Guerra et al. (2010), the costa may end
	below or in the leaf apex, or be excurrent. Orientation of leaf cells
	(diagonal rows vs. parallel rows) is not distinct according to the illustrations in Guerra <i>et al.</i> (2010).
-	Upper leaves lanceolate to linear-lanceolate, without
_	shoulders; branches of chloronema divergent
3	Capsule with straight apiculus; stomata all over the capsule
	surface; upper leaves erect; tuberous parts of rhizoid system
	packed with lipid drops and having oblique cross walls; leaf
	margin denticulate in upper part
	Ephemerum crassinervium subsp. sessile
	(Ephemerum sessile) (rr)
	(======================================

-	Capsule with curved apiculus; stomata confined to capsule base; upper leaves recurved; tuberous parts of rhizoid system packed with starch grains and having transverse cross walls; leaf margin denticulate
4	Ripe spores finely papillose, each covered by a whitish veil
	Ephemerum serratum
	(Ephemerum minutissimum Lindb.) (w) Note: This is not what was hitherto named E. serratum, but what was named E. minutissimum.
-	Ripe spores warty, not covered by veil
	Ephemerum stoloniferum
	$(=Ephemerum\ serratum\ auct.\ in\ traditional,\ but\ incorrect\ use)$ (rr)
Key	to species of <i>Gymnostomum</i> group
1	Basal lamina cells narrowly rectangular, (14) 16–40 (50) × (5) 6–7 μm, costa 20–40 μm wide at mid-leaf; leaves only 0.5 mm long, erect; plants tiny, only 2 mm tall; annulus of large inflated cells, persistent; peristome absent; brownish ovoid to elongate rhizoidal gemmae present among the tomentum <i>Gyroweisia tenuis</i> (rr)
-	Basal lamina cells short or long rectangular, 14–18 µm long, or if longer then costa 50–70 (90) µm wide at mid-leaf; leaves various; plants sometimes taller; annulus of small cells, persistent or falling
2	Plants > 5 mm high; leaves 5–7 times as long as wide
_	Plants 2–5 mm high; leaves very short, 0.4–0.5 mm long, broadly ovate to oblong, sometimes lingulate or spathulate, rarely ovate-lanceolate; 2–4 times as long as wide; multicellular gemmae present in leaf axils; margins of upper leaves unistratose; plants dull green
	Gymnostomum viridulum (r)
3	Plants vivid green, fresh green; costa in transverse section with dorsal band of stereids only, 25–45 μ m wide at leaf base; lamina cells 4–10 μ m; leaves 1 mm long; plants usually only a few millimeters (to 2 cm) tall; leaf scarcely tapered, and broadly rounded or obtuse at the apex
	Gymnostomum calcareum (w)

Key to species of *Hydrogonium*

References: Limpricht (1890), Nyholm (1989), Ahrens in Nebel and Philippi (2000), Garilleti in Guerra *et al.* (2006)

- Axillary gemmae numerous, clavate, consisting of few (ca 10) cells; dorsal side of costa with cells prorate at distal and proximal ends, appearing like paired mamillae.....
 - *Hydrogonium consanguineum* (Barbula indica auct.) (n.s.)
- Axillary gemmae few, ovoid, pointed at both ends, consisting of many (*ca* 100) cells; dorsal side of costa with simple papillae, without doubly prorate cells.....

Key to species of Tortella

References: Köckinger and Hedenäs (2017), Erzberger and Papp (2018)

- - Leaf margin entire; leaves not squarrose when moist; plants
- forming ± well defined cushions on soil or calcareous rock. 2

 Cells of ventral surface of costa linear, smooth, not covered by isodiametric papillose lamina cells; leaf apex broadly
- 3 Plants to 5 cm tall; leaves sometimes fragile; cells on dorsal surface of costa near leaf apex elongate and smooth (stereids), rarely papillose; stem mostly without central

	strand; peristome teeth twisted 1.5–3 times to the left
	Tortella tortuosa (w)
-	Plants usually smaller; leaves fragile with most tips broken off transversely; cells on dorsal surface of costa near intact leaf apex quadrate and papillose; stem mostly with central strand; peristome teeth twisted 1/2 turn to the left
4	Leaves when dry curled up to the apex, intact leaf tip often like a cork-screw; individual shoots of cushion well discernible from above and shaped like a ball of wool; marginal cells transversely elongate (wider than long) or quadrate, with bulging outer walls causing the margin to be neatly crenulate; thermophilic plants growing at low elevations
-	Dry leaves straight or slightly flexuose in their apical third and mostly oriented perpendicular to the shoot axis ('lying on the cushion surface'); individual shoots of cushion thereby less well discernible from above; marginal cells elongate (longer than wide) or quadrate, outer walls only slightly bulging; cryophilic plants of higher elevations
Key	to species of Trichostomum
-	Leaf apex cucullate, cuspidate; costa slightly excurrent; basal lamina cells rectangular, incrassate, upper lamina cells rounded, papillose; capsule elliptical, brownish, on a red seta; plants 1–2 cm tall; dioicous.
	Note: <i>T. viridulum</i> = <i>T. crispulum</i> var. <i>angustifolium</i> might merit specific rank; it produces often sporophytes and has very narrow, longly acuminate leaves (Meinunger and Schröder 2007: as species; illustrated in Schlüsslmayr 2005); taxonomic status controversial (Caspari <i>et al.</i> 2018).
-	Leaf apex not cucullate; costa usually excurrent as a mucro; capsule reddish brown, on a yellow seta

Key to species of Weissia

1	Capsule immersed in perichaetial leaves, on very short seta, lid not dehiscent 2
_	Capsule clearly exserted above the leaves
2	Seta shorter than the capsule; spores coarsely papillose; leaves narrow lanceolate from a broad sheathing base, with incurved margins and excurrent costa, strongly crispate
	when dry
-	Seta <i>ca</i> 1 mm long, longer than the capsule, mostly 2 capsules in a single perichaetium, clearly overtopped by the comal leaves; capsule without a peristome, closed by an
	epiphragm; lid differentiated but usually not dehiscent;
	plants to 5 mm tall
3	Spores > 20 μ m in diameter4
-	Spores < 20 µm in diameter
4	Capsule with peristome (but peristome mostly rudimentary); upper leaf margins hardly involute; capsule shortly cylindric, sulcate when dry (similar to <i>Cynodontium</i>) **Weissia rutilans** (r)
-	Capsule without a peristome, closed by an epiphragm, sometimes incomplete; exothecial cells with incrassate walls <i>Weissia brachycarpa</i> (w)
	Note: Attention should be paid to var. <i>obliqua</i> with leaves strongly incurved above and capsule oblong-ovate on short seta, when young often just emergent.
5	Peristome well developed, reddish-brown to yellowish brown (but may become pale with age); costa 30–80 μm wide
-	Peristome rudimentary and pale or lacking6
6	Peristome rudimentary, always pale, often not extending above capsule mouth; costa 70–100 µm wide near leaf base
-	Peristome completely lacking, capsule closed by an epiphragm
7	Costa 70–130 µm wide near leaf base <i>Weissia condensa</i> (w) Note: The leaf apex in <i>W. condensa</i> is cucullate like in <i>Trichostomum crispulum</i> , however, in that species the leaf margin is plane and not strongly inrolled as in <i>W. condensa</i> . <i>T. crispulum</i> is also a larger plant.
-	Costa up to 50 μm wide near leaf base; spores 17–34 μm
	Weissia hrachycarna (w)

	to species of Seligeriaceae rences: Nyholm (1987), Blockeel <i>et al.</i> (2000), Brugués and Guerra (2015)
1	Plants commonly > 1 cm tall; leaves with distinct alar cells
•	Blindia acuta (n.s.)
_	Plants usually < 1 cm tall, often minute; alar cells not distinct
	2
2	Leaves ± distinctly 3-ranked
_	Leaves not 3-ranked 4
3	Plants 2–5 (6) mm, blackish; leaves distinctly to indistinctly
•	trifarious, with long, smooth to nearly smooth subula, ±
	secund; peristome reduced; spores large, (16) 20-33 (40)
	μm, coarsely papillose; ventral costa superficial cells in
	section 8–11 μm, dorsal superficial costa cells in section 6–8
	μm
-	Plants 1–2.5 mm, light green; leaves indistinctly trifarious,
	with ± scabrous subula, ± recurved; peristome teeth
	lanceolate-triangular, 70–100 μm long, not reduced; spores
	ca 18 μm, smooth to slightly papillose; ventral costa
	superficial cells in section 6–9 μm, dorsal superficial costa
	cells in section 4–6 µm
4	Capsules without peristome, widened at mouth; leaf margins
	denticulate in lower third; seta to 2 mm long
	Seligeria donniana (r)
_	Capsules with peristome
5	Seta arcuate when moist; mature capsules ovoid to shortly
	cylindrical, narrowed at mouth when dry and empty.
	Blindiadelphus 6 Seta straight when moist; mature capsules semiglobose,
-	becoming wide-mouthed when dry and empty7
6	Leaves abruptly narrowed into long, fine subula completely
U	filled by the excurrent costa; costa usually 45–65 µm wide at
	base; plants mostly in crevices of moist siliceous rock
	protected by overhangs
_	Leaves gradually narrowed to broad acumen; costa ending
	in leaf apex, lamina extending into the leaf apex; costa
	usually 20–30 (40) µm wide at base; plants exclusively on
	small dolomitic stones at ground level below shaded rock
	walls in Corno-Quercetum
	Blindiadelphus campylopodus (Seliaeria campylopoda) (rr)

7 Narrow and linear leaves mixed with broader leaves; spores (9) 10–14 μm.....8 Leaves from ovate-lanceolate base narrowed into stout point, formed by excurrent costa; leaf lamina opaque in Perichaetial leaves long, reaching or almost reaching 8 capsule, longly excurrent costa filling subula; seta surface cells short rectangular, mostly 15-26 µm long, 2-4 times as Seta much longer than perichaetial leaves, percurrent costa not filling subula, lamina extending to apex; seta surface cells narrow, linear, mostly $28-50~\mu m$ long, 4-8 times as **Key to species of Grimmiaceae** (Coscinodon: 3; Grimmia: 24–64; Racomitrium: 4–9; Schistidium: 10–23) References: Blom (1996), Blom in Nyholm (1998), Erzberger and Schröder (2008), Erzberger (2009), Maier (2010), Erzberger et al. (2016) 1 Basal leaf cells elongate, with incrassate nodulose-sinuose walls; seta elongate (capsules exserted), mostly dextrorse (forming a right-handed helix like a normal screw) when dry (sinistrorse in R. canescens, R. lanuginosum); stem without central strand. *Racomitrium* 4 Basal leaf cells never at the same time elongate and incrassate nodulose-sinuose; seta often short, untwisted or Costa in transverse section nearly homogeneous, composed 2 of cells of ± equal size; seta short, capsule immersed; columella attached to operculum, both fall off the capsule together; stem (of sporophyte-bearing plants) often with central strand (central strand lacking or narrow and indistinct in S. apocarpum, S. brunnescens subsp. griseum, S. elegantulum, S. flaccidum, S. lancifolium, S. papillosum). Schistidium 10 Costa in transverse section composed of cells of different size, usually with large guide cells, often with stereids or hydroids; capsules immersed to exserted; columella not attached to lid, remaining in the urn when the capsule

3	Leaves plicate, often at leaf base with additional costa-like structures of several layers; calyptra large, campanulate- mitrate, covering nearly all of ripe urn; peristome teeth cribrose (extensively perforated); stem with a central strand
-	of few cells <i>Coscinodon cribrosus</i> (rr) Leaves plicate or not, rarely with additional costa-like structures; calyptra smaller, mostly mitrate (cucullate in <i>G. ovalis, G. montana, G. crinita, G. orbicularis</i>), peristome teeth usually not or only slightly perforated; stem with central strand (absent in <i>G. hartmanii</i> and sometimes <i>G. elatior</i>). <i>Grimmia</i>
4 (1)	Hair point present, papillose; seta sinistrorse when dry5
_	Hair point absent or present, not papillose, but often denticulate; seta dextrorse when dry6
5	Lamina smooth; hair point papillose and sinuose-erose dentate, decurrent at margins; seta strongly papillose
-	Lamina papillose; hair point not sinuose-erose dentate, decurrent or not; seta smooth
6	Hair point absent; lamina cells papillose, but papillae flat and inconspicuous (best seen in cross section). Subgenus <i>Codriophorus</i>
-	Hair point usually present, but sometimes lacking; lamina cells not papillose (but sometimes pseudopapillose with projecting cell walls). Subgenus <i>Bucklandiella</i> 8
7	Leaves broadly lanceolate; leaf apex broadly rounded with distant teeth; alar cells forming distinct ± decurrent auricles; supra-alar cells not differentiated
-	Racomitrium aciculare (Codriophorus acicularis) (n.s.) Leaves narrow lanceolate with obtuse apex, without teeth, at most very slightly denticulate by prominent cell corners; alar cells not differentiated; supra-alar cells hyaline or yellowish-hyaline, incrassate and esinuose, forming a distinct marginal band
8	Basal lamina cells nodulose; ventral surface of costa in cross section flat towards base, not canaliculate; 4–8 guide cells free at ventral surface in the median part; hair point (if present) with wide insertion, often slightly decurrent9

Basal lamina cells thick-walled and porose, but not nodulose; ventral side of costa in cross section canaliculate (with a ± rounded profile); 2–4 guide cells free at ventral surface; costa in mid-leaf mostly bistratose, narrow (60–80 μm) in lower part (costa 2 (3)-stratose with 3–4 ventral cells in basal part, 2-stratose with 2–3 ventral cells in mid-leaf, 2-stratose with 2 ventral cells in upper part); hair point lacking or short (< 0.4 mm); marginal supra-alar cells in conspicuous row consisting of 10–15 hyaline or yellowish-hyaline, thin-walled, pellucid esinuose cells......

......Racomitrium microcarpon

(Bucklandiella microcarpa) (n.s.)

9 Costa predominantly bistratose, weakly convex on dorsal side; reaching to hair point; 4–8 guide cells free at ventral surface; hair point usually well developed and flexuose, but absent in some Hungarian populations; leaf margin unistratose, narrowly revolute to 3/4 leaf length; innermost perichaetial leaf ovate, rarely brevipilose, hyaline.......

Racomitrium heterostichum

(Bucklandiella heterosticha) (rr)

12	Perichaetial leaves broadly elliptical to broadly ovate, distinctly wider than upper stem leaves; lamina cells small, rounded, 6–9 µm in upper part of leaf; central strand of 3–10
	cells; papillae frequent on dorsal and ventral sides of lamina; plants of siliceous substrates
-	Perichaetial leaves not distinctly wider than upper stem leaves; lamina cells larger, ± sinuose, 10–12 μm in upper
	part of leaf; central strand of 20–30 cells; papillae usually more frequent on dorsal than on ventral side of lamina; plants of usually calcareous substrates
13 (10)	Exothecial cells in lower third of capsule predominantly
	isodiametric, but sometimes mixed with patches of elongate
	cells; urn length: width ratio often less or equal to 1.3; leaf
	margins or back of costa mostly denticulate with papillae or
	projecting cells in upper third of leaf
-	Exothecial cells predominantly elongate; urn length: width
	ratio usually > 1.3; leaf margins and dorsal side of costa
	smooth or denticulate (S. crassipilum, S. helveticum; for S.
14	dupretii see 15)
14	hyaline cells only; plants on stones near running water
	(Danube)
_	Spores smaller, 8–13 μm; hair point various, but usually
	present; plants not specifically riverine
15	Exothecial cells irregular in size and shape, forming an
	irregular pattern; perichaetial leaves narrow, 0.5-0.6 mm
	wide, hardly concealing the urn; hair point narrow at
	insertion, consisting of few cells, up to 0.15 (0.25) mm long;
	at least some leaves with smooth margins; plants small
	Schistidium dupretii (w)
-	Exothecial cells usually isodiametric, often quadrate, but
	sometimes with patches of elongate cells, forming a ±
	regular to very regular pattern; perichaetial leaves narrow
	or wide, usually concealing major part of the urn; hair point
	various, but not conspicuously narrower than the chlorophyllose part of the leaf apex; margins usually
	distinctly to conspicuously denticulate-papillose
16	Urn 0.7–1.0 mm long and 0.5–0.65 mm wide; leaves
	narrowly lanceolate, margins and back of costa strongly

	papillose down to below mid-leaf, with size of papillae increasing towards apex; exothecial cells arranged in neat vertical rows; peristome teeth 320–400 µm long; perichaetial leaves 0.5–0.8 mm wide; plants small
-	Urn 0.9–1.35 mm long and 0.5–0.85 mm wide; leaves broader, with costa and margins less distinctly papillose; exothecial cells less neatly arranged; peristome teeth 400–700 µm long; perichaetial leaves 0.8–1.5 mm wide; plants medium-sized
17 (13)	K+ yellow
17(13)	K+ red 19
18	Peristome usually lacking; hair point usually long, 0.15–0.85
10	mm, finely denticulate; perichaetial leaves plicate; lid mamillate; plants small to medium sized
-	Peristome well developed; hair point short, 0-0.45 mm,
	coarsely spinulose; lid rostrate, but beak sometimes short;
	plants small Schistidium confertum (rr)
19	Leaf margins or back of costa denticulate with papillae or projecting cells; peristome teeth 350–400 μm long, in freshly deoperculate capsules stellate-straight, scarcely
	twisted, orange; urn usually with 0–4 (6) stomata at base, but sometimes more, pores often rudimentary or lacking; central strand distinct, > 6 cells <i>Schistidium crassipilum</i> (c)
-	Leaf margins and dorsal side of costa smooth (few papillae may occur at leaf apex immediately below the hair point, in
	S. helveticum also further down)20
20	Stomata absent (absent to few in forms of <i>S. crassipilum</i> with
_0	smooth margins and costa, see 19); plants small, in compact cushions
_	Stomata present, (3) 6–10 per urn; plants medium-sized to
	large with conspicuous hair points23
21	Plants usually black, glossy; hair point absent or short (< 0.2
	mm) on all leaves, coarse and coarsely spinulose with short
	and broad, patent to squarrose spinulae, terete, not
	decurrent; exothecial cells forming an irregular pattern
	Schistidium helveticum (Schistidium singarense) (w)
_	Plants brownish; hair point absent or short in lower leaves,
	conspicuous in perichaetial leaves, 0.15–1.0 mm long, coarse

	and terete, but flattened towards insertion, not or shortly
	decurrent, but embracing parts of upper lamina; exothecial
	cells up to 50-80 µm long, forming a regular pattern.
	Schistidium brunnescens 22
22	Leaves ovate-lanceolate to ovate triangular, often with
	shoulder, 1.85-2.6 mm long, usually with longitudinal
	bistratose ridge-like striae on one or both sides of costa;
	costa 58–90 μm wide in central part of leaf
	Schistidium brunnescens subsp. griseum (r)
-	Leaves oblong, ovate or short ovate-triangular, 1.1–2.0 mm
	long, shoulders and ridge-like striae absent; costa 38–60 μm
	wide in central part of leaf
	Schistidium brunnescens subsp. brunnescens (w)
23 (20)	Hair point erect, terete, narrow throughout, not decurrent,
	hyaline, but often brownish at insertion; central strand
	absent; urn usually ovate in shape; upper lamina cells
	isodiametric, not sinuose, not distinctly incrassate; lamina
	with irregular bistratose patches; plants in mats or
	decumbent tufts on ± shaded calcareous rocks
	Schistidium elegantulum subsp. elegantulum (w)
-	Hair point stiff and straight, usually flattened at base,
	decurrent; central strand always distinct (> 20 cells); urn
	usually cylindrical in shape; upper and especially lower
	lamina cells strongly sinuose, distinctly incrassate (similar
	to cells of Racomitrium); lamina rarely with bistratose
	patches; plants in hoary tufts on exposed or moderately
	shaded calcareous rocks
24 (3)	Plants with sporophytes25
-	Plants without sporophytes
25	Seta short, not longer than capsule, capsule immersed to
	emergent 26
-	Seta longer than capsule, capsule exserted above the cushion
	31
26	Seta arcuate (moist)27
-	Seta straight (moist)30
27	Lamina bistratose from apex to widest part of leaf,
	semicircular in section; costa only slightly prominent on
	dorsal side; peristome present
-	Lamina unistratose or with bistratose patches, margins
	unistratose or histratose leaves + concave costa +

20	Professional about leaves and the best mark and a
28	Peristome absent; leaves concave in basal part only,
	obtusely keeled towards apex; lamina unistratose with
	bistratose margin and bistratose patches.
	Grimmia anodon (r)
_	Peristome present; lamina completely unistratose (margins
20	sometimes locally bistratose)
29	Leaves spathulate, widest above mid-leaf; hair point long to
	very long, broadly inserted; margin completely unistratose;
	seta as long as capsule
-	Leaves not spathulate, widest at or below mid-leaf; hair
	point not particularly long, with narrow insertion; margin
	partly bistratose; seta shorter than capsule
20(26)	Grimmia plagiopodia (r)
30 (26)	Leaves plicate; costa recessed in deep ventral groove; some
	plicae pluristratose at base, resembling additional costae;
	calyptra wide, campanulate, plicate, nearly enclosing the urn; peristome cribrose
	Leaves not plicate, semicircular in section; other characters
-	different
31 (25)	Seta arcuate (moist)32
31(23)	Seta straight (moist)
32	Lamina bistratose in upper part of leaf; costa recessed in a
32	deep grove, prominent on dorsal side, in transverse section
	without hydroids but with an additional layer of wide cells
	on the dorsal side of the guide cells; lamina and costa usually
	with scattered hemispherical papillae; central strand absent
	in weak stems and branches
	Note: Compare also <i>G. trichophylla</i> with guide cells in 2 layers.
_	Lamina usually unistratose in upper part of leaf, sometimes
	with bistratose margins or striae; costa not grooved; in
	transverse section with hydroids, without additional wide
	cells (except in G. trichophylla); lamina and costa epapillose;
	stems with central strand
33	Leaves ovate, widest at or shortly above mid-leaf, apex ±
	broad; lamina usually unistratose, margins unistratose or
	bistratose in 1 or more cell rows; gemmae unknown34
_	Leaves lanceolate, from ovate base tapering to acuminate
	apex; lamina unistratose or with bistratose patches, margins
	unistratose or bistratose; gemmae sometimes present35

34	Basal paracostal cells usually elongate, 4–8 times as long as wide; capsule ovate, lid mamillate, flat; leaf margins
	unistratose or bistratose in 1 cell row at one side of leaf only,
	not thickened, recurved on both sides at mid-leaf; costa
	slightly attenuate towards leaf base, mamillose dorsally at
	insertion
_	Basal paracostal cells mostly shortly rectangular to
	quadrate, 2-4 times as long as wide; capsule oblong, lid
	usually rostrate; leaf margins pluristratose, thickened,
	recurved on both sides, on one side from base, on the other
	side from above base to above widest part of leaf; costa not
	attenuate towards leaf base
35	Leaves strongly asymmetric, one side rounded in outline,
	opposite side straight; margin of straight side recurved;
	costa S-shaped, towards apex not median, but approaching
	one of the margins; in cross section at mid-leaf with two
	narrowly elliptical guide cells arranged obliquely with
	respect to leaf plane; rope-like young shoots present in the
	interior of the cushion; gemmae unknown; narrow central
	strand of thin-walled cells contrasting sharply with the
	incrassate orange-tinged stereid-like cortical cells
	Grimmia funalis (rr)
-	Leaves symmetric to slightly asymmetric, costa not S-
	shaped, median at leaf apex; young shoots of different shape
	or absent; short-stalked gemmae sometimes present at leaf
	base
36 (35;	Costa above mid-leaf rectangular in section, with edges
64)	angulate to winged; hair point conspicuously denticulate;
	lamina sometimes partly bistratose; gemmae sometimes
	present
	Note: G. hartmanii also can have an angulate costa, but lacks a central strand.
_	Costa in section rounded, not angulate or winged; hair point
	denticulate or smooth; gemmae present or absent37
37 (36;	Lamina cells at leaf apex longer than wide, slightly sinuose;
52)	costa very wide at leaf base, sometimes $> 80 \mu m$, with 6 (8)
32)	guide cells at insertion; leaf margin revolute on one side,
	recurved on opposite side; hair point denticulate; gemmae
	unknown
	Complete (1)

-	Lamina cells at leaf apex rounded; costa usually less wide;
	hair point nearly smooth; gemmae sometimes present38
38	Costa in transverse section at leaf insertion and leaf base
	with two layers of enlarged cells, i.e. 1–2 (3) cells in addition
	to ventral guide cells, in size and position intermediate
	between guide cells and other costal cells; plants of siliceous
	substrates
-	Guide cells at leaf base in one layer; substrate various39
39	Guide cells 4 at leaf insertion; leaves erecto-patent to
	spreading when moist; plants usually on calcareous
	substrates
-	Guide cells 6 at leaf insertion; leaves erecto-patent to
	squarrose when moist; plants of siliceous substrates, often
	near water
40 (31)	Leaves semicircular in transverse section ± from base to
	apex41
-	Leaves ± keeled, not semicircular in transverse section (G.
	longirostris has leaves keeled in basal and semicircular in
	apical part)42
41	Leaves ovate, usually with rounded apex; detached leaves lie
	flat on a slide; margin unistratose in bistratose upper part of
	leaf; most basal cells oblate, incrassate, few elongate
	paracostal cells
-	Leaves from ovate base lanceolate, tapering from distinct
	shoulder to narrow apex; detached leaves on a slide usually
	fold along the costa and appear in lateral view; margin
	bistratose; most basal cells elongate, some nodulose,
	towards margin shorter
42	Costa in transverse section recessed in a deep groove in
	upper part of leaf; leaves with distinct shoulder; basal cells
	short rectangular to quadrate with incrassate transverse
	walls; hair point denticulate
-	Costa not recessed in groove; leaves without shoulder; basal
	cells elongate-rectangular with thin transverse walls; hair
	point smooth, sometimes short (< 0.4 mm)43
43	Stem without central strand; plants usually with numerous
	short branches; basal lamina cells with strongly thickened
	longitudinal walls, porose, cells from leaf base to mid-leaf
	elongate and nodulose; ventral side of costa in cross section
	canaliculate (with a + rounded profile). 2-4 guide cells free

-	at ventral surface, bistratose at mid leaf composed of 2 types of cells; hair point lacking or short (< 0.4 mm), marginal supra-alar cells in conspicuous row consisting of 10–15 hyaline or yellowish-hyaline, thin-walled, pellucid esinuose cells; margins recurved to revolute on both sides
	types of cells (guide cells, substereids and larger dorsal cells); margin recurved on one side
	Grimmia longirostris (rr)
44 (24)	Leaf apex without any hyaline cell; leaves strongly asymmetric, costa towards apex not median, but approaching one of the margins; rope-like young shoots present in the interior of the cushion (muticous leaves can occur in <i>G. anodon</i> , too)(male plants of)
-	Leaf apex with hair point or at least with some hyaline cells
45	Central strand absent; hair point lacking or short (< 0.4 mm), or with cell lumina clearly visible, denticulate; globular
-	multicellular gemmae sometimes present at leaf tips
46	Plants sparsely branched; basal lamina cells not conspicuously incrassate or porose, towards margin usually short but neither widened nor hyaline; costa at mid-leaf usually 3–5-stratose; cells of leaf apex with cell lumen clearly visible; hair point denticulate; globular multicellular gemmae often present at leaf tips (or some leaf tips deformed due to gemma production)
-	Plants usually with numerous short branches; basal lamina cells with strongly thickened longitudinal and thin transverse walls, porose, cells from leaf base to mid-leaf elongate and nodulose; ventral side of costa in cross section

	canaliculate (with a ± rounded profile); 2–4 guide cells free at ventral surface; hair point lacking or short (< 0.4 mm), marginal supra-alar cells in conspicuous row consisting of 10–15 hyaline or yellowish-hyaline, thin-walled, pellucid esinuose cells; costa at mid-leaf mostly bistratose
	(Bucklandiella microcarpa) (n.s.)
47	Lamina usually unistratose in upper part of leaf, margin unistratose or pluristratose in one or several rows48
-	Lamina bistratose in upper part of leaf or lamina unistratose with some bistratose cell rows, margin pluristratose in several rows
48	Lamina unistratose, margin unistratose (or at one side of leaf a single marginal row bistratose)49
-	Lamina usually unistratose, margin pluristratose in one or several rows
49	Leaves spathulate, widest above mid-leaf; margin completely unistratose, plane
_	Leaves not spathulate, widest at or below mid-leaf; margin unistratose or partly bistratose, plane or recurved50
50	Leaves spoon-like, concave; margin unistratose to partly bistratose, plane; basal cells short. <i>Grimmia plagiopodia</i> (r)
_	Leaves not spoon-like, not concave, with an obtuse keel, leaf halves forming a right or obtuse angle in transverse section; margins unistratose or on one side of leaf a single unthickened bistratose row of cells, recurved on both sides around mid-leaf; costa slightly attenuate towards leaf base, mamillose dorsally at insertion; basal paracostal cells usually elongate, 4–8 times as long as wide.
	Grimmia orbicularis (w)
51	Leaves strongly asymmetric, one side rounded in outline, opposite side straight; margin of straight side recurved; costa S-shaped, towards apex not median, but approaching one of the margins; in cross section at mid-leaf with two narrowly elliptical guide cells arranged obliquely with respect to leaf plane; rope-like young shoots present in the interior of the cushion; gemmae unknown; narrow central strand of thin-walled cells contrasting sharply with the incrassate orange-tinged stereid-like cortical cells

-	Leaves ± symmetric, costa not S-shaped, median towards
	apex; young shoots of different shape or absent; short-
=0	stalked gemmae sometimes at leaf base
52	Leaves ovate, widest at or shortly above mid-leaf; apex ±
	broad; leaf margins pluristratose, thickened, recurved on
	both sides, on one side from base, on the other side from
	above base to above widest part of leaf; leaf halves forming
	approximately a right angle in transverse section; basal cells
	mostly shortly rectangular to quadrate, 2–4 times as long as
	wide; gemmae unknown
-	Leaves lanceolate, from ovate base tapering to acuminate
	apex; gemmae sometimes present
53 (47)	Lamina continuously bistratose at least in upper part of leaf
	(in <i>G. laevigata</i> margin in part unistratose)54
-	Lamina unistratose even in upper part of leaf, with some cell
	rows bistratose; margin bistratose in several rows62
54	Leaves semicircular in transverse section
_	Leaves not semicircular in transverse section
55	Costa biconvex, prominent on ventral and dorsal sides, with
	a centrally placed group of hydroids, guide cells indistinct;
	plants of exposed calcareous rocks
	Grimmia teretinervis (Schistidium teretinerve) (n.s.)
_	Costa not biconvex, guide cells distinct; substrate calcareous
	or siliceous
56	Leaves from ovate base lanceolate, tapering from distinct
	shoulder to narrow apex; detached leaves on a slide usually
	fold along the costa and appear in lateral view; margin
	bistratose; most basal cells elongate, some nodulose,
	towards margin shorter
_	Leaves ovate, usually rounded or obtuse at apex; detached
	leaves lie flat on a slide; margin bistratose or unistratose;
	basal cells short or long
57	Margin towards leaf base unistratose in a widening zone;
37	guide cells in a shallow groove with incrassate ventral walls
	in mid-leaf; most basal cells oblate, incrassate, few
	paracostal cells elongate; hair point strongly denticulate
	Mote: <i>G. laevigata</i> is usually confined to siliceous rocks (Erzberger 2009).
	Its tufts disintegrate easily.

-	Margin bistratose throughout bistratose part of lamina;
	costa not grooved, guide cells hardly discernible, ventral
	walls not incrassate; basal paracostal elongate cells usually
	extending far towards margin, where cells are shorter; hair
= 0(= 4)	point smooth or faintly denticulate. <i>Grimmia tergestina</i> (w)
58 (54)	Costa in transverse section of upper part of leaf appearing
	recessed in a ± deep groove; leaves with or without
	shoulder; basal cells with thin or \pm incrassate transverse
	walls
-	Costa not grooved ventrally; leaves without shoulder; basal
=0	cells elongate with thin transverse walls
59	Costa without hydroids, ventral side of pluristratose costa
	with 1 to several layers of wide cells of guide-cell-like
	appearance; lamina and costa usually with scattered
	hemispherical papillae, rarely smooth; a wide band of
	strongly elongate paracostal cells with thin transverse walls
	present at leaf base; leaves without distinct shoulder
	Grimmia elatior (rr)
-	Costa with hydroids; costa and lamina without
	hemispherical papillae; basal cells with ± incrassate
	transverse walls, paracostal basal cells shorter, elongate in
	few rows; leaves with ± distinct shoulder60
60	Leaves strongly plicate, some plicae pluristratose at base,
	resembling additional costae; shoulder poorly developed;
	hair point nearly smooth
-	Leaves faintly plicate, without additional costae; shoulder
	distinct; hair point denticulate
61 (58)	Costa above mid-leaf rectangular in section, edges angulate
	to winged; in cross section at insertion with 4 (6) guide cells;
	leaf margin thickened, recurved on one side and revolute on
	the other; hair point denticulate; capsule ribbed when dry;
	short-stalked gemmae sometimes at leaf base
	Grimmia muehlenbeckii (w)
-	Costa rounded at dorsal side, but occasionally unevenly so to
	somewhat angulate in lower part of leaf; in cross section at
	insertion with 6, rarely only 4 guide cells; leaf margin not
	thickened, usually recurved on one side only; hair point
	smooth; capsule smooth when dry

62 (53)	a centrally placed group of hydroids, guide cells indistinct; plants of exposed calcareous rocks
-	Costa not biconvex, guide cells distinct; substrate calcareous or siliceous
63	Leaves strongly asymmetric, one side rounded in outline, opposite side straight; margin of straight side recurved; costa S-shaped, towards apex not median, but approaching one of the margins; in cross section at mid-leaf with two narrowly elliptical guide cells arranged obliquely with respect to leaf plane; rope-like young shoots present in the interior of the cushion; gemmae unknown; narrow central strand of thin-walled cells contrasting sharply with the incrassate orange-tinged stereid-like cortical cells
	Grimmia funalis (rr)
-	Leaves symmetric to slightly asymmetric, costa median at leaf apex; young shoots of different shape or absent; short-stalked gemmae sometimes present at leaf base
64	Leaves ovate, widest at mid-leaf; basal cells shortly rectangular, marginal cells incrassate; margins plane throughout; gemmae unknown
-	Leaves lanceolate, from ovate base tapering to acuminate apex; margin recurved to revolute at least on one side; gemmae sometimes present
	to species of <i>Hedwigia</i> ences: Hedenäs (1994), Erzberger (1996)
1	Mid-leaf cells mostly with 1, rarely with 2 peltate papillae per cell on dorsal side; hyaline hair point usually reflexed when dry
-	Mid-leaf cells predominantly with several, ± branched papillae per cell on dorsal side; leaves straight or falcate2
2	Hyaline hair point of sterile shoots conspicuously white, strongly contrasting with the leaf lamina, occupying 1/4–

1/2 of leaf length; median lamina cells often with 2 papillae placed near proximal and distal cell ends on dorsal side, the papillae of adjacent cells seemingly merging together,

Key to species of Bartramiaceae

Note on the examination of *Philonotis*: Since the species of this genus show a high degree of variability, it is advisable to examine well-grown plants and avoid juvenile stages. This also applies to the examination of leaves, which should be taken from the tomentose part of the stem, i.e. growth of the previous year, since the first leaves of a season tend to be untypical.

The shape of perigonial leaves (acuminate or obtuse) is not a reliable character for species identification: Meinunger and Schröder (2007) 3: 94.

References: Buryová (unpublished), Bergamini (2001), Frey et al. (2006)

1	Leaves lanceolate, not sheathing at base. <i>Philonotis</i>
_	Leaves narrowly lanceolate to linear lanceolate, sheathing at
	base2
2	Lamina cells with striate cuticle; leaves curled or crispate
	when dry; seta about 1 cm long with erect, globose
	capsule
-	Lamina cells mamillose. <i>Bartramia</i>
3	Leaves with whitish sheathing base, abruptly contracted
	into lanceolate limb; cells mostly elongate, rarely quadrate
	to shortly rectangular
-	Leaf base weakly sheathing, not conspicuously whitish; cells
	shorter, at most shortly rectangular4
4	Capsule overtopping leaves on long straight seta; tufts to 5
	cm tall
_	Capsule immersed among leaves on short seta, appearing
	lateral; plants to > 10 cm tall
5 (1)	All lamina cells with mamilla at distal end; leaves 1.2–2 mm
3(1)	long, 2.5–4 times as long as wide, margins plane, not
	recurved
	1 monous marcinea (11)

-	Lamina cells only in upper half of leaf with mamillae at distal end, lower cells smooth, or at least some mamillae at
	proximal end (especially in lower part of leaf)6
6	Lamina cells only in upper half of leaf with mamillae at distal
U	end (some cells without mamilla), lower cells smooth; leaves
	short, 0.6–0.9 (1) mm long
-	Lamina cells with mamillae at proximal end, or some with
	mamillae in mid-cell, or cells with mamillae at distal end
7	only in uppermost part of leaf or margin
7	Cells uniformly rectangular throughout, of ± equal width;
	margin mostly plane with single teeth; leaves not plicate
	Philonotis caespitosa (r)
-	Upper lamina cells ± linear, distinctly narrower than basal
	rectangular cells; margin recurved, partly toothed with
	geminate teeth; leaves plicate or not
8	Basal paracostal cells > $60 \mu m$ long (at least some cells),
	conspicuously translucent, leaves distinctly homomallous,
	gradually tapering, hardly plicate below; costa very robust,
	60–180 μm wide near base <i>Philonotis calcarea</i> (rr)
-	Basal paracostal cells up to 50 μm long, not conspicuously
	translucent; leaves rarely weakly homomallous, ± abruptly
	tapering from broad base to narrow acumen, distinctly
	plicate below; costa usually only to 90 µm wide near base
	to species of Bryaceae
	ences: Demaret (1993), Nyholm (1993), Ahrens in Nebel and Philippi (2001)
Guerra	a et al. (2010), Erzberger and Schröder (2013)
4	
1	Plants with subterranean stolons and erect secondary stems
	with large terminal rosette and scale-like lower leaves,
	comal leaves in general > 5 mm long. <i>Rhodobryum</i> 2
-	Plants without a conspicuous terminal rosette and without
	subterranean stolons, leaves < 5 mm long. <i>Bryum</i> s.l.
_	(incl. Imbribryum, Ptychostomum)
2	Leaf apex narrowly pointed; costa ceasing below apex; leaf
	rosette consisting of (11) 16–21 (27) leaves; costa with 1–2
	(3) layers of dorsal stereids covered by 2–3 layers of wider
	superficial dorsal cells; margins weakly recurved; plants to 5

	cm tall, dark green, with leaves 9–10 mm long and patent whether wet or dry; usually on shady acid humus
-	Rhodobryum roseum (r) Leaf apex broadly pointed; costa excurrent; leaf rosette consisting of (18) 21–35 (52) leaves; costa with (3) 4–7 layers of dorsal stereids, superficial dorsal cells unistratose to irregularly bistratose; margins strongly recurved to revolute; plants to 3 cm high with leaves 5–6 mm long and incurved when dry; usually on base-rich soil over limestone.
3	Plants whitish-green or silvery; shoots julaceous, leaves closely imbricate when moist and when dry, concave, broadly ovate to ovate
-	Plants not conspicuously whitish-green or silvery; shoots not or rarely julaceous
4	Capsule gibbous on curved seta, with oblique mouth and long neck; leaves concave, imbricate; lamina cells 14–25 µm wide; plants in dense tufts with densely julaceous shoots, whitish or pale green above, usually vinaceous below; in crevices of basic rocks, often near waterfalls
-	Capsules symmetrical or slightly asymmetrical; shoots julaceous, but not vinaceous below; lamina cells 8–16 µm wide
5	Costa excurrent as short, stout apiculus, 60–100 µm wide at leaf base; upper leaf cells green (in young leaves); plants short and bud-like, julaceous; capsules elongate-ovate, light-brown when ripe; seta 2–4 cm long; spores 16–20 µm; bulbils in leaf axils lacking
_	Ptychostomum funkii (Bryum funckii) (rr) Costa mostly ending distinctly below leaf apex, rarely percurrent or excurrent, less wide; upper leaf cells hyaline, without chlorophyll; plants slender, thin, often silvery white; capsules small, ovate or obovate to ellipsoidal, dark red when ripe, getting blackish later; seta shorter, to 2 cm long; spores 8–14 μm; plants often with bulbils in leaf axils
6	Bulbils present in leaf axils, either foliate or with rudimentary leaf primordia. (<i>Bryum dichotomum</i> group)7
-	Bulbils in leaf axils lacking 10

7	Bulbils < 200 μm long, several per leaf axil, leaf primordia either rudimentary or only in uppermost part (< 1/3) of bulbil
-	Bulbils > 200 μm long, solitary or several per leaf axil, leaf primordia in upper third, half or two thirds of bulbil9
8	Bulbils brilliant yellow, usually <i>ca</i> 5 per leaf axil; leaf primordia rudimentary
-	Bulbils not brilliant yellow, often 20–30 per leaf axil; leaf primordia peg-like, incurved or erect.
	Bryum gemmiferum (r)
9	Bulbils solitary (rarely 2) in leaf axils; leaf primordia acuminate, erect, not incurved, occurring in upper half or two thirds of bulbil; bulbils up to 400 µm long (rarely more)
_	Bulbils several per leaf axil; leaf primordia rounded to
	obtuse, incurved, occurring in upper third of bulbil
	Bryum barnesii (r)
10 (6)	Filiform gemmae present in leaf axils
	Filiform gemmae lacking13
11	Leaves mostly abruptly narrowed to filiform acumen; leaves
	spirally twisted when dry; margin plane or recurved in lower part of leaf only
-	Leaves gradually narrowed to short acumen, flexuose, but not spirally twisted when dry; margin broadly or narrowly
	recurved from base to near apex
12	Leaf base red, contrasting green part of leaf (discolorous);
	margin with distinct border of 3–5 (10) rows of narrow,
	incrassate cells, border distinctly decurrent along stem;
	usually ± robust plants with conspicuous tomentum of red
	rhizoids, to > 4 cm tall (up to 10 cm)
	Ptychostomum pseudotriquetrum (incl. var. bimum)
	(Bryum pseudotriquetrum) (w)
_	Leaf base not contrasting rest of leaf, concolorous (either
	green or if red then whole leaf red); marginal border
	distinct, but narrower, to 3 rows, not decurrent; less robust
	plants, not tomentose
13 (10)	Leaves spathulate or ovate, widest at or above middle,
	suddenly contracted into long, filiform point; cells at leaf

-	base reddish, discolorous; lamina cells ± 20 µm wide14 Leaves widest below middle, more gradually narrowed to short or long point; cells at leaf base concolorous or discolorous
14	Leaves not spirally twisted when dry, closely imbricate and evenly arranged along ± julaceous stem; border often indistinct, 1–2 (3) cells wide; capsules very rare
-	Leaves spirally twisted when dry; margin with distinct border 2–5 cells wide; capsules not infrequent, to 5 mm long
15	Leaves not decurrent, margins not recurved, except in lower part; stems julaceous; rhizoids coarsely papillose, papillae tall and to 5 µm wide; lamina cells 15–20 µm wide
-	Leaves slightly decurrent, margins often recurved to apex; plants not julaceous; rhizoids less coarsely, but densely papillose; lamina cells 16–23 (30) µm wide
16	Plants dioicous; capsules brown when ripe, cernuous
-	Plants synoicous; capsules dark red when ripe, cernuous to pendulous
17 (13)	Robust plants, often with metallic sheen and reddish colour, stem evenly foliated, leaves stiff, densely imbricate when dry; costa ending below leaf apex or percurrent to shortly excurrent; marginal border indistinct
- 18	Characters different
-	Imbribryum alpinum (Bryum alpinum) (w) Plants not glossy, not red; mid-leaf cells 10–18 (28) μm wide, not incrassate; costa stout, excurrent into short point; rhizoidal tubers absent, but fragile shoots sometimes developed in leaf axils
	•

19	Plants with rhizoidal tubers
	(Note: several other species occasionally or regularly also produce
	rhizoidal tubers, e.g. <i>Imbribryum alpinum, Ptychostomum imbricatulum,</i>
	Ptychostomum capillare, Ptychostomum elegans, Ptychostomum moravicum, Ptychostomum torquescens, Bryum bicolor, Bryum barnesii,
	Bryum gemmiferum. These are not keyed out here!)
-	Plants lacking rhizoidal tubers (but compare Ptychostomum
	<i>imbricatulum</i> sometimes with rhizoidal tubers)26
20	Fully developed rhizoidal tubers small, < 100 (115) μm21
-	Fully developed rhizoidal tubers > 120 μm22
21	Rhizoids pale violet, finely papillose, tubers yellowish to
	orange or reddish-brown, 70-80 μm, cells not to slightly
	protuberant
_	Rhizoids pale yellowish brown, tubers bright crimson to
	reddish-brown, to 100 µm long, irregularly spherical, cells
	distinctly protuberant
22	Rhizoids and tubers yellow to orange-yellow; costa
	becoming dark purple with age
_	Tubers red; rhizoids not yellow
23	Rhizoids usually deep violet (old rhizoids dark red and
	coarsely papillose, young rhizoids pale reddish and only
	weakly papillose); tubers red to reddish-brown or orange, at
	the end of long rhizoids, never axillary, to 200 µm, cells not
	to slightly protuberant
_	Rhizoids paler, not violet
24	Leaves with distinct border of 2-3 (4) rows of elongate,
	narrow cells; median lamina cells 15-20 μm wide; tubers
	glossy, bright crimson red, on short rhizoids at the stem
	base and frequently also in leaf axils above the soil, opaque
	in transmitted light, (130) 180-260 µm in diameter, tuber
	cells 30-35 (45) µm in diameter, strongly protuberant, with
	2-3 µm thick walls as seen in profile; plant of slightly acidic
	to highly basic habitats
-	Leaves not or scarcely bordered; lamina cells 10-16 μm
	wide; tuber cells not protuberant; tubers never axillary25
25	Plants growing in dense, nearly cushion-like yellowish to
	brownish-green or green turfs, with numerous basal
	rhizoids forming a + dense felt costa stout in unner leaves

	excurrent in long, denticulate point; basal lamina cells beside the costa quadrate or shortly rectangular; mid-leaf
	cells 10–12 µm wide, incrassate; rhizoidal tubers 120–180
	(220) µm in diameter, pale brownish to bright red, tuber
	cells usually $< 45 \mu m$ long, not protuberant; plants of dry-
	warm calcareous habitats
-	Plants growing in lax turf, mostly reddish-green, without
	conspicuous rhizoid felt; costa in upper leaves only shortly
	excurrent; basal lamina cells beside the costa rectangular;
	mid-leaf cells $10\text{-}14$ (16) μm wide, slightly incrassate or
	thin-walled; rhizoidal tubers ± spherical, (180) 190–260
	(330) μm, red, tuber cells usually not protuberant, (18) 24-
	55 (65) μm long; plants growing on base-rich non-
	calcareous soil
	Imbribryum subapiculatum (Bryum subapiculatum) (w)
26 (19)	Plants growing in tufts usually > 4 cm tall, leaves often
	strongly decurrent, plants often with dense tomentum of
	reddish rhizoids
-	Plants smaller, leaves usually not or scarcely decurrent29
27	Leaf base concolorous, not contrasting with upper lamina;
	border often bistratose; lamina cells 16–24 μm wide; rhizoid
	tomentum scarce or lacking; leaves conspicuously longly
	and broadly decurrent; soft plants to 10 cm tall, often
	pinkish to flesh-coloured
_	Leaf base discolorous, reddish, contrasting with upper green
	lamina; border unistratose, conspicuous; laminal cells 12–14
20	μm wide; rhizoid tomentum often conspicuous
28	Plants dioicous; sporophytes rare; spores usually 12–18 μm.
	Ptychostomum pseudotriquetrum var. pseudotriquetrum
	(Bryum pseudotriquetrum var. pseudotriquetrum) (w)
-	Plants synoicous, nearly always with sporophytes; spores
	usually 15–25 μm.
	Ptychostomum pseudotriquetrum var. bimum
20(2()	(Bryum pseudotriquetrum var. bimum, Bryum bimum) (rr)
29 (26)	Leaf base not contrasting upper lamina in colour,
	concolorous, plants often reddish to pinkish; marginal
	border often partially bistratose, sometimes inconspicuous;
	capsule narrow-mouthed, lid with ± acute mamilla
-	Leaf base red, in contrast with upper green lamina,

	variegated red; border unistratose, conspicuous or indistinct
	to lacking; lid sharply or obtusely pointed
30	Endostome cilia ± long, nodose or with short appendages;
	spores (15) 18–24 (26) μm; plants dioicous; often with
	wine-red to pinkish colour
-	Endostome cilia short to rudimentary, rarely longer and
	nodose; spores > 28 μm; plants autoicous; seta usually long;
	leaf border yellowish; plants green or sometimes pale
0.4	reddish 32
31	Capsule elongate pyriform, with curved neck, not
	conspicuously contracted below mouth when dry and
	empty; marginal border conspicuous, to 3 cells wide, often
	bistratose (in well developed plants)
	Ptychostomum pallens (Bryum pallens) (r)
-	Capsule short, symmetric, turbinate when dry and empty,
	with conspicuous contraction below mouth; marginal
	border inconspicuous, often only 1-2 cells wide
22	
32	Exostome teeth usually with oblique cross-walls connecting
	lamellae, exostome partially united to endostome; spores usually > 40 µm; endostome segments with narrow, slit-like
	perforations; border less conspicuous, only locally
	bistratose; plants sometimes pale reddish
_	Exostome teeth without cross-walls between lamellae,
	endostome free; spores smaller, 28 (35) µm; endostome
	segments with oval perforations; leaf margin with yellowish,
	conspicuous regularly bistratose border; seta to 5 cm long;
	plants green
33 (29)	Leaf margin without border or border indistinct34
_	Leaf margin with distinct border36
34	Capsule with narrow mouth, elongate pyriform mostly
	slightly asymmetric, gibbous, red-brown to blackish when
	ripe; cilia of variable length, nodose, not appendiculate;
	plants synoicous; spores 18–25 μm
	Ptychostomum intermedium (Bryum intermedium) (rr)
-	Capsule with wide mouth, cilia appendiculate, or plants
	without capsules, dioicous

35	Plants small, 0.5–1 cm tall, whitish-green, julaceous; leaves broadly ovate, strongly concave, imbricate, crowded at bud-
	like apex; costa excurrent as short, but stout apiculus (of <i>ca</i>
	10% of total leaf length or less), 60-100 μm wide at leaf
	base; lamina cells thin-walled, lax, $20-35 \times 15 \mu m$; marginal
	border lacking (or consisting of a single row of incrassate,
	hardly narrowed cells); capsule turbinate (strongly
	contracted below mouth) when dry and empty
-	Plants not bud-like, taller, 0.5-2 cm; leaves ovate to ovate-
	lanceolate, not strongly concave, gradually narrowed to long
	point formed by excurrent costa (of ca 20-30% of total leaf
	length), entire or weakly denticulate; lamina cells longer,
	40–70 μm long; leaves enlarged and crowded towards shoot
	apex; leaf margin recurved to revolute from base to near
	apex; margin with \pm indistinct border formed by 2-3 (4)
	rows of elongate, narrow cells; plants dioicous, but often
	with sporophytes; capsule wide-mouthed, pendulous; cilia
	long appendiculate; spores 10–12 μm
36 (33)	Plants without sporophytes
	plants cannot be identified without ripe capsules
- 0 -	Plants with ripe capsules
37	Cilia short to rudimentary; spores large, usually > 20 µm
	(18–50 µm); plants usually synoicous; capsule narrow-
	mouthed 38
-	Cilia long, nodose or appendiculate; spores small, usually <
	20 μm (10–24 μm); plants dioicous, synoicous, autoicous or
38	polyoicous; capsule mouth wide or narrow
30	Exostome teeth with oblique cross-walls between lamellae; endostome attached to exostome; lid with sharp point; costa
	longly excurrent
	Note: Compare also <i>P. warneum</i> .
_	Exostome teeth usually without cross-walls; endostome
	free; capsule obovate to pyriform; lid flat, mamilla indistinct;
	leaves ovate-lanceolate, upper leaves longly acuminate;
	costa excurrent into long, smooth or slightly denticulate
	cuspidate point; exostome teeth at tip with thickened cell
	wall remnants

	(Bryum archangelicum, B. imbricatum) (rr)
39	Spores 15–25 μm; capsule mouth wide or narrow40
-	Spores (at least majority) not exceding 16 µm; capsule largemouthed41
40	Capsules ± gibbous, elongate pyriform, mostly somewhat curved and asymmetric, narrow-mouthed, cernuous to pendulous; leaf margin with indistinct border or 2–3 rows of narrow, elongate cells; plants synoicous
-	Capsules cylindrical to clavate pyriform or elongate pyriform, not curved, symmetric, large-mouthed, sometimes inclined; leaf margin with distinct border of 2–6 rows of elongate, narrow incrassate cells; plants autoicous or polyoicous
41	Plants synoicous; red leaf base distinct, leaf margin with distinct border of narrow, elongate, incrassate cells
-	Plants dioicous; leaf base often concolorous to indistinctly red; leaf margin with few rows of narrow, elongate cells forming indistinct border
	t o species of <i>Pohlia</i> ences: Erzberger (2005), Guerra in Guerra <i>et al.</i> (2010)
1	Plants rarely with sporophytes; usually with numerous axillary gemmae (bulbils) in leaf axils in upper part of stem, rarely bulbils solitary (in <i>P. andalusica</i>); lamina cells 8–10 times as long as wide; costa reaching leaf apex; leaf margin weakly denticulate at apex. (<i>Pohlia annotina</i> agg.)
2	Plants without axillary bulbils
-	plants dull when dry

3	Leaf primordia distinctly laminate in form; bulbils (including primordia) usually narrowed towards the apex and the base in outline, not larger than 600 μm, yellowish-green when young, red-brown when mature; leaf primordia 3–5 (8), laminate, erect, comprising <i>ca</i> 1/2 of total bulbil length, inserted at the same level at flattened bulbil apex; plants glossy, leaves straight, appressed to erect when dry
	Pohlia andalusica (rr)
-	Leaf primordia peg-like, rarely becoming laminate on very large ovate bulbils; bulbils green, yellow, orange, red according to age; plants dull or glossy4
4	Plants glossy when dry; bulbils mostly 150–300 (450) µm
-	long and to 60 µm wide, rather uniform in shape, oblong-
	linear to linear-vermicular; leaf primordia 1–2, reaching 1/4
	of total bulbil length
	Note: The bulbils of this plant differ from those of <i>P. annotina</i> , which may
	be similar, by the following characters: body of the bulbil 1–2 cells wide,
	with 1–2 primordia consisting of 1–2 cells (<i>P. annotina</i> : bulbil body opaque, more than 2 cells wide, with 2–3 (4) leaf primordia mostly
	consisting of more than 2 cells. See photographs in Guerra et al. (2010), p.
	192.
-	Plants dull when dry; bulbils extremely variable in size and
	shape, obovate, obconic to elongate and narrowly turbinate,
	occasionally bulbiform, rarely vermicular, different forms
	often present on a single stem, mostly more than 80 μm wide at the broadest point and 150–300 (550) μm long,
	opaque, with (2) 3–4 (5) peg-like, erect primordia
	(occasionally becoming laminate with age); leaves patent
	when dry; leaf cells uniformly wide from costa to margin
	Pohlia annotina (r)
5 (1)	Capsules longly cylindric, with neck as long as or longer than
- (-)	urn, horizontal; leaves narrowly lanceolate, margins
	recurved, denticulate; costa excurrent
	Pohlia elongata (n.s.)
_	Capsules shortly cylindric to ovoid, with shorter neck,
	drooping, pendulous6
6	Lamina cells of upper leaves to 5-6 times as long as wide,
	mostly > 16 μm wide; vegetative leaves short, only 2–4 times
	as long as wide; stems and leaf base often reddish7
-	Lamina cells of upper leaves longer, 6–14 μm wide; leaves
	mostly > 4 times as long as wide; leaf base reddish or not8

7	Plants whitish green, with waxy water-repellent surface; leaves predominantly ovate (observe older stems), ± distinctly decurrent; usually 1–3 cm high plants; rarely with sporophytes
-	Plants brownish or pale green, not waxy, easily wetted; leaves lanceolate to ovate-lanceolate, hardly decurrent; plants only to 5 mm tall; occasionally with sporophytes
8	Plants with rhizoidal gemmae, usually sterile, rarely > 1 cm tall9
_	Plants without rhizoidal gemmae, usually > 1 cm tall10
9	Gemmae brownish, spherical to pyriform, with smooth surface, not knobbly, $75-150 \times 70-90$ µm; plants very small, only to 5 mm high; lower leaves lanceolate with costa ending below leaf apex, upper leaves narrowly lanceolate with costa excurrent; lamina cells very narrow and elongate, $70-120 \times 9-12$ (15) µm
-	Gemmae yellowish, rounded, with protuberant cells giving a knobbly appearance, $5070 \times 4050 \mu m$; leaf apices more strongly denticulate and lamina cells narrower and longer than in preceding species, $70180 \times 610 \mu m$, wider in lower leaves
10	Lamina cells very long and narrow, about 10 times as long as wide, 6–10 μm wide and up to 150 μm long; plants conspicuously glaucous green with metallic sheen; leaf base, stem and costa reddish; capsules cernuous, ellipsoid, neck half as long as urn, pale brown
-	Lamina cells shorter and wider, only 5–7 times as long as wide, 10–12 µm wide and up to 70 µm long; capsule yellowish brown, elongate pyriform to elliptical; leaves erect, lanceolate, denticulate at apex
11	Plants paroicous, spores $2025~\mu m$ in diameter; habitat various, also in wetlands
	smooth: growing exclusively in hogs Pohlia sphaanicola

12 -	Note: doubtfully recorded. Plants dark green		
Rhiz	to species of <i>Mnium</i> (1; 4–7), <i>Plagiomnium</i> (8–13) omnium (2) ences: Koponen in Nyholm (1993), Sauer in Nebel and Philippi (2001)		
1	Leaves unbordered, bluntly toothed down to mid-leaf		
-	Leaves with a border of narrow elongate cells, often multistratose2		
2	Leaf margins entire, leaves roundish oval; stems densely tomentose in lower part		
3	Leaf margins toothed 3 Teeth of leaf margins usually in pairs (geminate); sterile shoots erect, leaves spirally arranged. <i>Mnium</i> pp4		
-	Teeth of leaf margins simple; sterile shoots procumbent or arcuate, with complanate leaves. <i>Plagiomnium</i> 8		
4 -	Lamina cells 13–17 µm wide		
5	Leaves <i>ca</i> twice as long as wide, broadly lanceolate to obovate; lamina cells without collenchymatous thickenings, not arranged in distinct rows; stems with terminal rosettes; often more than 1 seta per perichaetium; synoicous; leaves flexuose when dry, nearly smooth and only very slightly undulate when moist; dorsal surface of costa usually without teeth, rarely with few teeth; peristome (exostome) dark red-brown		
-	Leaves at least 3 times as long as wide, lanceolate; lamina cells with or without collenchymatous thickenings; leaves equal in size along stem, not forming rosette; only 1 seta per perichaetium; synoicous or dioicous; peristome (exostome) yellowish, greenish-yellow or brownish to golden-brown, not dark red-brown		
6	Dorsal surface of costa with teeth; dioicous7		

-	Dorsal surface of costa without teeth (a few sometimes
	present in upper leaves); costa reaching into ± cuspidate leaf apex; cell corners with distinct thickenings; marginal teeth
	small and blunt; stems and leaf base often brownish;
	·
7	synoicous
7	Leaf base not decurrent (or decurrent leaf bases not easily
	seen); costa usually ending below leaf apex (except
	sometimes in the uppermost leaves); lamina cells without
	collenchymatous thickenings
_	Leaf base distinctly decurrent; costa ending in acute apex; lamina cells with collenchymatous thickenings; cells at costa
	much larger than cells at leaf margin; numerous sharp teeth
	usually present on dorsal side of costa of all well-developed
	leaves; marginal teeth sharp
8 (3)	Leaves narrowly lingulate (except in immature plants),
0(3)	conspicuously decurrent; transversely undulate when wet
	and dry; leaf cells 10–16 µm wide
	Plagiomnium undulatum (c)
_	Leaves orbicular to oval or ovate, decurrent or not, not
_	transversely undulate; leaf cells larger9
9	Leaves serrate down to mid-leaf
,	Plagiomnium cuspidatum (c)
_	Leaves serrate down to base 10
10	Leaf base decurrent along stem
_	Leaf base not decurrent
11	Synoicous, often with sporophytes; lamina cells
	isodiametric, with thickenings <i>Plagiomnium medium</i> (n.s.)
_	Dioicous; lamina cells longer than wide, without thickenings
12	Leaves narrowly and often only shortly decurrent; sterile
	shoots prostrate; lamina cells 1.5–2 times as long as wide;
	costa weak, often ending below leaf apex; at least some teeth
	nearly perpendicular to margin, formed by 1–3 (4) cells
	Plagiomnium affine (c)
_	Leaves longly and broadly decurrent; sterile shoots arcuate;
	lamina cells twice as long as wide; costa strong apically,
	percurrent to excurrent; marginal teeth directed forward
	and formed by 1–2 cells
13	Synoicous; lid longly rostrate; lamina cells not porose,
	isodiametric to shortly elongate 35–50 x 20–32 um with

	trigones (collenchymatous thickenings); marginal teeth blunt
-	Dioicous; lid conical; lamina cells porose, elongate, 40–77 ×
	22–37 μm; marginal teeth sharp
	Plagiomnium ellipticum (r)
Key	to species of Orthotrichaceae
Refe	rences: Schäfer-Verwimp in Nebel and Philippi (2001), Guerra <i>et al.</i> (2014) arrós <i>et al.</i> (2016), Blockeel (2017)
Codo	onoblepharon (2), Lewinskya (15–17), Nyholmiella (13), Orthotrichum (9–10 25), Pulvigera (12), Ulota (4–8), Zygodon (2)
1	Plants with or without sporophytes, often with spindle-shaped gemmae having exclusively transverse walls, in leaf axils; if plants fertile then calyptra cucullate; lamina cells papillose or smooth
-	Plants usually with sporophytes, or if sterile then gemmae occurring on leaf lamina or at leaf apex; calyptra large and conspicuous, mitriform or campanulate, often hairy, leaf cells papillose
2	Upper and median lamina cells smooth; autoicous, capsules frequent, with double peristome; gemmae very sparse, with pale walls; costa strong, 70–85 (100) µm wide in lower third of leaf, ending below apex or percurrent, sometimes slightly excurrent; leaves erect-spreading; rare blackish-green plants to 2 cm high, on trees (<i>Fagus, Abies</i>) particularly in knotholes and seapage areas on the trunk, in Hungary around water-filled knotholes on <i>Quercus cerris</i> , often together with <i>Anacamptodon splachnoides</i>
-	Upper and median lamina cells papillose; dioicous, capsules rare, gymnostomous (without peristome); gemmae generally abundant, ovoid to ellipsoid, consisting of 3–6 cells with greenish to brownish walls, without longitudinal walls; costa weak, 30–40 (50) μm wide in lower third of leaf, ending below apex, rarely percurrent or slightly excurrent; leaves erect-appressed, ± flexuose, usually curved and homomallous when dry; green plants, brownish below, on the bark of living trees, also saxicolous
	Zygodon rupestris (r)

3	Marginal cells at leaf base hyaline, median basal cells incrassate and usually linear; leaves usually crispate when
	dry (except <i>Ulota coarctata</i> and <i>U. hutchinsiae</i>); seta usually
	longer than capsule, and capsule therefore exserted (far)
	above leaves; calyptra conspicuosly hairy. <i>Ulota</i> 4
-	Marginal cells at leaf base not forming hyaline border,
	median basal cells rectangular, not incrassate; leaves not
	crispate when dry (but somewhat curled in some species not
	yet recorded in Hungary); seta usually not much longer than
	capsule, capsule therefore often immersed, emergent or
	shortly exserted above leaves (more conspicuously exserted
	in Orthotrichum anomalum and Lewinskya speciosa);
	calyptra hairy or not. <i>Orthotrichum</i> (s.l., including
4	Lewinskya, Pulvigera, Nyholmiella)9 Leaves straight and appressed when dry, in appearance
4	much like <i>Orthotrichum</i>
_	Leaves curled or crisped when dry
5	Capsule pale, elongate pyriform with contracted mouth and
U	long neck, smooth and lacking striae when empty; on bark of
	deciduous trees
_	Capsule ellipsoid to narrowly cylindrical, not contracted at
	mouth, sulcate when empty; on siliceous rocks
6	Capsules spindle-shaped (fusiform) when empty, not
	contracted below mouth; sum of capsule length + seta length
	ca 5–8 mm; endostomial segments 8, linear, fragile, mostly
	uniseriate, ornamentation obliquely striate at base; leaves
	scarcely crisped when dry (each leaf is more strongly bent at
	its base than towards the tip, i.e. the radius of curvature
	increases from base to apex)
-	Capsules ± contracted below mouth (urceolate) when dry and empty; sum of capsule length + seta length <i>ca</i> 3–5 mm;
	endostomial segments fragile or persistent, not striate at
	base; leaves strongly crisped or not when dry. <i>Ulota crispa</i>
	s.l. (w)
7	Capsules when dry and empty conspicuously constricted
,	below mouth, ribs very close to each other in upper half of
	urn (intercostal spaces can hardly be seen); exothecial
	bands 4–6 cells wide, reaching to mouth or nearly so;
	exostomial teeth pairs not easily splitting, usually bordered

	tooth; endostomial segments 8, strong, persistent, shining,
	broad at base; leaf base wide, strongly concave, usually abruptly narrowing towards the leaf lamina; basal marginal
	leaf cells forming a broad band (7–15 cell rows) along the
	leaf base; leaves usually strongly crisped (with radius of
	curvature ± constant from leaf base to apex, compare
	however <i>U. bruchii</i>); operculum without differentiated basal
	ring
_	Capsules when dry and empty only slightly or not
	constricted below mouth, ribs separated by ± broad furrows
	in upper half of urn; exothecial bands 2-4 (5) cells wide,
	visibly separated from the mouth by a ring of small thin-
	walled cells in 2–6 rows; pairs of exostome teeth partially
	splitting in empty capsules, teeth not bordered by a hyaline
	halo; operculum with or without orange basal ring8
8	Endostome segments incurved when dry, uniseriate with
	incrassate and prominent transverse wall remnants,
	persistent, hardly fragile; all the cells of the exothecial bands
	hyaline with pale yellow incrassate lateral walls; leaves
	markedly crisped when dry, abruptly narrowing from a
	concave base; operculum without differentiated basal ring
	Ulota intermedia (r)
-	Endostome segments variably bent when dry, uniseriate or
	irregularly biseriate with thin transverse wall remnants,
	fragile, easily lost; cells of exothecial bands evenly pale
	yellow, at least in the two central rows; leaves slightly or
	moderately crisped when dry, gradually narrowing from a
	plane to slightly concave base; operculum usually with an
0(0)	orange to reddish basal ring
9 (3)	Leaves ending in hyaline point; capsule pale brown, smooth
	to weakly striate, almost immersed in leaves; peristome
	teeth separating at maturity. <i>Orthotrichum diaphanum</i> (cc)
-	Leaves without hyaline tip; peristome teeth separating or
10	not 10
10	Upper lamina cells large, 16–24 µm in diameter; leaves ovate-lingulate to elliptical with rounded apex (sometimes
	apiculate); plants 0.5–1 cm high, sometimes with leaf-borne
	gemmae; usually growing on wood or stones in the flood
	zone of rivers
	20110 01 11, 010

	Note: found only once in an artificial habitat in the Duna-Tisza-interfluve; see Erzberger and Papp (2000).
-	Upper lamina cells smaller, 10-18 μm in diameter; leaves
	ovate to lanceolate with acute to obtuse apex, sometimes
	apiculate11
11	Plants dioicous, usually without capsules; margins plane to
	incurved; leaves sometimes covered with conspicuous
	brown gemmae; plants usually corticolous
-	Plants autoicous, almost always with capsules; gemmae, if
	present, green and inconspicuous; leaf margins recurved;
	plants corticolous or saxicolous
12	Leaves lanceolate, densely covered with brown filamentous
	gemmae; leaf margins plane; large plants, 3-4 cm tall or
	more
-	Leaves ovate-lingulate, with rounded apex, with few to many
	small greenish gemmae; margins plane to incurved; small
	plants. <i>Nyholmiella</i> 13
13	Leaf margin plane; upper lamina cells with 1 papilla per cell
	(rarely 2); gemmae predominantly on ventral side of leaf
	lamina; capsule with double peristome
	Nyholmiella obtusifolia (Orthotrichum obtusifolium) (c)
_	Leaf margin incurved from base to somewhat cucullate apex;
	upper lamina cells with (1) 2–3 papillae per cell; gemmae on
	both sides of leaf lamina; capsule without peristome
	Nyholmiella gymnostoma
	(Orthotrichum gymnostomum) (n.s.)
14 (11)	Capsules with superficial stomata (phaneropore).
()	Lewinskya 15
_	Capsules with immersed stomata (cryptopore).
	Orthotrichum19
15	Outer peristome teeth erect or spreading when dry, coarsely
	papillose, sometimes with preperistome; capsules with 8 ±
	conspicuous striae, immersed to emergent, ovoid-obloid;
	outer peristome teeth in 8 pairs soon separating; calyptra
	strongly hairy, the hairs rising above apex of calyptra;
	lamina occasionally bistratose above and towards margins;
	leaf cells with simple papillae; 1.5–5 cm tall robust plants,
	commonly on (siliceous) rock but sometimes also on
	deciduous trees
	Lewinskya rupestris (Orthotrichum rupestre) (r)

Note: *Ulota hutchinsiae* might key out here, because it has superficial stomata, grows on siliceous rocks and has erect to spreading peristome teeth, sometimes with a preperistome, and a very hairy calyptra, and unlike other *Ulota* species, has the leaves not crisped, but straight and appressed when dry. However, its capsules are clearly exserted, strongly contracted and cylindrical when dry and empty, and careful examination of the coloured leaf base will reveal the characters typical of *Ulota* (hyaline marginal and incrassate median basal cells).

- Capsules pale, elongate-cylindrical, strongly sulcate when dry, hemi-emergent to shortly exserted; calyptra naked or with few short hairs; peristome teeth united in 8 pairs (or partly separating when old), regularly recurved when dry, touching surface of capsule throughout their length; inner peristome segments coarsely papillose (rarely nearly smooth); leaves mostly with a broad, short apex; mostly on trees, but also on rock.

Lewinskya affinis (Orthotrichum affine) (cc) Note: Lewinskya fastigiata was distinguished from L. affinis as variety already by Boros (1968), but neglected during the last decades. It has been resurrected recently as a good species (Vigalondo et al. 2020), but is missing in the most recent Hungarian checklist (Erzberger and Papp 2020) pending verification of specimens. It can be distinguished from L. affinis inter alia by capsules immersed or hemi-emergent, and having broad exothecial bands 4 cells wide near capsule mouth (often 6–8 cell rows below) vs. narrow bands 2–3 cells wide near capsule mouth (sometimes 4–6 below) in L. affinis.

- Capsule cylindrical with long neck, weakly sulcate in upper half or total length, emergent to clearly exserted, gradually narrowed to the seta, seta 0.75–3 mm long; peristome teeth

	united in 8 pairs (or partly separating when old); inner peristome segments 8, papillose, with slightly irregular margins; calyptra usually densely hairy; plants forming tufts to 3 cm tall; on deciduous trees, rarely on rock
18	Seta (1) 1.5–2.5 (3) mm long, as long as or longer than the
	urn, capsule clearly exserted; capsule wall smooth or with
	short striae in upper half only; ripe calyptra conical or
	fusiform, with golden or colourless hairs
	Lewinskya speciosa (Orthotrichum speciosum) (c) Note: When capsules are not strongly exserted, confusion with <i>L. affinis</i> might occur, especially when old capsules of <i>L. speciosa</i> sometimes are sulcate to near base. However, in a cushion of <i>L. speciosa</i> nearly always some smooth capsules will be present. Additional useful characters to separate <i>L. speciosa</i> from <i>L. affinis</i> : (i) strongly hairy calyptra (calyptra with only few or no hairs in <i>L. affinis</i>); (ii) peristome teeth arcuate, only the tips touching the capsule surface (regularly recurved in <i>L. affinis</i>); (iii) stomata confined to lower half of capsule (<i>L. affinis</i> : stomata not too frequent, scattered all over the capsule, some occasionally close to capsule
_	mouth). Seta 0.75–1 (1.3) mm long, as long as or less long than the
	urn, capsule half emergent; capsule wall with distinct striae
	in total length; ripe calyptra oblong-conical, with greenish-yellowish hairs
10(14)	(Orthotrichum speciosum var. brevisetum)
19 (14)	Outer peristome teeth erect or spreading when dry, often
	with preperistome (prostome); plants mostly on rocks20
-	Outer peristome teeth recurved when dry, touching surface of empty capsule; prostome lacking; plants mostly on trees
20	Vaginula with numerous golden-yellow hairs (also
20	numerous golden-yellow paraphyses between the
	antheridia); calyptra campanulate, straw-coloured, with
	(moderately) numerous yellow, papillose hairs; capsule
	wide-ovoid to shortly urn-shaped; inner peristome
	segments 8 or 16, well developed; capsule wall with 8
	pronounced striae, intermediate striae weak or lacking;
	plants densely tomentose, in tall lax cushions; on non-
	calcareous rocks
_	Vaginula naked or with a few hyaline hairs (very rarely with
	numerous hairs); inner peristome lacking; capsule with 16 ±
	equally strong striae (but intermediate striae often

	somewhat shorter); on base-rich rock21
21	Capsules exserted above leaves, elongate-cylindrical, only
	very slightly constricted below mouth when dry; calyptra
	long conical to campanulate, with few to numerous papillose
	hairs; seta 2-4 mm long, as long as or longer than capsule,
	reddish brown; outer peristome teeth striate horizontally
	below, vertically above
_	Capsules immersed or only shortly exserted, wide ovoid or
	urn-shaped when empty, slightly constricted below mouth, ±
	abruptly contracted into seta; seta shorter than the capsule,
	pale; outer peristome teeth striate, papillose-striate or
	papillose-reticulate; inner peristome segments usually
00(10)	lacking
22 (19)	Exothecial bands of capsule wall formed by 2 rows of well-
	differentiated yellowish cells (locally 3-4 cells wide);
	vaginula with many long hairs; spores 15–23 μm
	Orthotrichum patens (c)
	Note: Confusion with O. stramineum, the only other (Hungarian) corticole
	species with long vaginula hairs, can be avoided by taking into account the
	following characters: (i) spore size in just ripe capsules (<i>O. patens</i> : 16–20
	(24) μm, <i>O. stramineum</i> : mostly 12 μm); (ii) capsule shape (<i>O. patens</i> :
	wide ovoid, abruptly (often forming a 90° angle) contracted into seta, dry
	empty capsules not longitudinally contracted, O. stramineum: capsule
	more cylindrical, gradually narrowed to seta, dry empty capsule always
	strongly contracted); (iii) width and length of capsule striae (0. patens:
	only 2–3 cells wide, <i>O. stramineum</i> : 4–6 cells wide and mostly longer); (iv)
	endostome sculpture (<i>O. patens</i> : papillose, <i>O. stramineum</i> : smooth); (v) apical openings in exostome teeth (<i>O. patens</i> : mostly lacking, <i>O.</i>
	stramineum: present); (vi) papillosity of upper lamina cells (<i>O. patens</i> :
	cells with 1–4 (6) simple or forked papillae, <i>O. stramineum</i> : less papillose,
	with small, mostly unbranched papillae); (vii) time of spore ripening
	(most easily observed, when both species occur together) – is 1 month
	ahead in <i>O. patens</i> . A hairy vaginula is also observed in <i>O. urnigerum</i> , a
	saxicole species with an erect exostome.
_	Exothecial bands of capsule wall formed by 4 or more rows
	of well-differentiated cells; vaginula hairy or not; spores
	usually smaller (but 18–24 µm in <i>O. rogeri</i>)23
23	Vaginula with numerous long hairs (best seen near young
23	capsules); capsule elongate-cylindrical, longitudinally
	. ,
	contracted when dry; gradually narrowed into seta; empty
	capsule wall tough; capsule striae (3) 4–6 cells wide; inner
	peristome with 8 or 16 segments (the intermediate
	segments often consisting of few cells only); outer peristome

teeth with apical window-like openings, extremely rarely splitting into 16 single teeth; spores mostly 12 μm (10–15 μm); calyptra campanulate, with some smooth hairs, rarely naked, the tip conspicuously blackish far down.....

.....Orthotrichum stramineum (w)

......Orthotrichum rogeri (rr)

- Plants with monomorphic leaves, leaves of all shoots similar, independent of sex; endostome segments incurved when dry, smooth or papillose, without longitudinal striae...........25
- Upper leaves from ovate base lanceolate, widest at lower third, apex acute or obtuse, rarely rounded; capsule gradually narrowed to the seta, half emergent to almost exserted; neck and seta pale brown, contrasting with urn, with 8 striae 6–8 cells wide; stomata (mostly) 'pseudophaneropore', i.e. hardly covered by broadly rounded edges of surrounding cells; inner peristome with 8 or more often 16 segments (8 longer and 8 shorter, the latter sometimes reduced to 1–2 cells); calyptra short, to 1.4 mm long; gemmae unknown; exostome teeth yellowish, ornamented in upper part with papillae and lines......

.....Orthotrichum pallens (c)

Note: Orthotrichum moravicum is closely related to O. pallens, but differs in 16 inner peristome segments of \pm equal length (not 8 longer and 8 shorter as in O. pallens) and nearly as long as the exostome teeth; the endostome processes have conspicuous lateral appendages (Plášek $et\ al.$ 2009). O. moravicum was described from the Czech Republic and might be expected in Hungary.

 Upper leaves frequently ovate-lanceolate (widest shortly below middle) to oblong-lanceolate, acuminate or obtusely rounded (sometimes apiculate with 1–2 hyaline end cells); capsule immersed to half emergent, suddenly or gradually narrowed to seta; stomata covered by surrounding cells to a

	variable extent, mostly in a single capsule some stomata hardly covered, some covered half or more by irregularly protruding mamillae of surrounding cells; inner peristome mostly with 8 segments; exostome orange, ornamentation various
26	Capsule abruptly narrowed to the seta; exostome teeth in persistent pairs, ornamented with tall papillae (rarely some lines); endostome segments 8, with enlarged base, usually shorter than exostome teeth; gemmae very common
_	Capsule gradually narrowed to the seta; exostome teeth pairs splitting at tips, ornamented in upper part with low papillae and some fine lines; endostome segments 8 or 16, long and narrow, not noticeably widened at base, as long as exostome teeth, gemmae rare <i>Orthotrichum pumilum</i> (c)
Key	to species of Aulacomnium
-	Stems tomentose; gemmae rare; cells at leaf base pluristratose, brownish; plants to 10 cm tall
-	Stems not tomentose, nearly always with gemmae in stalked globose clusters at shoot apices; cells at leaf base unistratose; plants 1–3 cm high
	(1)
Key	to species of <i>Fontinalis</i>
- -	At least lower stem leaves keeled; stem and branch leaves arranged in 3 rows

Key to species of *Plagiothecium* References: Erzberger and Baráth (2017), Erzberger *et al.* (2020)

1	Leaves whitish green, 2–5 mm long, imbricate, strongly transversely undulate; decurrent alar cells with straight
	outer walls; on forest soil
-	Plants without this combination of characters2
2	Median lamina cells narrow, < 10 μm wide; small plants usually only 2–3 cm long3
-	Median lamina cells > 10 μm wide; medium-sized to large plants
3	Leaves 0.9–1 mm long and <i>ca</i> 0.35 mm wide, symmetric gradually acuminate to a long point, often with uniseriate gemmae and rhizoids at the tip; lamina cells 5 (8) µm wide
-	Leaves longer and wider (mostly > 0.5 mm wide) asymmetric, apex less longly drawn out, usually without gemmae (but gemmae frequent in leaf axils); lamina cells wider
4	Leaf apices plane; leaves asymmetric usually with one curved and one straight side, to 1.2 mm long, alar cells decurrent in 1–2 rows, not clearly delimited, not auriculate with straight outer walls, rectangular
-	Leaf apices curved downward towards substrate when moist; leaves asymmetric with two curved sides, > 1.2 mm long, alar cells forming a distinct group, decurrent in 2–4 (5) rows, mostly somewhat auriculate, often with weakly bulging outer walls, isodiametric or rectangular; capsule horizontal to inclined, often gibbous
	Plagiothecium curvifolium (w)
5	Decurrent alar cells inflated, oval to rounded, with bulging walls, sometimes only a few cells rounded and most cells rectangular or quadrate; lamina cells not wider than 16 µm
-	Alar cells not inflated, the decurrent cells rectangular, not rounded; lamina cells wider than 16 µm or not8

- Most alar cells strongly inflated, rounded, forming an oval group usually less long, often wider, decurrent in 3–4 (6) rows; leaves without rhizoid initials at apex; all leaves distinctly asymmetric; medium-sized plants. *Plagiothecium denticulatum*.
- 7 Lamina cells in ± distinct transverse rows, therefore leaves sometimes transversely undulate when moist; margins at apex often completely without denticulations; leaves 2.5–4 mm long, strongly complanate, strongly asymmetric, at least some with one straight and one curved side; decurrent alar cells in 4–6 (8) rows; plants of wet habitats......

- Lamina cells not in transverse rows, leaves not transversely undulate when moist; margins at apex often denticulate; leaves 1.5–2.5 (3) mm long, less strongly complanate, asymmetric, mostly with both sides curved; decurrent alar cells in 3–5 rows; plants of humid, but not particularly wet habitats.......*Plagiothecium denticulatum* var. *denticulatum* (*Plagiothecium denticulatum*) (w)

- Mid-leaf cells (65) 75–140 (155) \times 15–22 (25) μm (4–6 times as long as wide) in \pm transverse rows; apical end cell

rhomboidal; leaf apex usually with (few) rhizoid initials..... Note: Wolski and Krawczyk (2020) recently described a new species closely related to P. nemorale, Plagiothecium angusticellum, which they also identified in a collection by L. Vajda from Börzsöny Mts. P. angusticellum is missing in the latest Hungarian and European checklists (Erzberger and Papp 2020, Hodgetts et al. 2020), and the differentiation from *P. nemorale* does not seem straightforward. Key to species of Orthothecium Plants medium-sized to robust, often reddish, leaves 2-4 mm long, strongly plicate longitudinally..... **Orthothecium rufescens** (rr) Note: missing in Erzberger and Papp (2020), recently discovered by Németh and Schmotzer in a single site in Bükk Mts. Plants delicate, mostly yellowish green, rarely reddish, leaves mostly 1.7 mm (to 2 mm) long, without longitudinal plicae **Orthothecium intricatum** (r) Key to species of Fabronia Leaf margins with unicellular teeth........ *Fabronia ciliaris* (rr) Leaf margins with pluricellular teeth, appearing ciliate.....Fabronia pusilla (r) Key to species of *Palustriella* regularly complanately pinnately branched. Stems tomentose, paraphyllia abundant; stem leaves cordatetriangular; alar cells numerous, forming a triangular group ascending the margin *Palustriella commutata* (r) Stems irregularly or subpinnately branched, not tomentose,

Key to species of Campyliadelphus

- Stem leaves straight, sometimes curved, narrowly triangular lanceolate, about four times as long as wide, from ovate base gradually narrowed to long linear, ± channelled acumen; costa strong, reaching higher than 2/3 leaf length; leaf margin ± distinctly denticulate; wetland species......

Key to species of Campylium

- Leaves often abruptly narrowed into a long acumen, 1–2.3 mm long; creeping, often pinnately branched plants.....

(Campylium stellatum var. protensum) (r)

Key to species of *Drepanocladus*

- Costa weak, mostly extending not far above mid leaf; leaves straight or falcate; acumen channeled; alar cells incrassate, porose, often reaching costa, distinctly decurrent; leaves 2–3 mm long, gradually and longly pointed from broadly lanceolate base; stem leaf insertion slightly curved......
 -Drepanocladus polygamus (Campylium polygamum) (r)

-	Alar cells in relatively small groups reaching 40–60% distance from leaf margin to costa, the cell walls often
	incrassate; leaves often strongly falcate
3	Leaves lanceolate to linear-lanceolate, longly and finely
3	acuminate; costa reaching into apex4
-	Leaves broadly ovate-lanceolate, shortly acuminate,
	gradually narrowed from base to apex, concave; costa
	ending in leaf apex; plants robust, turgid, to 30 cm long,
	brownish yellow
4	Ratio of median leaf cell length (in µm) to leaf length (in
	mm) 18.0-24.5; alar cells often incrassate; leaves longly and
	finely pointed; often very robust plants, to 20 cm long; costa
	strong, (50) 70-100 µm wide at base, reaching to leaf apex
	Note: To compute the aforementioned ratio, examine 8–10 leaves from a
	homogeneous part of stem under the microscope, measure length of the longest and the shortest leaf and cell length of the longest and the shortest
	leaf cell, then compute mean values and use these for computing the
	ratios (Hedenäs and Bisang 2002).
-	Ratio of median leaf cell length (in μm) to leaf length (in
	mm) 23-36.5; alar cells thin-walled; costa relatively weak,
	mm) 23–36.5; alar cells thin-walled; costa relatively weak, 30–75 μm wide at base, ending far below leaf apex
	30–75 μm wide at base, ending far below leaf apex
-	30–75 µm wide at base, ending far below leaf apex
Hygro	30–75 µm wide at base, ending far below leaf apex
Hygro	30–75 µm wide at base, ending far below leaf apex
Hygro radic	30–75 µm wide at base, ending far below leaf apex
Hygro	30–75 µm wide at base, ending far below leaf apex
Hygro radic	30–75 µm wide at base, ending far below leaf apex
Hygreradic	30–75 µm wide at base, ending far below leaf apex
Hygro radic	30–75 μm wide at base, ending far below leaf apex
Hygreradic	To species of Amblystegium s.l. (incl. Amblystegium ale, Serpoleskea confervoides) Leaves without costa or costa short and double, not reaching mid-leaf 2 Leaves with single costa extending to or above mid-leaf 3 Capsule curved; perichaetial leaves without costa; on shaded calcareous rocks
Hygreradic	To species of Amblystegium s.l. (incl. Amblystegium pamblystegium, Pseudoamblystegium subtile, Pseudocampylium ale, Serpoleskea confervoides) Leaves without costa or costa short and double, not reaching mid-leaf
Hygreradic	To species of Amblystegium s.l. (incl. Amblystegium pamblystegium, Pseudoamblystegium subtile, Pseudocampylium ale, Serpoleskea confervoides) Leaves without costa or costa short and double, not reaching mid-leaf
Hygreradic	To species of Amblystegium s.l. (incl. Amblystegium ale, Serpoleskea confervoides) Leaves with single costa extending to or above mid-leaf
Hygreradic	To species of Amblystegium s.l. (incl. Amblystegium amblystegium, Pseudoamblystegium subtile, Pseudocampylium ale, Serpoleskea confervoides) Leaves without costa or costa short and double, not reaching mid-leaf
Hygreradic	To species of Amblystegium s.l. (incl. Amblystegium ale, Serpoleskea confervoides) Leaves without costa or costa short and double, not reaching mid-leaf

3	Costa reaching leaf apex, often bent in upper third4
-	Costa reaching mid-leaf, straight7
4	Costa very strong, mostly > 40 µm wide at leaf base, hardly narrower towards the tip, mostly reaching to leaf apex, often slightly bent in upper third; plants growing in or near water
-	Costa less strong, distinctly narrowed towards the tip, bent above or not; plants not aquatic but growing in moist or wet habitats6
5	Leaves narrowly rounded at apex; branch leaves widest at (1/4) 1/3–1/2 leaf length; stem little and irregularly branched; plants growing in lime-poor flowing water
-	Leaves distinctly acuminate at apex; branch leaves widest at 1/8–1/4 leaf length; stem often densely and pinnately branched; plants growing in base-rich or lime-rich flowing water
6	Leaf cells at mid leaf rhombic or hexagonal, 2–4 times as long as wide; leaves distinctly widened in lower third and contracted at leaf base, often narrowed ± abruptly towards leaf tip; costa always with a distinct bend
-	Leaf cells at mid leaf often elongate; leaves not conspicuously contracted at their base, gradually narrowed towards the tip; costa not to at most slightly bent
7	Slender (to medium-sized) plants, leaves to 1 mm long; variable with regard to length of costa and length of lamina cells (from 2–3 to 4–6 times as long as wide)
	robust var. <i>juratzkanum</i> with leaves denticulate at base) (cc)
_	Medium-sized to robust plants, leaves > 1 mm long8
8	Leaves decurrent along stem <i>Pseudocampylium radicale</i> (<i>Campylium radicale, Amblystegium radicale, A. saxatile</i>) (r)
-	Leaves not decurrent along stem, distantly arranged along stem; leaf margin entire; leaves distinctly contracted at leaf base

Key to species of Calliergon s.l. incl. Straminergon

- Plants almost unbranched; costa reaching 3/4 or slightly more of leaf length; stem leaves oblong, ± imbricate; alar cells in decurrent group ascending up margin; often with rhizoids at leaf apex.....
 -Straminergon stramineum (Calliergon stramineum) (rr)

- Alar cells very sharply differentiated, rarely reaching costa, in a strongly concave group; leaves shorter and broader; ca
 1.5 times as long as wide; plants pinnately branched with turgid main stems and short, thinner branches, green, yellowish or brownish; dioicous... Calliergon giganteum (rr)
 Note: Depauperate plants may approach C. cordifolium.

Key to species of Scorpidium

- Plants turgid; stem leaves ovate to ovate-orbicular, strongly concave, apex acute or shortly acuminate; costa rarely reaching mid-leaf, often short and double or lacking......
 - Scorpidium scorpioides (rr)
- Plants not turgid; stem leaves ovate-lanceolate, falcate, moderately concave, longly acuminate; costa reaching to 1/2-2/3 leaf length.

Key to species of Pseudoleskeella

Key to varieties of Abietinella abietina

References: Düll-Hermanns (1981) and Smith (2004)

Key to species of *Thuidium*

- Terminal cells of branch leaves with several apical papillae (often therefore appearing truncate or forked)2

-	Costa of stem leaves ending below leaf apex; apical leaf cells short; tips of stem leaves erect; cells of paraphyllia with a centrally placed papilla; stem tip ± straight, not hooked
3	Stem leaves with a long, fine acumen, the apex consisting of 3–4 uniseriate elongate cells (easily broken off!), perichaetial leaves without cilia.
-	Stem leaves more shortly pointed, apex not uniseriate; perichaetial leaves ciliate
Key	to species of Eurhynchium
-	Leaves 1.2–2 times as long as wide, patent; leaf apex narrow, margins forming an angle of 15–45°
-	Leaves 1.2–1.3 times as long as wide, appressed; leaf apex broadly pointed, margins forming an angle of 45–85°
Key	to species of Rhynchostegium
1	Mid-leaf cells lax, 10–16 μm wide, 3–5 times as long as wide; leaves subrotund
2	Mid-leaf cells 4–10 µm wide, 6–17 times as long as wide2 Leaves widely elliptical, concave, apex obtuse and apiculate; entire or denticulate only at apex; costa reaching mid-leaf; yellowish green to brownish, often silvery, glossy julaceous plants with dense leaves
-	Leaves ovate to elliptical, apex acute or acuminate; margin denticulate from base to apex3
3	Leaf apex acute; dark green plants with long straight branches and broadly ovate leaves denticulate all around; stem and branch leaves similar; basal cells weakly differentiated; aquatic or subaquatic plants in and by streams
_	Leaf apex acuminate 4

Key to species of Cirriphyllum

- Stem leaves more gradually narrowed to lanceolate acumen; costa ending in dorsal spine.......*Cirriphyllum crassinervium* (*Eurhynchium crassinervium*) (w)

Key to species of Oxyrrhynchium

- 1 Primary stems rhizomatous, subterranean, with scale-like leaves; lamina cells 4–6 μ m wide; leaf apex often twisted by 180°.....
 - ... **Oxyrrhynchium schleicheri** (Eurhynchium schleicheri) (w)
- 2 Stem leaves ovate to triangular lanceolate, gradually narrowed to apex, 1.5–2 mm long; branches with complanate, almost distichous leaves, very glossy; synoicous or autoicous, often with capsules; leaves longly decurrent, recurved at base; costa reaching into leaf acumen, occasionally ending in dorsal spine; stem leaf cells 60–90 μm long.
 - Oxyrrhynchium speciosum (Eurhynchium speciosum) (r) Note: Rhynchostegium riparioides is somewhat similar in habit, but distinguished by a smooth seta, if present.

Stem leaves rounded ovate heart-shaped, rather suddenly narrowed to apex, 1-1.2 mm long; plants decumbent, irregularly pinnate, to 10 cm long; leaves complanate or not; dioicous, rarely with capsules; costa reaching 1/2 to 3/4 leaf length, usually ending in conspicuous dorsal spine; stem leaf cells 40–60 µm long Oxvrrhynchium hians (Eurhynchium hians) (cc) Key to species of Rhynchostegiella Leaves narrow lanceolate or more often linear-lanceolate, 6-1 10 times as long as wide, longly acuminate; margins entire (or very finely denticulate near apex only); cells of leaf apex elongate; costa reaching into leaf apex; lamina cells 10-20 times as long as wide; frequently with sporophytes, seta smooth; plants forming glossy, silky yellowish green mats..... Leaves more broadly lanceolate. 3–5 times as long as wide. bluntly pointed or acute, denticulate along margins; cells of leaf apex short, 4:1; costa of variable length; seta rough......2 Mats dull dark green, costa reaching into leaf apex; leaf cells 2 5-7:1 *Rhynchostegiella teneriffae* (r) Mats glossy, light green; costa reaching shortly above midleaf; leaf apex narrowly lanceolate; leaf cells 8-10:1: on sunny or sheltered, warm sandstone and limestone rocks...... Note: the seta is smooth in **R. curviseta** var. **laeviseta** (W.E. Nicholson and Dixon) Podp. (rr) Key to species of *Homalothecium*

- 2 Plants irregularly branched, rarely ± pinnate, to 15 cm long;

-	stems usually loosely attached to substrate; branches straight; basal leaf cells porose, leaf margin irregularly finely denticulate or entire; capsules usually curved, endostome well developed; plants yellowish green
	to species of <i>Brachythecium</i> s.l. (<i>Brachytheciastrum</i> hythecium and <i>Sciuro-hypnum</i>)
1	Leaves with sharply set-off, long and narrow acumen (hair point); stem leaves ovate or narrowly ovate; transition between leaf lamina and acumen short, but not abrupt; lamina cells 6–12 µm wide
_	tommasinii) (w) Leaves more gradually narrowed upwards, or with broad
	apex
2	Costa long, ending at least 80% up the leaf3
-	Costa shorter, ending 50-70% up the leaf6
3	Alar groups reaching costa; costa mostly percurrent; autoicous, sporophytes frequent; operculum conical; seta ± rough above, smooth below
	<i>Sciuro-hypnum populeum</i> (<i>Brachythecium populeum</i>) (w) Alar groups not reaching costa, small, marginal, costa not
_	percurrent; seta rough4
4	Leaves strongly plicate, stem leaves triangular or narrowly triangular-ovate, ± gradually narrowed towards apex; medium-sized plants; dioicous, sporophytes rare; operculum conical
-	Leaves plane, abruptly narrowed into apical portion; medium-sized or small plants; operculum conical or rostrate

5	Medium-sized plants; stem leaves not decurrent, abruptly
	narrowed into short acumen on average 3 cells wide, often
	twisted 180°; costa very stout in the lower part of the leaf,
	distinctly narrowing above, ending shortly below leaf tip or
	reaching into it; stems prostrate, branches irregular,
	ascending; operculum rostrate
	Sciuro-hypnum flotowianum (Eurhynchium flotowianum,
	Cirriphyllum reichenbachianum) (w)
	Note: S. flotowianum resembles S. populeum in habit, but can be
	distinguished from that species by the costa not ending in leaf apex and
	leaf margins recurved only below.
-	Small plants; stem leaves distinctly decurrent, abruptly
	narrowed towards narrow apical portion that is at least 250
	μm long, not twisted; costa less stout; dry branches strongly
	curved; operculum conical Sciuro-hypnum reflexum
	(Brachythecium reflexum) (rr)
5 (2)	Alar groups reaching costa; plants of moist or periodically
	wet environments
	Sciuro-hypnum plumosum (Brachythecium plumosum)(rr)
_	Alar groups not reaching costa
7	Alar cells dilated, thin-walled and hyaline, forming a large,
	distinct, decurrent group; plants of moist or periodically wet
	environments
_	Alar cells not inflated, alar group hardly decurrent; plants of
	usually dry habitats (except <i>B. mildeanum</i>)8
8	Alar groups extending up along margin in a band or an ovate
U	group9
	Alar groups different
9	9 1
9	Dioicous, sporophytes rare; leaves erect and imbricate,
	therefore branches usually julaceous; margin of branch
	leaves entire or very finely denticulate; branch leaf costa
	smooth, rarely ending in an indistinct spine on abaxial side;
	seta smooth
-	Autoicous, sporophytes frequent; leaves erect to patent;
	margin of branch leaves distinctly to coarsely denticulate;
	branch leaf costa frequently ending in a distinct spine on
	abaxial side; seta at least partly rough
	Brachythecium campestre (r)

10	Leaves ± distinctly plicate; plants large to medium-sized; alar groups small, marginal, mainly consisting of quadrate or
-	rectangular cells; plants dioicous or autoicous
11	Median lamina cells 35–65 μm long; plants dioicous; margin of branch leaves denticulate to finely so; seta smooth or rough; costa ending in distinct or indistinct spine on abaxial side; leaves very strongly plicate, reminding of a <i>Homalothecium</i>
-	Median lamina cells 40–150 μm long; plants autoicous or dioicous; margin of branch leaves almost entire to coarsely denticulate; seta smooth; costa rarely or often ending in spine on abaxial side
12	Costa reaching 75–85 % up the leaf, very stout at leaf insertion; margin of branch leaves denticulate; seta rough (but sporophytes rare); median lamina cells 35–60 µm long; plants growing on base-rich, but not calcareous rocks
-	Costa reaching 50–75 % up the leaf; margin of branch leaves denticulate to finely so; seta smooth; median lamina cells 50–65 µm long; plants growing in dry calcareous habitats, on small stones at the ground, never on rock. **Brachythecium laetum** (rr)**
13	Dioicous; costa smooth or rarely ending in a small spine on abaxial side; margin of branch leaves finely denticulate to almost entire; median lamina cells 40–130 µm long; stem leaves elongate-triangular, near apex with a long and narrow, often twisted piliferous portion; plants growing in dry calcareous habitats
-	Autoicous; costa often ending in a distinct spine on abaxial side; margin of branch leaves distinctly to coarsely denticulate; median lamina cells 54–150 µm long; stem leaves triangular-ovate, near apex narrow but not piliferous; plants growing in base-poor to base-rich habitats
14 (10)	Stem leaves 0.35–0.6 (0.9) mm wide; plants small (to medium-sized); alar groups small, marginal, mainly consisting of quadrate to rectangular cells, distinctly delimited; branch leaf spine of costa and prorate cell ends of

	costa strong, rarely absent
-	Stem leaves wider; plants large; alar groups indistinctly delimited
15	Seta smooth; some lamina cells distinctly prorate
	Brachytheciastrum olympicum (rr)
-	Seta rough; lamina cells of stem leaves hardly prorate, of
	branch leaves prorate
	(Brachythecium velutinum) (cc)
16	Seta smooth; costal spine absent or rarely present and very
	weak
-	Seta rough or partly rough
17	Leaf margin entire or finely denticulate; alar cells regularly
	rectangular; median lamina cells of stem leaves $813~\mu m$
	wide; rarely with sporophytes; plants of moist habitats
	Brachythecium mildeanum (w)
-	Leaf margin denticulate; alar cells quadrate to short
	rectangular; median lamina cells 7–9 μm wide; capsules
	often inclined; often with sporophytes; plants in mostly
	dense, but usually small turfs, with erect shoots, in open
	deciduous forests in calcareous areas.
40	Brachythecium capillaceum (r)
18	Branches often complanate; costal spine relatively strong;
	seta rough above, smooth below; leaf margin strongly
	denticulate; plants growing in different types of forest, on
	forest litter, humus-rich soil and soil-covered stones and
	rocks, sometimes even on rotting wood; also in spruce
	plantations
	Branches not complanate; costal spine absent or
_	occasionally present and very weak; seta completely and
	conspicuously rough; leaf margin denticulate; common
	plants of nutrient-rich habitats with very wide ecological
	amplitude
	2) dony diecium i dedudum (cc)

Key to species and varieties of *Hypnum* including *Buckia*

1	Alar cells increasing in size towards leaf base, $12-20~\mu m$ wide; medium sized plants
-	Alar cells homogeneous, of equal size, hardly > 10 µm in diameter; small plants
2	Plants complanately ± regularly pinnately branched, pale to whitish green, leaves strongly curved, margins denticulate in upper half, in particular the branch leaves; alar cells yellowish, forming excavate auricles, somewhat decurrent; growing mostly on soil
-	Plants irregularly pinnate, green to yellow-green; leaves denticulate only at apex, rarely down to mid-leaf; variable; growing mostly on bark and rock. <i>Hypnum cupressiforme</i> 3
3	Plants complanate, branches usually less wide than principal stem4
-	Plants not complanate, in general julaceous, branches as wide as the principal stem, mostly robust plants with ovatelanceolate to linear-lanceolate, ± concave leaves
4	Plants usually forming mats, leaves mostly strongly falcate, usually turned towards the substrate; alar group slightly to strongly excavate
-	Plants usually forming wefts, leaves ± straight or slightly falcate, usually oriented away from the substrate, shoots often slightly julaceous; alar group conspicuous, strongly excavate, alar cells incrassate, often brownish
5	Leaves ovate, rather abruptly narrowed to the acumen, concave; dioicous; plants julaceous, often erect, in yellowish green to brownish green mats; on calcareous rocks
-	Leaves ovate-lanceolate, gradually and longly acuminate; autoicous, often with capsules; leaves weakly falcate, denticulate all around, in particular branch leaves; alar groups fairly distinct; capsule erect or curved, lid rostrate; plants pale green, 2–4 cm long; on tree bases and tree trunks **Hypnum pallescens* var. reptile** (w)

Key to species of Taxiphyllum

-	Costa short, double,	ceasing before	re mid-leaf, rai	ely not
	developed; leaves ±	complanate;	plants to 1 c	m long,
	pinnately branched,			
	cucumber smell	Taxi	iphyllum wissgr	illii (w)

Key to species of Calliergonella

- Leaves straight, ovate or elongate-ovate, above suddenly narrowed to a rounded, occasionally apiculate apex; stem and branch tips pointed due to closely imbricate leaves.......
- Calliergonella cuspidata (w)
 Leaves ± falcate-secund, longly and gradually tapering from

Key to species of Rhytidiadelphus

Key to species of *Neckera* s.l. (incl. *Alleniella, Exsertotheca, Neckera*)

Leaf apex obtuse to very shortly acuminate, apiculate; leaves 1.5–2 mm long; mid leaf cells 20–60 µm long..... Leaf margin narrowly recurved from base to apex, at least 3 on one side, broadly incurved in basal part on other side: plants small, mostly to 2-3 cm (5 cm) long; often with filiform small-leaved flagelliform caducous branches; dioicous, sporophytes rare, exserted... *Neckera pumila* (n.s.) Leaf margin broadly incurved on one side at leaf base, otherwise flat, not narrowly recurved; plants medium-sized to robust, 3-20 cm long; flagelliform branches rare; autoicous or dioicous, sporophytes immersed or exserted...4 Leaves gradually tapering from mid-leaf to sharp tip; plants 4 medium-sized, 3-5 (10) cm long, leaves to 2.5 mm long; nearly exclusively epiphytic; autoicous, often with immersed capsules *Neckera pennata* (r) Leaves ± parallel-sided for most of their length, abruptly tapering to short, obtuse tip; plants robust, 5-20 (30) cm long, leaves 2.5-4 mm long; mostly saxicole, more rarely epiphytic; dioicous, sporophytes rare, capsule exserted..... Exsertotheca crispa (Neckera crispa) (w) **Key to species of** *Thamnobryum* Reference: Mastracci (2003) Branch leaves distinctly concave and glossy, with a short and coarsely dentate apex: shoots unbranched or irregularly branched; submarginal cells not elongate..... Thamnobryum neckeroides (n.s.) Branch leaves plane, with an elongate and dentate apex; shoots dendroid; submarginal cells elongate, sharply delimited from the other lamina cells towards the costa..... Thamnobryum alopecurum (w) Key to species of *Isothecium* Margins denticulate to below mid-leaf; branch leaves longly and finely pointed; branching ± dendroid..... **Isothecium myosuroides** (r)

Key to species of *Anomodon* s.l. including *Pseudanomodon, Claopodium*

1	Leaves narrowed to an attenuate apex; leaf cells unipapillose or pluripapillose 2
-	Leaf apex broad or narrow, obtuse or bluntly pointed; leaf cells pluripapillose
2	Leaf terminating in a long, hyaline, uniseriate point; lamina cells pluripapillose.
_	Leaf not terminating in uniseriate hyaline point; lamina cells
	unipapillose
3	Leaf apex triangular-acuminate, denticulate; plants often
	with attenuate (progressively smaller leaved) branches
	Pseudanomodon attenuatus (Anomodon attenuatus) (c)
-	Leaf apex rounded or apiculate, entire4
4	Leaf base decurrent, irregularly tearing when removed; papillae only on the outer side of the leaf base; leaves gradually narrowed to tapering apex, homomallous, robust
	plants, to 10 cm long
	Leaf base auriculate; auricles rounded, all of their margin
_	with high, partly branched papillae; leaves abruptly
	contracted from broadly oval base to an almost linear,
	lingulate, paralell-sided apex, not homomallous; slender
	plants, to 5 cm long
	process, to a contraction of the process of the pro

GLOSSARY

Most of the definitions of this glossary are compiled from the following treatments: Magill (1990), Malcolm and Malcolm (2000), Frey et al. (2006), Damsholt (2002), Paton (1999), Smith (2004), Watson (1981).

abaxial - see dorsal side away from stem or axis; back, dorsal, or lower

surface of leaf or costa (opposed to adaxial)

acidophilous acid-loving, preferring an acid habitat, growing on

substrata that are acid in reaction

acrocarpous refers to the production of archegonia and later sporophytes at the apices of the main stems; the

main stems are usually erect, tufted and sparingly

branched

acumen (pl. acumina)

acuminate

a slender, tapering point

slenderly tapered with an angle of less than 45°;

longer than acute

sharp pointed, with terminal angle less than 90° but acute

greater than 45°

adaxial - see ventral side towards stem or axis; ventral or upper surface

of a leaf or costa (opposed to abaxial)

adventive thalli lateral thalli, e.g. in Metzgeria, which break away

and form a means of asexual reproduction

a depression, e.g. on the surface of a spore

underleaf in leafy liverworts; see underleaf

air chamber specialized internal air-containing cavity common in

most complex thalloid liverworts, e.g. Marchantiales referring to cells at basal margins (angles) of a leaf; these cells are often differentiated in size, shape or color from other leaf cells, e.g. Dicranum; see

auricles

alveola (pl. alveolae) amphigastrium

(pl. amphigastria)

androecium antheridia and surrounding leaves (perigonium),

the male inflorescence

see alar cells

angular cells

angulate

alar cells

angled, having angles or corners

annulus (pl. annuli) in stegocarpous mosses a zone of variously

differentiated cells between the capsule urn and operculum, facilitating opening of the capsule; cf.

valve

antheridium (pl. antheridia) male gametangium; a multicellular globose to broadly cylindric, stalked structure containing

spermatozoids

antical the dorsal surface of a stem; the leaf margin

oriented towards the shoot apex of a longitudinal or obliquely inserted leaf (opposed to postical); in leafy liverworts with conduplicate leaves the antical lobe

may also be called dorsal lobe

aperture an opening or hole

apical at apex, summit or point of a structure

apical lamina in *Fissidens* the part of the leaf above or distal to the

(pl. laminae) vaginant and dorsal laminae apiculus (pl. apiculi) a short, abrupt point

apophysis a strongly differentiated, sterile neck at the base of a capsule, between seta and urn; e.g. Polytrichum (see (pl. apophyses)

hypophysis, neck)

appendiculate with short, thin, transverse projections (see

nodulose)

archegonium female gametangium or sex organ; a multicellular, (pl. archegonia) flask-shaped structure consisting of a stalk, venter,

neck and containing an ovum

arcuate curved

areola (pl. areolae) small angular or polygonal surface area differing in

colour or structure from the surrounding area. forming a pattern or network; e.g. air chambers on a

thallus: see also alveolae

the cellular network of a leaf or thallus areolation

ascending pointing obliquely upward; away from the substrate attenuate

slenderly tapering

auricle a small, ear-like lobe; often present at the basal

margins of leaf in mosses; e.g. Climacium (see alar cells); also on thalli and other organs in liverworts

autoicous with archegonia and antheridia in separate clusters

on the same plant (see synoicous, paroicous, dioicous, monoicous) [the suffix "-oecious" strictly applies only to sporophytic or diploid sexuality, and thus is inapplicable to bryophytes; however, it is nonetheless frequently employed (autoecious) as an alternative spelling and pronounciation to "-oicous"] the upper angle formed by the axis and any organ

that arises from it; e.g. leaf and stem, stem and

branch

axillary hair hair in the leaf axils

axil

axis (pl. axes) the main stem; a conceptual line around which

structures (leaves, branches, etc.) develop

basal at the base, bottom or proximal end (opposed to

terminal or distal)

basal cell cell at the base; in leaves, frequently differentiated

cells of the lower 1/4-1/3 of a leaf (cf. lamina cells); cell at point of attachment of an axillary hair

basal membrane a short tube or cylinder often supporting segments and cilia of the endostome; e.g. Bryum; or teeth of

the haplolepidous peristome; e.g. Tortula

biconvex convex on both sides, e.g. the costa of Grimmia

teretinervis

bifid divided into two parts, see bilobed, bifurcate **bifurcate** Y-shaped or forked; leaves divided into two ± equal

parts

bilobed divided into two lobes or segments

bistratose composed of two cellular layers; e.g. leaf blades that

are two cells thick

border cells along a leaf margin that differ from other leaf

cells in their shape, size, colour, or wall thickening

bract a modified leaf associated with a gametangium or

gemma-cup (cf. perichaetial leaf, perigonial leaf)

brevipilose with a short hair

brood bodies any structures that function as asexual propagules

such as gemmae, bulbils, tubers or reduced

branches or leaves

bulbil vegetative propagule; a small, deciduous, bulb-like

axillary propagulum

bulbousswollen, bulb-likecaducouseasily or soon falling away

calcicole growing in calcareous habitats, as on dolomite and

limestone

calcicolousgrowing on a limy substratumcalcifugeavoiding calcareous habitats

calciphile favouring calcareous habitats, as on dolomite and

limestone

calyptra (pl. calyptrae) a membraneous covering of haploid tissue over the

developing sporophyte, derived from the archegonial venter. In mosses the venter generally ruptures near the base, is carried upwards by elongation of the seta, and frequently expands to form a protective covering over the capsule. In liverworts the structure is strictly an epigonium that ruptures near the apex and remains at the base of

the seta; see vaginula

campanulate bell-shaped; referring to a calyptra that is elongated

and cylindrical; a campanulate-cucullate calyptra is cylindrical and split on one side only, e.g. *Ceratodon purpureus*; a campanulate-mitrate calyptra is cylindrical and undivided or equally lobed at the

base, e.g. Grimmia pulvinata

canaliculate channelled (lengthwise); as in leaves or thalli

than the control of t

capitulum (pl. capitula) head; e.g. Sphagnum the sporangium; ter

the sporangium; terminal spore-producing part of the sporophyte; in most mosses it is differentiated into an apical operculum, central urn (spore-bearing region) and a sterile basal neck or hypophysis; in most liverworts and hornworts it is a uniform structure containing spores and elaters or pseudoelaters and usually opens by splitting into 4 (or 2) valves carinate like a boat keel; winged

catenulate chain-like; e.g. arrangement of leaves on the stem of

some species of Leskeaceae

cauline of the stem

caulonema in mosses the second of the two growth stages of a (pl. caulonemata)

typical protonema (the first stage is called chloronema). A caulonema typically has pigmented walls and oblique cross-walls, whereas chloronema has hyaline walls and transverse crosswalls. Only the caulonema ordinarily produces the buds which develop into the familiar gametophytes

of an adult moss

central strand a small group of elongate cells forming a central axis

of some stems and thalli, usually colored and thin-

walled in transverse section

nodding or drooping, orientation of a capsule with cernuous

longitudinal axis intermediate between

horizontal and pendulous

channelled hollowed out like a gutter and semicircular in cross

section (cf. keeled)

tissue composed of cells containing chloroplasts chlorenchyma chlorocvst

green (chlorophyllose) cell; generally used in contradistinction to hyalocysts; e.g. leaves of

Sphagnum, Leucobryum

chloronema

in mosses the first of the two growth stages of a (pl. chloronemata) typical protonema (the second stage is called

caulonema). A chloronema typically has hyaline walls and transverse cross-walls, whereas a caulonema has pigmented walls and oblique crosswalls. Only the caulonema ordinarily produces the buds which develop into the familiar gametophytes

of an adult moss

chlorophyllose containing chlorophyll; generally green unless

masked by some other pigments

ciliate fringed with hair-like appendages (see fimbriate) cilium (pl. cilia) a delicate hair- or tooth-like structure fringing a leaf,

thallus or other structure, or alternating with the processes of the inner peristome (endostome)

circinate curved in a circle; e.g. leaves of Sanionia uncinata clavate

thickened towards the apex; club-shaped

cleistocarpous indehiscent; capsule without a regular mechanism for opening; e.g. capsule lacking a operculum and annulus and hence opening irregularly (opposed to

stegocarpous)

collenchymatous with cell walls more heavily thickened at the (leaf or

exothecial cell) angles; e.g. Aulacomnium palustre;

(see trigones)

the central, sterile tissues in the sporogenous region columella

of a capsule in most mosses and hornworts

comal tuft a tuft of leaves at tip of stem or branch; e.g. *Bryum*comissure junction; e.g. in leaves of *Sphagnum*, the seam

between adjacent cell walls of hyalocysts and

chlorocysts

complanate flattened or compressed, such as leaves flattened

into more or less one plane, e.g. Plagiothecium

complicate-bilobed a bifid leaf with the two segments folded together

longitudinally

concave having an outline or surface curved like the interior

of a circle or sphere

concolorous said of two or more parts of structures that have the

same colour

conduplicate strongly folded longitudinally along the middle; e.g.

the leaves of *Fontinalis antipyretica* or the sheathing part of the leaves of *Fissidens*; in liverworts like *Scapania* short for bilobed-conduplicate, one lobe

folded over the other

conical cone-shaped; operculum of *Bryum*

connate joined, used when similar parts of an organism are

fused together

connivent directed or pointing together, though not fused, as

the tips of leaf lobes inclined and converging

towards one another

convex having an outline or surface curved like the exterior

of a circle or sphere

cordate heart-shaped, said of leaves attached at the broad

end

cortex (adj. cortical) the outer layer of cells of the stem, surrounding the

medulla

corticolous growing on bark

costa (pl. costae) nerve or midrib of a leaf or thallus, always more

than one cell thick

crenulate with minute rounded teeth

costate having a midrib (costa), opp. ecostate

cribrose finely perforated; e.g. peristome teeth of *Grimmia* **crispate, crisped** wavy; often used more loosely to mean variously

curled, twisted and contorted

cryptopore having immersed stomata, with the guard cells

sunken below level of the exothecial cells and often

± covered by them (opposed to phaneropore)

cucullate hooded or hood-shaped; a calyptra split along one

side only; also used to describe leaves strongly concave and erect or inflexed at the tips, like a

monk's cowl

cuneate (cuneiform) wedge-shaped

cuspidateending abruptly in a stout, rigid point (cf. apiculate)cuticlean extracellular cutinized layer on the epidermis of

most complex thalloid liverworts, leaves or stems of mosses, setae and capsules of mosses and capsules

of hornworts

cygneous curved like a swan's neck; e.g. seta in many species

of Campylopus

cvlindrical elongate and circular in transverse section deciduous falling off, compare caducous, fugacious decumbent with stem prostrate, but with ascending tips

decurrent with basal leaf margins extending down the stem

past the leaf insertion as ridges or narrow wings;

e.g. Plagiomnium elatum

dehiscent the capsule opening regularly by means of an

annulus and operculum or valves (opposed to

indehiscent)

dendroid having a growth habit like a tree

with sharp teeth directed outward (see denticulate) dentate denticulate finely toothed (see dentate; subjectively distinct) referring to capsule after the operculum has fallen deoperculate dextrorse forming a right-handed helix (like in a normal

screw); e.g. twist of seta (opposed to sinistrorse) dimorphic occurring in two forms; e.g. with leaves of two forms dioicous with archegonia and antherida on separate plants diplolepidous peristome type originating from 2 rows of cells on the outer surface of the tooth and one on the inner

surface; usually consisting of a double ring of teeth, the exostome and the endostome; cf. haplolepidous

also disciform, flattened into a plate or disk discolorous said of two or more parts or structures that differ in

colour

discoid

distal away from the base or point of attachment; towards

the apex of a leaf or stem: the outer, convex face of a

spore (opposed to proximal)

distichous leaves alternating in two opposite rows; e.g.

Fissidens, Distichium

divergent (1) spreading from a point of attachment, (2)

spreading in opposite directions

dorsal (of leaves) the abaxial, back or lower surface; (of

peristome teeth) the outer face; (of stems or thalli) the upper surface, away from the substrate (opposed to ventral); see also antical (liverworts

with conduplicate leaves)

dorsal lamina part of the leaf blade opposite the sheathing base, at (pl. laminae) the back of the costa and below the apical lamina;

e.g. Fissidens

flattened on the dorsal and ventral sides dorsiventrally

compressed

ecad an organism that is modified by its environment echinate

with stiff bristles (spore surface)

ecostate without a costa, opp. costate edentate without teeth; i.e. entire leaf margins elater

a differentiated elongate cell, dead at maturity and (pl. elateres, elaters) normally with one to three helicoidal wall thickenings, found interspersed among the spore

mass in most liverwort capsules; function: to break up and subsequently help disperse the spores

a solid that has an elliptical outline ellipsoidal elliptical oblong with convex sides or ends

elongate stretched out; e.g. linear

emarginate broadly notched at the apex (compare with retuse,

more narrowly notched)

partially exposed, referring to capsules or perianths emergent

only partly projecting beyond the tips of perichaetial

leaves (cf. exserted, immersed)

the inner circle of a diplolepidous peristome, formed endostome

> from contiguous periclinal wall-pairs of the primary and inner peristomial layers; typically a weak membraneous structure consisting of a basal membrane bearing segments and cilia. The endostome is homologous with the haplolepidous

peristome

without teeth; ± smooth on the margin; not divided entire

into lobes, e.g. leaves, thalli

epapillose without papillae; opp. papillose

ephemeral short-lived; refers to an organism which completes

its life cycle in a single season

the outer cell layer of a stem or thallus; frequently epidermis

fragile and ephemeral (adj. epidermal)

lacking hairs or a hair point, opp. pilose (hairy) epilose epiphragm in Polytrichaceae: a circular membrane formed by

(pl. epiphragmata) the expanded tip of the columella and attached to the ends of the peristome teeth; it partially closes the mouth of the capsule after the lid has dropped

away. Similar structure in Weissia brachycarpa, W.

condensa, W. rostellata

epiphyte a plant growing on another plant

with leaves directed toward stem apex; with leaf erect

margins curved upward (adaxially); with capsules

straight, not curved

erecto-patent spreading at an angle of 45° or less (cf. spreading,

patent)

ragged or irregularly notched as if gnawed by an erose

animal

not wavy, opp. sinuose esinuose

evolute rolled out, of a leaf lobule which is unrolled or plane,

not inflated

hollowed out excavate

excurrent extending beyond the apical margin; e.g. an awn

formed by a protruding costa

exostome the outer circle of a diplolepidous peristome,

formed from contiguous periclinal wall-pairs of the outer and primary peristomial layers; missing or

rudimentary in haplolepidous peristomes

exothecial with reference to the exothecium

exothecium the outermost layer of the capsule wall, consisting of

exothecial cells, the capsule epidermis

explanate flattened and spread out

exserted projecting and exposed; e.g. capsules or perianths

held clear of the tips of perichaetial leaves (cf.

emergent)

falcate curved like the blade of a sickle falcate-secund strongly curved and turned to one side

fascicle a group, cluster or bundle of branches; e.g. branches

in Sphaanum

fertile producing sex organs (antheridia, archegonia), opp.

sterile

fibril fine, fiber-like wall thickenings

fibrillose with fine, fiber-like wall thickenings (fibrils; applied

to Sphagnum hyalocysts where fibrils may be spiral

or annular)

slender and elongate, filamentous, thread-like filiform

filament a single row of cells attached end-to-end, long and

sometimes branched

fimbriate fringed, generally with radiating cell walls of partly

eroded marginal cells, e.g. stem leaf apices of

Sphagnum fimbriatum

flagellum (pl. flagella),

fugacious

refers to a long thin shoot with small (reduced) adj. flagelliform

leaves, functioning as a means of asexual

propagation

flexuose slightly and irregularly bent, twisted, or wavy leafy or leaf-like; closely covered with leaves foliose

vanishing or readily falling away (see caducous,

deciduous

furcate forked (see bifurcate)

furrowed

fusiform spindle-shaped, narrow and tapered at both ends a specialized gametangia-bearing branch, producing gametangiophore

either archegonia (archegoniophore) or antheridia

(antheridiophore)

gametangium vessel bearing gametes; e.g. archegonium,

antheridium (pl. gametangia)

gametoecium gametangium and surrounding leaves

(pl. gametoecia) androecium, gynoecium)

gametophore the leafy stem of the gametophyte, in mosses

produced from buds on the caulonema stage of the

protonema and usually bearing sex organs

(gametangia) later

gametophyte the haploid, sexual generation; in bryophytes, the

dominant generation

geminate arranged in pairs, e.g. the marginal teeth in *Mnium* **gemma (pl. gemmae)** uni- or multicellular, filamentous, globos

uni- or multicellular, filamentous, globose, ellipsoidal, cylindrical, stellate or discoid brood bodies, relatively undifferentiated, serving in

vegetative reproduction (cf. brood body)

gemma cup cup-shaped, gemmae-containing structure of thalline (*Marchantia*) or foliar (*Tetraphis*) origin,

presumably structured to aid in distribution of

gemmae by water splash (cf. splash-cup) bearing gemmae

gemmiferous,

globose

guard cells

gemmiparous

gibbous swollen or bulging on one side

glabrous smooth, not papillose, rough or hairy (see naked)

spherical

the specialized inflatable epithecial cells surrounding a moss or hornwort stoma, usually paired and kidney-shaped, but in some species of the moss family Funariaceae only a single cell with a central slit-like opening, sometimes also used for the cells surrounding air-pores in the epidermis of

thallose liverworts

guide cell large, highly vacuolated, thin-walled and

longitudinally arranged cells found in a median layer across the costa of many mosses, part of conducting parenchyma; e.g. taxa of Pottiaceae characteristically have guide cells sandwiched between two layers of stereid cells in the costal

cross section

gynoecium a structure containing female reproductive organs

(pl. gynoecia) (archegonia)habitat local environment

hair point usually formed by an excurrent costa; e.g. *Tortula*

hamate (or hamulose) hook shaped, more abruptly curved than falcate;

syn. uncinate

haplolepidous peristome type originating from one row of cells of

the outer surface of the tooth and two rows of cells on the inner surface (homologous to the endostome

of diplolepidous mosses)

heterogeneous made up of contrasting or unrelated parts; opp.

homogeneous

heteromorphous having different shapes

heterothallic male plants much smaller than female plants, e.g. in

Riccia frostii

homogeneous uniform or composed of similar parts; opp.

heterogeneous

homomallous pointing the same way (opposed to heteromallous)

homomorphoushaving the same shapehyalinecolorless or transparent

hyalocyst large, empty, water-storage cell as in leaves of

Sphagnum and Leucobryum

hyalodermis differentiated external cells; e.g. thick-walled,

enlarged or hyaline; stem epidermis of enlarged, hyaline cells; i.e. chloroplasts very few or lacking; such cells may be protoplasmic (e.g. *Cephalozia*) or

not (e.g. Sphagnum) at maturity

hydroid tracheid-like conductive cell in the central strand of

some bryophytes, especially mosses, sometimes also

in the costa

hypophysis a strongly differentiated, sterile neck at the base of **(pl. hypophyses)** the capsule, between seta and urn; e.g. *Polytrichum*

closely appressed and overlapping; e.g. with the leaf margins overlapping like shingles on a roof

immersed submerged or below the surface; referring to a

capsule or perianth exceeded by the blades or awns of the perichaetial leaves (cf. emergent, exserted); or

to sunken stomata (see cryptopore) cut or dissected into sharp divisions

incised cut or dissected into sharp divisions

inclined bent down; capsules that are between the erect and

horizontal positions, i.e. drooping

incrassate with thickened cell walls

imbricate

incubous lying upon; an oblique leaf insertion in which the

antical (distal) leaf margins are oriented toward the dorsal stem surface; when viewed from above the antical leaf margins will overlap the postical (proximal) leaf margins of the leaves directly above; found in some leafy liverworts; e.g. *Lejeunea* (cf.

succubous)

incurved curved upward (adaxially) and inward, subjectively

stronger than inflexed and weaker than involute; applied to leaf tips and margins (opposed to

recurved)

indehiscent describes a capsule that does not dehisce by means

of a lid or valves; the spores are released by the rupture or decay of the capsule wall; syn.

cleistocarpous

inflated swollen or bladderlike; puffed up; e.g. the lobules of

certain species of Frullania or Lejeunea, or alar cells

of Cratoneuron (opposed to explanate)

inflexed bent upward (adaxially) and weakly inward; applied

to leaf margins or leaves on a stem (cf. incurved,

involute, inrolled; opposed to reflexed)

inflorescence a structure containing sexual reproductive organs

innovation a new branch, especially one that arises below an

inflorescence

involucral flap a flap of thallus tissue covering the gynoecium; e.g.

Pellia epiphylla

involucre a protective sheath of tissue of thalline origin,

surrounding a single antheridium, archegonium or sporophyte in certain liverworts; often used loosely as a general term of any sheath-like structure

surrounding sporophytes or gametangia

involute rolled upward (adaxilly) and tightly inward, applied

to leaf margins (syn. inrolled; cf. incurved; opposed

to revolute)

irregular asymmetric; not fitting any detectable pattern

isodiametric about as broad as long; applied to cells about the same diameter in all directions; including square,

rounded, or hexagonal

julaceous smoothly cylindric; like a catkin, referring to stems

or branches with strongly imbricate leaves

K+ colour reaction with a 2% solution of potassium

hydroxide KOH, giving either reddish or yellowish colours, e.g. in Pottiaceae or *Schistidium*

keel a ridge formed along a sharp fold of a leaf

(Fontinalis), or perianth (Lejeuneaceae), in liverworts in particular the line of the fold between the smaller dorsal and the larger ventral lobe of a conduplicate bilobed leaf, or between the larger

dorsal and the smaller ventral lobe of a leaf

keeled sharply folded along the middle, like the keel of a

boat; V-shaped in cross section; e.g. leaves of *Grimmia pulvinata, Fontinalis antipyretica* (cf.

carinate)

laciniate fringed with appendages coarser than cilia and

more than one cell wide; cf. ciliate, fimbriate with depressions or perforations on a surface

lacunose with depressions or perforations on a surface lamella (pl. lamellae) parallel photosynthetic ridges or plates along a leaf

blade, or costa; e.g. Polytrichum, Pterygoneurum

lamina (pl. laminae) the flattened, generally unistratose and green part

of the leaf blade excluding the costa and border; the expanded part of a thallus (see apical, dorsal and

vaginant laminae)

lamina cell cell of a lamina (see areolation)

laminate like a lamina, said of leaf primordia with a flat blade lanceolate lance-shaped, in bryology narrow and tapered from

near the base; narrowly ovate-acuminate [used elsewhere for narrowly elliptical and tapering

equally to both ends]

lateral at the side; e.g. lateral branches or thallus wings (cf.

basal and terminal)

loose; referring to large thin-walled cells, as well as

to nature and spacing of leaves on stem, or of stems

in a tuft

leaf a photosynthetic outgrowth from the stem; in

bryophytes generally consisting of a unistratose lamina with or without a multistratose costa; (also called phylloid, indicating that only analogue to but

not identical with the leaf of higher plants)

lenticular doubly convex, lens-shaped

lid see operculum

limb the upper part of a leaf (the opposite of base)

limbidium (pl. limbidia) border, differentiated leaf margin, e.g. in Fissidens

bryoides

linearvery narrow, elongate with nearly parallel sideslingulatetongue-shaped; oblong with a slightly broadened

apex

lobe any segment of a divided (lobed) leaf or other organ;

the larger segment of an unequally divided leaf in

leafy liverworts (see lobule)

lobule a small lobe; e.g. the smaller segment of an

unequally divided leaf in leafy liverworts

lumen (pl. lumina) the cell cavity

lunate crescent shaped

mamilla (pl. mamillae) strongly bulging surface of a cell; also used for

various hollow papilla-like protuberances without associated local wall thickening; i.e., with the cell

lumen extending into the protuberances

mamillate convex to hemispherical with a blunt central

projection; e.g. operculum of *Grimmia orbicularis*

mamillose with mamillae

marginal at the margin, especially as applied to a leaf

marsupiuma modified shoot calyptra; associated with(pl. marsupia)development of sporophyte in a pouch-likestructure that penetrates downward into the

substrate (geocauly)

mat a densely interwoven, horizontal growth form; e.g.

Brachythecium, Hypnum

median central, middle; e.g. median leaf cells are from the

upper middle of a leaf, midway between costa and

margin (see mid-leaf)

medulla the interior, non-cortical part of the stem midrib chief median part of a thallus (cf. costa)

mitrate (mitriform) conic and undivided (similar to a bishop's mitre) or

equally lobed at base, referring to calyptrae (opposed to cucullate; see campanulate) e.g.

Grimmia

monoicous bisexual; with antheridia and archegonia on the

same plant, including autoicous, synoicous,

paroicous, and polyoicous (opposed to dioicous;

monoecious, see note after autoicous)

mucilage cell a specialized cell that is either filled with or secretes

gelatinous, water absorbent substance. presumably a mucopolysaccharide (syn. slime cell)

mucro (pl. mucrones) a short, abrupt point (see apiculus)

muticous without a point or awn mvcorrhiza

fungal hyphae in symbiosis with some liverworts neck (1) in mosses: the sterile basal portion of a capsule

between the base of the theca (urn) and the top of the seta (syn. apophysis, hypophysis (2) the

cylindrical upper portion of an archegonium

nodulose with nodular thickenings; minutely knobbed; sometimes referred to intracellular wall thickening;

e.g. Racomitrium

wider than long oblate

slanted; e.g. an oblique leaf insertion is one that is oblique

between transverse and longitudinal

obloid a solid with an oblong profile

oblong rectangular with rounded corners or ends obovate egg-shaped with apex broader than base obovoid an inversely ovoid solid

broadly pointed; more than 90° - used by some obtuse

authors to mean blunt or rounded

ocellus (pl. ocelli) an idioblastic leaf cell having one large oil body and

lacking chloroplasts, also found in underleaves, bracts and perianths of certain leafy liverworts

(Frullania)

a membrane-bound, terpene-containing organell oil-body

unique to the cells of liverworts

oil-cell an idioblastic cell characterized by a very large oil

> body. common in thalloid liverworts;

Marchantia

operculum the lid covering the mouth of most moss capsules; usually separated from the mouth by an annulus to (pl. opercula)

open the capsule (see stegocarpous)

orbicular nearly circular widely elliptical oval

outline of an egg with base broader than apex ovate

ovoid an egg-shaped solid

palmate with finger-like lobes radiating from centre

papilla (pl. papillae) ornamentation. а solid microscopic

protuberance of cell wall (see papillose, and slime papilla)

papillose bearing papillae; monopapillose - bearing one

simple unbranched papilla on the cell surface. Loosely applied to any minutely rough surface, that

may be strictly mamillose; e.g. pottioid leaves

paracostal beside the costa

paraphyllium small green outgrowths of various shapes, i.e., (pl. paraphyllia) filiform, lanceolate, scale- or leaf-like or sometimes

branched; produced randomly on the stems or branches of many pleurocarpous mosses; e.g. *Thuidium*. In hepatics, occasionally on stems or associated with perigonial bracts; e.g. *Scapania* (cf.

pesudoparaphyllia)

paraphysis hyaline or yellowish, usually uniseriate, hair often (pl. paraphyses) associated with antheridia and archegonia of

mosses

parenchyma a tissue of relatively undifferentiated, usually thin-

walled and isodiametric cells, with non-overlapping

end walls

paroicous with antheridia and archegonia in a single

gametoecium but not mixed, the antheridia in the axils of bracts just below the bracts surrounding the archegonia (paroecious; see note after autoicous)

patent of leaves spreading from the stem at an angle of 45°

or more

pectinate resembling a comb

pedicellate borne on a stalk or pedicel, such as most bryophyte

gametangia

pellucid translucent

pendulous

peltate shieldlike structure fixed on a central stalk

pendent (pendant) hanging downward (cf. inclined, pendulous); e.g.

pendent branch (*Sphagnum*, opp. spreading branch) hanging, pendent; e.g. capsules drooping and

inclined beyond horizontal; stems and branches that

hang

percurrent extending to the apex

perennial a plant in which the vegetative parts live year after

vear

perforate pierced through

perianth organ of foliar origin enclosing the archegonia in

most leafy liverworts

perichaetial leaf modified leaf or underleaf (bract; bracteole)

associated with the gynoecium; collectively forming

the perichaetium

perichaetium the gynoecium; strictly the ensheathing cluster of **(pl. perichaetia)** modified leaves or underleaves (bracts; bracteoles)

modified leaves or underleaves (bracts; bracteoles) and perianth, if present, enclosing the archegonia

perigonial leaf modified leaf or underleaf (bract; bracteole)

associated with the androecium; collectively

forming the perigonium

perigonium the androecium; strictly the cluster of modified (pl. perigonia) leaves or underleaves (bracts; bracteoles) enclosing

the antheridia

perigynium in some Jungermanniidean liverworts a tubular

extension of the stem tissue surrounding the gynoecium, elevating the female bracts and perianth

(if present)

peristome a circular structure, generally divided into 2ⁿ (i.e. 4,

(rarely multiple) row around the mouth of a capsule; (see endostome, exostome, prostome; also teeth, inner, outer, single and double peristome)

phaneropore with superficial stomata; with guard cells at the same level as other exothecial cells and not sunken

in chambers (opposed to cryptopore)

piliferous having a hair point

pinnate with numerous, spreading branches on opposite

sides of the axis and thus resembling a feather

pitted having small depressions or holes in the cell wall;

sometimes called pores; in chlorocysts they are conspicuous as depressions in thickened walls

between adjacent cells; e.g. Dicranum

plane flat, not curved or wavy, referring to leaf margins or

blade

pleurocarpous refers to the production of archegonia on short side

branches in mosses; as a result the sporophytes appear lateral; most pleurocarpous mosses are prostrate, highly branched and often mat-forming

plica (pl. plicae) longitudinal furrow or pleat

plicate with longitudinal furrows or pleats (plicae)

pluristratose in several layers

polyoicous with several forms of gametoecia on the same plant a small aperture, the opening in the wall of some

a small aperture, the opening in the wall of some cells; e.g. in leaf hyalocysts and hyalodermis of *Sphagnum*; also the central opening in a stoma (see

pseudopores, air pores, pit)

porose having pores

postical the ventral surface of a stem; that leaf margin

oriented towards the base of a longitudinal or obliquely inserted leaf (see proximal; opposed to antical); in leafy liverworts with conduplicate leaves the postical lobe may also be called ventral lobe

primordium the embryonic stage of a leaf or other organ, made **(pl. primordia)** up of undifferentiated (unspecialized) cells

procumbent spreading, prostrate

propagule reduced bud, branch or leaf serving in vegetative (propagulum) reproduction (syn. diaspore, see brood-body)

prorate having papillae or mamillae borne at the tips of cells, or formed by projecting cell ends; e.g.

Philonotis

prosenchyma a tissue made up of narrow elongate cells with

(pl. prosenchymata)

prosenchymatic prostome

prostrate protonema (pl. protonemata)

proximal

pseudopapillose

pseudoparaphyllium (pl. pseudoparaphyllia)

pseudoperianth

pseudopodium (pl. pseudopodia)

pseudopore

pyriform quadrate receptacle tapered overlapping end walls (opposed to parenchyma)

cells characteristic of prosenchyma

a rudimentary structure outside, and usually adhering to, the main peristome teeth: e.g. some species of *Orthotrichum*; (also called preperistome)

lying flat on the ground, creeping

a filamentous, globose or thalloid structure resulting from spore germination and including all stages of development up to the production of one or more gametophores. The protonema is extremely variable as to the amount of chlorophyll present, the degree of obliqueness of its end walls and the degree to which it branches (cf. chloronema, and caulonema). In liverworts a globose, short thalloid or filamentous structure generally gives rise to a single gametophore; in mosses the protonema is typically filamentous although Sphagnum, Andreaea and

Tetraphis have ± thallose protonemata

near the base or point of attachment; the internal

face of a spore (opposed to distal)

having projecting cell walls seen in transverse section of leaf lamina, similar to papillae, but at the cell wall between adjacent cells, not at the cell lumen (German: Pfeilerpapillen); e.g. Dicranum brevifolium, sometimes Flexitrichum flexicaule

Small, unistratose, filiform or foliose structure resembling paraphyllium, but restricted to the areas of the stem around branch primordia; often found in pleurocarpous mosses

a hyaline unistratose sheath outside the calyptra for protection of a single archegonium and the sporophyte (not derived from leaves, thus not being a true perianth)

an elongation of the gametophyte axis below the sporophyte in Sphagnum and Andreaea, serving the function of a seta; also applied to a similar extension

of a stem tip bearing clusters of gemmae

pore-like structure with a thin membrane that may be revealed by staining; e.g. in Sphagnum leaves, consisting of a fibril ring without an interior perforation

pear-shaped square or nearly so

a disc or wart-like mass of tissue bearing antheridia or archegonia and found directly on the thallus (e.g. Conocephalum), inside the thallus (e.g. Pellia), or elevated and terminating a gametangiophore (e.g. Marchantia)

recurved curved downward (abaxially) and inward; in leaves,

referring to margins, apices, or marginal teeth; in peristome teeth curved outward and ± downward

(opposed to incurved)

reflexed bent down (abaxially) and inward generally

referring to leaf margins or leaves on a stem

(opposed to inflexed)

reniformkidney-shapedreticulateforming a network

retort cell a type of cortical cell in some Sphagnum species

shaped like a retort with a projecting neck ending in

a pore

retuse notched at the apex (especially when referring to

leaf shape); less deep than emarginate

revolute rolled downward (abaxially) and backward,

referring to a leaf margin (opposed to involute)

rhizoid hair-like structure that functions in absorption and

anchorage; in liverworts and hornworts one-celled and usually hyaline; in mosses usually brown to reddish, simple or branched, multicellular filaments, generally with oblique end-walls (see tomentum)

rhizoid furrow a furrow in the stalk of Marchantiales, for conveying

rhizoids to the male or female receptacle

rhizoid initial rhizoid-generating cell; such cells are usually

smaller than neighbouring cells

rhizomatous having a slender underground stem, horizontal and

creeping, analogous with the rhizome of higher

plants

rhomboidal oblong-hexagonal

rostrate, rostellate beaked, narrowed into a slender tip or point

rostrum (pl. rostra) beak

rudimentary incompletely developed, vestigial (cf. reduced)
rugose with transverse wrinkles or undulations; e.g. leaves

of Neckera crispa

saccate sac-like; abruptly and deeply concave; e.g. forming a

sac

saxicolous growing on rocks

sclerodermis an internal tissue of thick-walled cells forming a

cylinder inside the hyalodermis; e.g. stem of

Sphagnum

secundturned to one side; e.g. leaves on a stemseptatedivided by cell walls; having partitions

serrate saw-toothed; with marginal teeth pointing forward

(towards apex)

serrulateminutely serratesessilewithout a stalk or seta

seta (pl. setae) elongated portion of the sporophyte between the

capsule and foot (cf. pseudopodium)

sexual condition denotes the distribution of sexual organs within /

among plants, see autoicous, dioicous, polyoicous,

synoicous

surrounding and clasping the stem, base of seta or sheathing

see vaginant lamina

capsule: sheathing base of leaves is frequently

composed of hyalocysts (cf. blade)

sheathing lamina

a stem plus leaves and other appendages

shoot shoulder an area of abrupt narrowing; e.g. the area on a leaf

where the leaf base is abruptly narrowed to the upper lamina or blade or a similar constriction on

an exostome tooth

sigmoid S-shaped

sinistrorse forming a left-handed helix (opposed to a normal

screw); e.g. twist of seta (opposed to dextrorse)

wavy, as in leaf margin, or as in intracellular wall sinuose

thickening of *Racomitrium* (see nodulose); opp.

esinuose

sinus the notch or indentation between two segments, as

in a bifid leaf

slime-papilla clavate mucilage-secreting cell, common on young (pl. papillae) leaves and often terminating very short, uniseriate

filaments (slime hair)

spathulate tapering proximally from a broad, rounded apex

spicate arranged in the form of a spike

spindle-shaped, narrrow (more than 3 times as long as wide) and

fusiform tapered at both ends

with sharp, pointed teeth; also very high, sharp leaf spinose

cell papillae or mamillae

spinulose minutely spiny

wall thickenings arranged spirally in the hyalocysts spiral fibrils

of Sphagnum species and giving the cells their

mechanical strength

spore a reproductive unit produced in a capsule as a result

> of meiosis; usually minute, mostly spherical and generally unicellular bodies that give rise on

germination to a protonema

layer(s) of cells lining the inside and outside of the spore sac

archesporium

sporogonium see sporophyte

sporophyte the spore-bearing generation; initiated by the

fertilization of an egg: remaining attached to the gametophyte and partially dependent on it; typically

consisting of foot, seta and capsule

spreading forming an angle of 45° or more (but less than 90°);

e.g. the adaxial angle between a leaf and stem (see widespreading, erecto-patent, squarrose, patent)

spreading branch branch that at least at the point of insertion turns

(Sphagnum) away from the stem (opp. pendent branch)

squarrose spreading at right angles

stegocarpous referring to capsules with a dehiscent operculum

(opposed to cleistocarpous)

stellate star-shaped

stem the main gametophyte axis of mosses and leafy

liverworts; grows by means of a single apical cell

stereid slender, elongate, thick-walled, fiber-like cell found in groups (stereid bands) in the costa or stems of

some mosses

sterile without reproductive structures or sporophytes;

> generally referring to absence of sexual structures but can also mean absence of asexual structures

stipitate having a stipe or special stalk-like base

stolon a slender elongate spreading branch or stem, with

small and often distinctly shaped leaves, which arches away from its parent plant like a runner of a strawberry plant, 'rooting' where it touches the

substratum by producing rhizoids

minute opening in the capsule wall of hornworts, stoma (pl. stomata)

and usually in the capsule neck of mosses;

surrounded or bordered by two guard cells

in layers; e.g. denoting thickness of leaves; i.e., uni-, stratose

bi-, multistratose (cf. seriate)

stria (pl. striae) fine ridges or lines

striate marked with fine ridges or lines (striae)

striolate finely ridged

strumose with a goiter-like swelling (or struma) at one side of

the base, applied to some capsules; e.g. Ceratodon,

Cynodontium strumiferum

a column; a one-celled, uniseriate or multiseriate, stylus (pl. styli)

subulate to triangular structure found between the lobule and the stem in certain leafy liverworts; e.g.

Frullania

substereid almost with the characteristics of a stereid, but with

walls not as strongly thickened

the substance on which a bryophyte grows; e.g. soil, substrate

bark, rock

subula (pl. subulae)

a long, slender point subulate slenderly long-acuminate

succubous lying under; an oblique leaf insertion in which the

antical (distal) leaf margins are oriented toward the ventral stem surface: when viewed from above the antical leaf margins will lie under or be overlapped by the postical (proximal) leaf margins of the leaves directly above; found in various leafy liverworts (cf.

incubous)

sulcate strongly plicate, with deep longitudinal furrows or

grooves; e.g. capsules of *Ulota*

supra-alar cells in Racomitrium the cells immediately above the alar

with antheridia and archegonia mixed in the same synoicous

gametoecium (synoecious: see note after autoicous)

rounded in cross section terete

terminal at the apex, tip or distal end (opposed to basal)

terrestrial growing on (dry) ground terricolous growing on soil (terrestrial) of or pertaining to a thallus thallose

thallus (pl. thalli) a ± flattened gametophyte, not differentiated into a

stem and leaves

theca (pl. thecae) the spore-bearing portion of a moss capsule (see

urn; opposed to hypophysis)

woolly, densely radiculose; beset with tomentum tomentose tomentum a felt-like covering of abundant rhizoids, on some stems or rarely leaves; e.g. stems of Dicranum (pl. tomenta) trifarious arranged in three rows or ranks; syn. tristichous trigones generally triangular or circular intracellular wall thickenings, found at the point where three (or more) cells meet; especially common in leaf cells of

liverworts (see collenchymatous)

tristichous in 3 ranks

truncate abruptly cut off or squared off at the apex

tuber (in liverworts) a geotropic outgrowth from the

shoot apex, composed of perennating tissue, allowing for aestivation and subsequent continued growth or vegetative reproduction; (in mosses) gemmae borne on rhizoids (rhizoidal gemmae), found in many acrocarpous mosses; e.g. Bryum

tuberculate with small warts

tuberous like a tuber (rhizoidal gemma), e.g. swollen parts of

underground persistent protonema of Ephemerum

tuberculate rhizoid

(pegged rh.)

rhizoid with uniformly dispersed intracellular wall projections or tubercles that increase the surface area per unit volume; in some thalloid liverworts;

e.g. Marchantiales (syn. pegged rhizoids)

cylindrical and apparently hollow

tubular

tubulose tubelike, usually referring to leaves with strongly

incurved or broadly overlapping leaf margins; e.g.

growth form with stems erect but radiating at the tuft

edges; small cushions; caespitose habit; e.g.

Orthotrichum

tumid inflated, swollen

turbinate shaped like an old-fashioned child's top, obconic, i.e. an inverted cone

turf growth form with stems erect, parallel and close

together; often covering extensive areas; e.g. Bryum

argenteum

turgid plump or swollen

uncinate hooked; tip bent in the form of a hook

underleaf ventral, variously modified leaf in most leafy

liverworts (syn. amphigastrium)

undulate wavy

unipapillose with a single papilla per cell

uniseriate in one series; applied to a hair-like structure

comprised of a single row of cells

unistratose one-layered; comprised of a single cell layer; e.g.

most bryophyte leaves

urceolate urn-shaped, applied to capsules constricted below a

wide mouth and abruptly narrowed to the seta

urn spore bearing portion of a capsule (opposed to neck;

syn. theca)

vaginant lamina in Fissidens, one of the two clasping laminae below

the apical lamina

vaginate sheathing

vaginula a ring or sheath enveloping the base of the seta,(pl. vaginulae) derived from the base of the archegonium and

surrounding stem tissue and remaining after the

separation of the calyptra

valve one of the parts or partially detached flap of tissue

into which the capsule of most liverworts and hornworts separates upon dehiscence; rare in

mosses (e.g. *Andreaea*)

ventral (of leaves) the adaxial, top, or upper surface; (of

peristome teeth) the inner face; (of stems or thalli) the lower surface, next to the substrate; (opposed to dorsal); see also postical (liverworts with

conduplicate leaves)

ventral scale a unistratose, leaf-like structure of epidermal origin,

often hyaline or reddish; ventral scales most commonly occur in two or four rows along the ventral surface in complex thalloid liverworts

ventricose bulging on one side below (like a stomach)

vermicular, vermiform worm-shaped; long, narrow and somewhat wavy,

commonly with rounded ends; usually applied to

cells

verrucose covered with small wart-like elevations

verruculose irregularly roughened

verticillate whorled

vesicularcomposed of blistersvesiculoseinflated, bladderlike

vitta (pl. vittae) a band or ribbon; in liverworts the longitudinal

stripe of longer, often thicker-walled cells in a leaf lamina resembling a nerve but only one cell layer

lannina resembling a nerve but only one (

thick; e.g. *Diplophyllum albicans*

wart a small elevation or protuberance (cf. papillae)
water sac an inflated or inrolled lobule or leaf part, usually

filled with water; e.g. Frullania

wedge-shaped see cuneate

weft a loosely interwoven, often ascending growth form:

e.g. Thuidium

wing a thin, flat membranous expansion or appendage

such as the margin of a spore; the keel of a perianth or folded leaf, or loosely applied to the lamina of a thallus or basal angles of leaves; i.e. alar cell region

xeromorphic adapted for dryness

zig-zag alternate from side to side; a line with numerous

reversing angles e.g. median line on diplolepidous

exostome tooth

zygote the product of the fusion of two gametes; fertilized

ovum before it has undergone mitosis or meiosis

INDEX OF GENERA

(accepted names in normal print, synonyms in italics)

Abietinella 18, 28, 74, 219

Acaulon 17, 24, 51, 64, 145

Alleniella 6, 19, 29, 47, 228, 229

Aloina 17, 24, 48, 145, 146

Amblyodon 26, 59, 63

Amblystegium 6, 18, 27, 77, 78, 82, 216, 217

Amphidium 24, 53, 71

Anacamptodon 13, 27, 33, 74, 78, 203

Anastrophyllum 20, 31, 32, 43

Andreaea 13, 17, 23, 44, 126, 246, 251

Aneura 5, 22, 39, 109

Anomodon 6, 19, 29, 33, 49, 75, 230

Anthoceros 20, 37

Antitrichia 29, 75, 78

Aphanorrhegma 51, 64, 65, 131

Apometzgeria 110

Apopellia 22, 39, 111

Archidium 24, 51

Asterella 6, 17, 22, 31, 32, 38, 112, 114

Athalamia 38, 113

Atrichum 23, 48, 126, 127

Aulacomnium 18, 26, 56, 61, 66, 69, 71, 211, 234

Barbilophozia 6, 16, 20, 41, 42, 85, 89, 90, 92, 93

Barbula 10, 12, 25, 58, 69, 71, 169, 171

Bartramia 26, 52, 67, 72, 73, 189

Bazzania 21, 40

Blasia 22, 39

Blepharostoma 21, 40

Blindia 25, 56, 66, 174

Blindiadelphus 25, 33, 56, 59, 72, 174

Brachydontium 25, 54

Brachytheciastrum 6, 19, 28, 79, 223, 226

Brachythecium 6, 19, 28, 33, 76, 79, 223–226, 242

Bruchia 24, 33, 51, 72

Bryoerythrophyllum 25, 58, 67, 71

Bryum 6, 8, 10, 26, 33, 49, 50, 60, 61, 64, 65, 73, 190–198, 232, 235, 250, 251

Buckia 6, 19, 28, 84, 227

Bucklandiella 176, 177, 184, 185

Buxbaumia 9, 10, 17, 23, 45, 130

Callicladium 28, 33, 83

Calliergon 6, 18, 27, 77, 218

Calliergonella 19, 28, 80, 81, 83, 228

Calypogeia 17, 21, 31, 40, 98, 99

Campyliadelphus 18, 27, 76, 79, 215

Campylium 18, 27, 76, 81, 215, 217

Campylophyllopsis 27, 81, 83

Campylopus 8, 9, 17, 24, 49, 54, 62, 63, 133, 134, 141, 236, 250

Campylostelium 25, 58, 71

Cephalozia 20, 31, 32, 86, 87, 240

Cephaloziella 16, 20, 31, 32, 42, 43, 44, 87, 88, 89

Ceratodon 24, 53, 55, 67, 70, 143, 144, 233, 249

Chenia 6, 25, 49, 50, 64

Chiloscyphus 21, 40, 104, 105

Chionoloma 25, 56, 69, 72

Cinclidotus 17, 25, 33, 50, 146, 147

Cirriphyllum 18, 28, 79, 221, 223, 224

Claopodium 6, 19, 28, 49, 75, 230

Clevea 6, 17, 22, 31, 38, 112, 113

Climacium 27, 75, 232

Cnestrum 24, 33, 55, 67, 142

Codonoblepharon 26, 55, 62, 64, 71, 203

Codriophorus 176

Cololejeunea 21, 31, 41, 108

Conardia 27,78

Conocephalum 6, 17, 22, 31, 38, 112, 113, 246

Coscinodon 9, 25, 175, 176, 181, 187

Cratoneuron 27, 76, 78, 240

Crossidium 17, 25, 33, 48, 147

Crossocalyx 20, 43

Ctenidium 29,82

Cynodontium 24, 33, 53, 55, 58, 67, 70, 72, 141, 142, 173, 249

Desmatodon 54, 59, 67, 153, 161, 168

Dialytrichia 17, 50, 146

Dichodontium 24, 58, 59, 66

Dicranella 17, 24, 33, 53, 55, 58, 59, 63, 72, 73, 135–137, 200

Dicranodontium 24, 59, 63

Dicranoweisia 24, 56, 70, 141, 142

Dicranum 9, 11, 24, 33, 55, 59, 63, 66, 134, 140, 141, 142, 231, 245, 246, 250

Didymodon 11, 12, 17, 25, 33, 56, 57, 61, 66, 68–72, 147–151

Diphyscium 23, 51, 69

Diplophyllum 17, 21, 36, 41, 94, 95, 252

Distichium 24, 36, 47, 236

Ditrichum 9, 24, 33, 58, 72, 73, 133, 143-145

Drepanocladus 11, 18, 27, 33, 76, 77, 79, 215, 216, 218

Encalypta 17, 23, 49, 53, 55, 56, 62, 69, 130, 131

Endogemma 6, 17, 21, 32, 41, 101–103

Entodon 29,81

Entosthodon 23, 53, 56, 60, 65, 66, 132

Ephemerum 18, 25, 51, 73, 169, 170, 250

Eucladium 25, 57, 66

Eurhynchiastrum 28, 79

Eurhynchium 18, 28, 76, 78, 79, 220-222, 224

Exsertotheca 6, 19, 29, 47, 228, 229

Fabronia 18, 27, 78, 82, 214

Fissidens 9, 17, 24, 33, 36, 47, 137-139, 232, 235, 236, 242, 251

Flexitrichum 5, 17, 24, 58, 63, 133, 143, 144, 246

Fontinalis 18, 27, 82, 211, 235, 241

Fossombronia 17, 22, 39, 111

Frullania 17, 21, 31, 32, 42, 106, 107, 240, 243, 249, 252

Funaria 9, 24, 54, 65, 131, 132, 239

Fuscocephaloziopsis 20, 32, 86, 87

Grimmia 9, 12, 25, 33, 55, 57, 61, 175, 176, 180–188, 232, 233, 235, 241, 242

Gymnocolea 20, 44, 91, 94

Gymnostomum 6, 18, 25, 54, 63, 70, 170, 171

Gyroweisia 6, 25, 54, 70, 170

Hamatocaulis 28, 76

Hedwigia 9, 11, 18, 26, 46, 49, 51, 70, 188, 189

Helodium 28, 33, 75

Hennediella 6, 18, 25, 54, 67, 153

Herzogiella 27,84

Heterocladiella 29, 74, 80

Heterocladium 7, 29, 74, 80

Hilpertia 6, 18, 25, 33, 49, 57, 58, 64, 158, 160, 163, 165

Homalia 29, 47, 228

Homalothecium 18, 28, 75, 222, 223, 225

Homomallium 29,84

Hookeria 27, 47

Hydrogonium 7, 12, 18, 25, 33, 61, 71, 171

Hygroamblystegium 6, 18, 27, 77, 78, 216, 217

Hygrohypnum 27, 76, 81

Hylocomiadelphus 29, 82

Hylocomium 29, 82, 83

Hymenostylium 25, 54, 72

Hypnum 6, 19, 28, 33, 81, 82, 84, 227, 228, 242

Imbribryum 6, 26, 50, 60, 61, 65, 73, 190, 193–195

Isopaches 6, 16, 20, 89, 91, 92

Isopterygiopsis 27, 48, 83

Isothecium 19, 29, 77, 79, 83, 229, 230

Jamesoniella 40, 41, 101

Jungermannia 6, 17, 21, 41, 101–103

Kindbergia 28, 79

Leiocolea 44, 103, 104

Lejeunea 22, 42, 108, 240, 241

Lepidozia 21, 41

Leptobryum 26, 59, 63

Leptodictyum 27, 47, 79, 221

Leptodon 29,74

Leptophascum 64

Lescuraea 28, 33, 75, 76

Leskea 28, 75, 216

Leucobryum 14, 17, 24, 53, 54, 63, 135, 234, 240

Leucodon 29,80

Lewinskya 14, 26, 52, 55, 62, 70, 71, 203, 204, 206-208

Liochlaena 6, 17, 21, 31, 32, 41, 101, 102

Loeskeobryum 29,83

Lophocolea 21, 31, 40, 44, 104, 105

Lophozia 6, 7, 16, 21, 31, 43, 85, 89, 91-94

Lophoziopsis 6, 16, 21, 85, 91-93

Lunularia 6, 17, 22, 37, 112

Mannia 6, 17, 22, 31, 32, 38, 114, 115

Marchantia 6, 17, 22, 37, 38, 112, 114, 231, 239, 243, 247, 252

Marsupella 17, 21, 42, 43, 100

Meesia 26, 59, 66

Mesoptychia 6, 16, 17, 21, 44, 89, 91, 94, 103, 104

Metzgeria 17, 22, 39, 110, 231

Microbryum 6, 17, 18, 25, 33, 49, 50, 54, 57, 69, 151–155

Microeurhynchium 28,78

Mnium 6, 18, 26, 50, 59, 64, 201, 202, 239

Myurella 27,80

Nardia 6, 17, 21, 41, 44, 100, 101

Neckera 6, 19, 29, 33, 47, 228, 229, 247

Neoorthocaulis 6, 16, 20, 41, 42, 85, 89, 90

Niphotrichum 176

Nogopterium 29, 80

Nowellia 20, 42, 86

Nyholmiella 26, 33, 52, 62, 70, 203, 204, 206

Obtusifolium 6, 16, 20, 89, 91

Orthodontium 26, 33, 56, 60, 73

Orthothecium 7, 18, 27, 83, 214

Orthotrichum 10, 13, 26, 33, 49, 52, 55, 62, 64, 70, 203–211, 246, 250

Oxymitra 6, 17, 22, 31, 115

Oxyrrhynchium 18, 28, 79, 221, 222

Oxystegus 56, 69, 72

Palustriella 18, 27, 75, 214

Paraleucobryum 24, 62, 139

Pedinophyllum 21, 31, 41, 106

Pellia 22, 39, 111, 241, 246

Phaeoceros 20, 37

Phascum 6, 9, 18, 64, 68, 151, 153, 166-168

Philonotis 8, 26, 52, 73, 189, 190, 245

Physcomitrella 51, 64, 65, 131

Physcomitrium 24, 33, 51–53, 64, 65, 131–133

Plagiobryum 49, 60, 65, 73, 191

Plagiochila 21, 40, 41, 102, 106

Plagiomnium 6, 18, 26, 50, 63, 201-203, 236

Plagiopus 26, 52, 67, 189

Plagiothecium 9, 10, 14, 18, 27, 33, 47, 81, 212-214, 235

Plasteurhynchium 28,76

Platydictya 27,82

Platygyrium 28,84

Platyhypnidium 220

Pleuridium 24, 51, 72, 143

Pleurochaete 66, 171

Pleurozium 29,81

Pogonatum 17, 23, 48, 126-129

Pohlia 9, 18, 26, 34, 59, 60, 65, 73, 136, 198-201

Polytrichastrum 17, 23, 48, 126-129

Polytrichum 17, 23, 34, 48, 126, 128, 129, 232, 240, 241

Porella 17, 22, 31, 42, 108

Pottia 6, 18, 49, 51, 54, 57, 64, 68, 152–155, 166, 167

Preissia 112

Protobryum 51, 64, 68, 153, 166

Pseudanomodon 6, 19, 29, 75, 230

Pseudephemerum 24, 51, 73, 143

Pseudoamblystegium 6, 18, 27, 216

Pseudocampylium 6, 18, 27, 34, 216, 217

Pseudocrossidium 18, 25, 58, 68, 155

Pseudoleskea 75, 76

Pseudoleskeella 18, 28, 74, 75, 80, 82, 219

Pseudoscleropodium 28,77

Pseudotaxiphyllum 27, 48

Pterigynandrum 27, 80, 82

Pterogonium 80

Pterygoneurum 18, 25, 48, 155, 156, 241

Ptilidium 22, 40

Ptilium 29, 82

Ptychostomum 6, 26, 49, 50, 60–62, 64, 65, 73, 190–198

Pulvigera 26, 62, 70, 203, 204, 206

Pylaisia 29,84

Pyramidula 23, 52, 53, 64, 131

Racomitrium 25, 34, 45, 175-177, 180, 184, 185, 243, 248, 250

Radula 17, 22, 41, 109

Reboulia 6, 17, 22, 38, 112-114

Rhabdoweisia 24, 55, 66, 72, 142

Rhizomnium 6, 18, 26, 50, 201

Rhodobryum 26, 50, 64, 190, 191

Rhynchostegiella 18, 28, 79, 222, 223

Rhynchostegium 18, 28, 34, 79, 220, 221

Rhytidiadelphus 7, 19, 29, 81-83, 228

Rhytidium 29,76

Riccardia 22, 39, 109, 110

Riccia 4, 6, 7, 11, 14, 17, 23, 31, 32, 37, 115-121, 239

Ricciocarpos 6, 17, 23, 115

Saelania 5, 17, 25, 55, 66, 143

Sanionia 28, 76, 234

Sarmentypnum 27,77

Scapania 17, 21, 31, 32, 42, 94-97, 235, 244

Schistidium 8, 10, 25, 34, 52, 68, 69, 175, 177-180, 186, 188, 241

Schistochilopsis 5, 6, 16, 21, 89, 91

Sciuro-hypnum 6, 19, 28, 76, 79, 223, 224, 226

Scleropodium 77

Scorpidium 18, 28, 77, 81, 218

Seligeria 8, 25, 52, 53, 56, 72, 174, 175

Sematophyllum 29, 34, 83

Serpoleskea 6, 18, 27, 216

Solenostoma 6, 17, 21, 41, 101-103

Sphaerocarpos 23, 32, 37

Sphagnum 10–12, 14, 17, 23, 34, 44, 122–126, 233, 234, 238, 240, 244–249

Sphenolobus 20, 43

Splachnobryum 25, 34, 65

Straminergon 6, 18, 27, 77, 218

Streblotrichum 12, 18, 25, 58, 71, 169

Syntrichia 6, 10, 11, 18, 25, 49, 57, 61, 68, 156–165

Syzygiella 6, 17, 20, 40, 41, 101

Taxiphyllum 19, 28, 34, 47, 83, 228

Tetraphis 23, 45, 61, 239, 246

Thamnobryum 12, 19, 29, 34, 74, 229

Thuidium 9, 18, 28, 49, 74, 219, 220, 244, 252

Timmia 12, 17, 23, 56, 67, 130

Tomentypnum 27,75

Tortella 10, 11, 18, 25, 57, 66, 67, 171, 172

Tortula 6, 9, 11, 12, 18, 25, 34, 49–51, 54, 57–59, 64, 68–71, 153–168, 232, 239

Trichocolea 21,40

Trichodon 24, 72, 73, 135, 143

Trichostomum 18, 25, 57, 58, 69, 172, 173

Trilophozia 6, 16, 21, 41, 89, 90

Tritomaria 6, 16, 21, 41, 89, 90, 91

Ulota 8, 26, 30, 55, 62, 70, 203-205, 207, 250

Warnstorfia 77

Weissia 18, 25, 34, 51, 54, 57, 69, 167, 173, 237 Zygodon 26, 53, 55, 62, 64, 71, 203